

# **Turnover and Stabilization of Organic Matter in Alpine Soils: Effects of Livestock Grazing and Reforestation**

**Dissertation zur**  
**Erlangung der wissenschaftlichen Doktorwürde**  
**(Dr. sc. nat.)**  
**vorgelegt der**  
**Mathematisch-naturwissenschaftlichen Fakultät**  
**der**  
**Universität Zürich**  
**von**  
David Hiltbrunner  
**aus**  
Luthern LU

## **Promotionskomitee**

Prof. Dr. Michael W.I. Schmidt (Vorsitz)  
Dr. Stephan Zimmermann (Leitung)  
Dr. Frank Hagedorn  
Dr. Pascal A. Niklaus

**Zürich, 2012**

---

## Contents

Summary	I
Zusammenfassung	II
Contents	V
List of Figures (Part A)	VII
List of Tables (Part A)	VII
Abbreviations	VIII

## Part A - Synopsis

<b>1. General Introduction</b>	<b>2</b>
1.1 Land use change as a source or sink for atmospheric CO <sub>2</sub>	2
1.2 Drivers of SOC dynamics as affected by land use (change)	4
1.2.1 C-input	4
1.2.2 Microclimate and physical soil properties	4
1.2.3 Soil microbiology	5
1.3 Non-CO <sub>2</sub> greenhouse gases	6
1.3.1 Methane	7
1.3.2 Nitrous oxide	7
<b>2. Objectives</b>	<b>8</b>
<b>3. Materials and Methods</b>	<b>9</b>
3.1 Study site and sampling design	9
3.2 Soil chemical and microbial analyses and gas fluxes	11
<b>4. Results and discussion</b>	<b>12</b>
4.1 Soil C stocks	12
4.2 Effects of land use (change) on C dynamics	14
4.2.1 Soil organic matter input	14
4.2.2 Microclimate and physical soil properties	16
4.2.3 Soil microorganisms	16
4.3 Methane and nitrous oxide fluxes	19
<b>5. Conclusions</b>	<b>22</b>
<b>6. Research perspectives</b>	<b>23</b>
<b>References</b>	<b>24</b>

## Part B – Publications

<b>Paper I</b>	<b>32</b>
<b>Paper II</b>	<b>42</b>
<b>Paper III</b>	<b>62</b>
 <b>Acknowledgements</b>	 <b>88</b>
<b>Curriculum vitae</b>	<b>90</b>

---

## List of Figures (Part A)

Fig. 1: Net change in forest area between 2000 and 2005.	3
Fig. 2: Study site with transects	9
Fig. 3: Modification of the soil surface by the trampling cattle	10
Fig. 4: Schematic illustration of the different trampling categories	10
Fig. 5: C stocks in the organic layer and the uppermost 25cm of mineral soils	13
Fig. 6: Natural abundance of the $^{13}\text{C}$ isotope in the SOM	15
Fig. 7: Fungi:Bacteria-ratios of the soils	18
Fig. 8: Fluxes of methane and nitrous oxide	21

## List of Tables (Part A)

Table 1 Compilation of grassland afforestation studies in temperate zones	6
Table 2 Overview of the data collection and the soil analyses	16

# **Turnover and Stabilization of Organic Matter in Alpine Soils: Effects of Livestock Grazing and Reforestation**

**Dissertation zur**  
**Erlangung der wissenschaftlichen Doktorwürde**  
**(Dr. sc. nat.)**  
**vorgelegt der**  
**Mathematisch-naturwissenschaftlichen Fakultät**  
**der**  
**Universität Zürich**  
**von**  
David Hiltbrunner  
**aus**  
Luthern LU

## **Promotionskomitee**

Prof. Dr. Michael W.I. Schmidt (Vorsitz)  
Dr. Stephan Zimmermann (Leitung)  
Dr. Frank Hagedorn  
Dr. Pascal A. Niklaus

**Zürich, 2012**





---

## Summary

Land use change (LUC) is - after fossil fuel combustion - the largest source of anthropogenic CO<sub>2</sub> emission. A major part of these emissions originate from tropical deforestation while, by contrast, the abandonment of agricultural land on the Northern hemisphere acts as a net sink for atmospheric CO<sub>2</sub>. Primarily in rural areas, such as alpine regions, socio-economic developments have led to a rapid reforestation of formerly managed grasslands in the recent past. Since mountain soils contain relatively large amounts of labile C, it is likely that afforestation and land management will have a particularly strong impact on the soil C dynamics, but investigations about grassland reforestation in alpine regions are scarce to date. In order to improve the current understanding, I assessed the impacts of (1) livestock grazing and (2) reforestation along a Norway spruce (*Picea abies*) chronosequence on a subalpine pasture in the community of Jaun (FR., Switzerland). The aim of my thesis was to gain a deeper insight into the effects of these two processes on (i) soil C storage, (ii) mechanisms driving the C dynamics, and (iii) greenhouse gas-fluxes.

Centuries of cattle grazing have led to the formation of terraces along the slope. On these cattle tracks, repetitive trampling led to the formation of bare, degraded steps, which covered approximately 10% of the total surface. The features of these bare step soils differed considerably from the ones of the surrounding, vegetated soils. Although step soils were slightly compacted, they primarily evolved by erosion. Soil C storage of the steps was 20% lower than the soil C found under vegetation; estimation on an areal basis, however, concluded that only 2-3% of the total C in the uppermost 25cm was lost through the trampling. Reflecting the physico-chemical properties of the soil, microbial biomass was 27-45% (bacteria and fungi, respectively) lower in the step soils, with the fungal community affected the most by the trampling. CO<sub>2</sub> emissions from the steps were 40% lower as a result of the absent root respiration and the reduced microbial biomass, whilst soil methane (CH<sub>4</sub>) uptake was significantly limited due to inhibited gas diffusion into the compacted step soils. In summary, summer grazing over a number of centuries has strongly affected both soil properties and functions, although trampling effects were confined to a rather small area. The modification of the microtopography

and the altered soil properties, however, are likely to further accelerate soil erosion of the bare soils.

Spruce plantation on the subalpine pasture had relatively moderate effects on the soil C storage. In the first three decades following tree establishment, SOC stocks remained largely unchanged. By contrast, in the 40-45 year old stands, C stocks of the mineral soils transiently decreased; this loss was partly compensated, however, by the additional C in the developing organic layer. In the mature spruce stand (>120 years), mineral soils stored approximately the same amount of C as the ones under pasture ( $\sim 130 \text{ t C ha}^{-1}$ ), but the organic layer in the old forest contained an additional  $20 \text{ t C ha}^{-1}$ . In contrast to the small impact on C storage, C dynamics were noticeably affected by the afforestation. Analysis of the  $^{13}\text{C}$  natural abundance of the SOM revealed a gradual replacement of grass-derived C by spruce-derived C, which was accompanied by a decline in SOC quality (increasing C/N ratio and smaller fraction of labile C). Soil microorganisms in the mineral soils, as analysed by their phospholipids and substrate-induced respiration, were not affected by the vegetation change, either in their total abundance or in their composition (fungal/bacterial-ratio). Fungi, however, were much more abundant in the organic layer, demonstrating their prominent role during litter decomposition. Soil respiration was significantly reduced in the spruce stands with lower soil temperatures; the fact that the decelerated C turnover under forest did not increase SOC storage, however, - at least not in the short term - suggests that it must have been offset by a decreased C input from spruce trees. Soil methane uptake increased by a factor of two to three in the older stands, implying that alpine regions, with their rapidly expanding forest, become an important sink for atmospheric  $\text{CH}_4$ . In summary, spruce plantation had small effects on soil C storage in centennial timescales as compared to the large amounts of C stored in the growing tree biomass. On the other hand, reforestation on alpine grasslands is likely to strongly enhance the soil sink for atmospheric  $\text{CH}_4$ .

## **Zusammenfassung**

Landnutzungsänderungen tragen - nach der Verbrennung fossiler Brennstoffe - am stärksten zum anthropogenen  $\text{CO}_2$ -Ausstoss bei. Der weitaus grösste Teil dieser Emissionen stammt aus der Abholzung tropischer Regenwälder, während die

Brachlegung von Landwirtschaftsland auf der Nordhalbkugel eine Netto-Senke für atmosphärisches CO<sub>2</sub> darstellt. Vor allem in den dünn besiedelten, alpinen Regionen, führten sozio-ökonomische Entwicklungen in den letzten Jahrzehnten zu einer raschen Wiederbewaldung von ehemaligen Weideflächen. Da in alpinen Böden relativ große Mengen an labilem C gespeichert sind, ist es wahrscheinlich, dass die Auswirkungen von Vegetationsänderungen und Beweidung auf den Bodenkohlenstoff hier besonders ausgeprägt sind. Trotzdem gibt es bis anhin nur wenige Studien, welche diese Zusammenhänge genauer erforscht haben. Um die Datengrundlage zu erweitern, habe ich in meiner Dissertation auf einer subalpinen Weide in Jaun (FR) entlang einer Fichten (*Picea abies*) Chronosequenz die Effekte von (1) Beweidung und (2) Wiederbewaldung auf die Boden C-Dynamik untersucht. Das Ziel war, genauere Erkenntnisse über den Einfluss dieser beiden Faktoren auf (i) die Kohlenstoffspeicherung der Böden, (ii) auf die Mechanismen für deren Veränderung und (iii) auf die Treibhausgasflüsse zu gewinnen. Die jahrhundertelange Beweidung führte zur Terrassierung des steilen Hanges. Auf den Terrassen hatten sich durch das regelmässige Trittmuster der Rinder im Laufe der Zeit vegetationsfreie, degenerierte Viehtritte gebildet, die etwa 10% der gesamten Geländeoberfläche bedeckten und sich in ihren Eigenschaften erheblich von den ungestörten Böden unterschieden. Obwohl leicht verdichtet, entstanden die Tritte in erster Linie durch Erosion. Die Boden C Speicherung in den Tritten war 20% geringer als in den ungestörten Böden; auf die Gesamtfläche bezogen wurden aber nur 2-3% des Bodenkohlenstoffes in den obersten 25cm durch die Folgen der Beweidung verloren. Die Analyse der mikrobiellen Phospholipide zeigte eine zwischen 27-45% geringere mikrobielle Biomasse in den Tritten und einen überproportionalen Rückgang der Pilze. Als Ergebnis der kleineren mikrobiellen Abundanz und dem Fehlen von Wurzelatmung waren die CO<sub>2</sub>-Emissionen in den Tritten um 40% niedriger als in den ungestörten Flächen. Zudem war die Methanaufnahme der verdichteten Trittböden aufgrund der geringeren Gasdiffusion signifikant reduziert. Zusammengefasst hat die jahrhundertelange Beweidung zu einer starken Veränderung der Bodeneigenschaften und -Funktionen geführt, obwohl nur ein kleiner Teil der Gesamtfläche davon betroffen war. Es ist aber anzunehmen, dass das entstandene Mikrorelief und die veränderten Bodeneigenschaften in Zukunft die Erosion begünstigen. Die Fichtenpflanzung hatte relativ geringe Auswirkungen auf die Kohlenstoffspeicherung der Weideböden. In den ersten drei Jahrzehnten nach der

Pflanzung blieben die C Vorräte im Mineralboden weitgehend unverändert. In den 40-45 Jahre alten Beständen hingegen hatten die Vorräte im Mineralboden vorübergehend abgenommen, wobei dieser Verlust teilweise durch den zusätzlichen C in der organischen Auflage kompensiert wurde. Im alten Fichtenwald (> 120 Jahre) enthielten die Mineralböden etwa die gleiche Menge an C wie diejenigen unter Weide ( $\sim 130 \text{ t ha}^{-1}$ ), wobei noch zusätzliche  $20 \text{ t C ha}^{-1}$  aus der organischen Auflage dazukommen. Trotz des relativ geringen Einflusses auf die C Speicherung führte die Fichtenpflanzung zu markanten Änderungen der C Dynamik. Die Analyse der natürlichen Abundanz von  $^{13}\text{C}$  im organischen Bodenmaterial offenbarte einen allmählichen Austausch von gras- durch fichtenbürtigen C, welcher mit einem Rückgang der Qualität des organischen Materials (weiteres C/N-Verhältnis und sinkender Anteil an labilem C) einherging. Die Mikroorganismen in den Mineralböden wurden weder in ihrer Gesamtmenge noch in ihrer Zusammensetzung (Pilz/Bakterien-Verhältnis) durch die Aufforstung verändert. In der organischen Auflage dominierten dagegen die Pilze, was ihre zentrale Rolle bei Streuzersetzung unterstreicht. Die Bodenatmung war - als Folge der niedrigeren Temperaturen - in den Baumbeständen geringer als in der Wiese. Dass aber trotz tieferer Atmungsraten keine C Akkumulation in den Waldböden stattgefunden hat, legt den Schluss nahe, dass auch der C-Eintrag im Wald tiefer sein muss als in der Weide. Begünstigt durch die geringeren Bodenwassergehalte war Methan-Aufnahme in den älteren Fichtenbeständen zwei bis drei mal höher als in der Weide. Dies bedeutet, dass alpine Regionen mit ihren schnell zunehmenden Waldflächen in Zukunft wichtige Senken für atmosphärisches  $\text{CH}_4$  darstellen könnten.

## Contents

Summary	I
Zusammenfassung	II
Contents	V
List of Figures (Part A)	VII
List of Tables (Part A)	VII
Abbreviations	VIII

## Part A - Synopsis

<b>1. General Introduction</b>	<b>2</b>
1.1 Land use change as a source or sink for atmospheric CO <sub>2</sub>	2
1.2 Drivers of SOC dynamics as affected by land use (change)	4
1.2.1 C-input	4
1.2.2 Microclimate and physical soil properties	4
1.2.3 Soil microbiology	5
1.3 Non-CO <sub>2</sub> greenhouse gases	6
1.3.1 Methane	7
1.3.2 Nitrous oxide	7
<b>2. Objectives</b>	<b>8</b>
<b>3. Materials and Methods</b>	<b>9</b>
3.1 Study site and sampling design	9
3.2 Soil chemical and microbial analyses and gas fluxes	11
<b>4. Results and discussion</b>	<b>12</b>
4.1 Soil C stocks	12
4.2 Effects of land use (change) on C dynamics	14
4.2.1 Soil organic matter input	14
4.2.2 Microclimate and physical soil properties	16
4.2.3 Soil microorganisms	16
4.3 Methane and nitrous oxide fluxes	19
<b>5. Conclusions</b>	<b>22</b>
<b>6. Research perspectives</b>	<b>23</b>
<b>References</b>	<b>24</b>

## Part B – Publications

<b>Paper I</b>	<b>32</b>
<b>Paper II</b>	<b>42</b>
<b>Paper III</b>	<b>62</b>
 <b>Acknowledgements</b>	 <b>88</b>
<b>Curriculum vitae</b>	<b>90</b>



---

## List of Figures (Part A)

Fig. 1: Net change in forest area between 2000 and 2005.	3
Fig. 2: Study site with transects	9
Fig. 3: Modification of the soil surface by the trampling cattle	10
Fig. 4: Schematic illustration of the different trampling categories	10
Fig. 5: C stocks in the organic layer and the uppermost 25cm of mineral soils	13
Fig. 6: Natural abundance of the $^{13}\text{C}$ isotope in the SOM	15
Fig. 7: Fungi:Bacteria-ratios of the soils	18
Fig. 8: Fluxes of methane and nitrous oxide	21

## List of Tables (Part A)

Table 1 Compilation of grassland afforestation studies in temperate zones	6
Table 2 Overview of the data collection and the soil analyses	16



## Abbreviations

$\delta^{13}\text{C}$ , $\delta^{15}\text{N}$	Stable Isotope Ratio of C and N (delta notation)
a.s.l.	Above Sea Level
C	Carbon
CO <sub>2</sub>	Carbon Dioxide
CH <sub>4</sub>	Methane
DOC	Dissolved Organic Carbon
ECD	Electron Capture Detector
FID	Flame Ionization Detector
GC	Gas Chromatography
GHG	Greenhouse Gas
IRMS	Isotope Ratio Mass Spectrometry
KCl	Potassium Chloride
LUC	Land Use Change
MS	Mass Spectrometry
NH <sub>4</sub> <sup>+</sup>	Ammonium
NO <sub>3</sub> <sup>-</sup>	Nitrate
N <sub>2</sub> O	Nitrous Oxide
OM	Organic Matter
Pg	Petagram (10 <sup>15</sup> g)
PLFA	Phospholipid Fatty Acids
SOC	Soil Organic Carbon
SOM	Soil Organic Matter

## Part A - Synopsis

# 1. General Introduction

## 1.1 Land use change as a source or sink for atmospheric CO<sub>2</sub>

Since the beginning of the industrial revolution, atmospheric CO<sub>2</sub> concentrations have dramatically increased as a result of human activity. In addition to fossil fuel combustion, land use change (LUC) had a major impact on the terrestrial carbon cycle. On a global scale, it is estimated that 156 Pg C were emitted into the atmosphere through changing land use between 1850-1990; approximately half the amount as from fossil fuel burning (Houghton and Goodale, 2004). By far the highest amount of CO<sub>2</sub> released through changing land use (about 60%) originates from tropical deforestation. Abandonment of cultivated areas as a result of socio-economic reasons in the Northern hemisphere, by contrast, resulted in a net uptake of atmospheric CO<sub>2</sub> by the terrestrial biosphere since the 1990s (Houghton, 2003).

Alpine regions in Europe and North America, associated with rapidly expanding forest cover, constitute “hotspots” of LUC with a considerable sink capacity for atmospheric CO<sub>2</sub>. Between 1980-2000, roughly 20% of the agricultural area in the European Alps has been abandoned (MacDonald et al., 2000; Tappeiner et al., 2003) and, for the most part, reforested by natural succession (Tasser et al., 2007); for example, in Switzerland the forested area in the Alps increased by 8% between 1995 and 2006, including an accumulation of tree biomass of 15 million m<sup>3</sup> (Brändli, 2010). However, in contrast to the higher amount of C stored in trees, the effects of afforestation on soil organic carbon (SOC) are much more ambiguous (Guo and Gifford, 2002; Laganriere et al., 2010; Paul et al., 2002; Poeplau et al., 2011; Post and Kwon, 2000). Globally, soils store up to 1500 Pg C in the uppermost meter (Jobbagy and Jackson, 2000) and even small changes in the rates of soil organic carbon (SOC) input and output may have substantial effects on C sequestration and atmospheric CO<sub>2</sub> concentrations (Amundson, 2001). LUC and land management practices are expected to strongly impact C dynamics (Hagedorn et al., 2010), particularly in alpine soils comprising a high fraction of labile C (Budge et al., 2011; Leifeld et al., 2009). As a result, soil C loss would at least partly offset the C sink of the growing trees, as reported by Thuille and Schulze (2006b) in a study investigating spruce afforestation on subalpine pastures. To date, grassland afforestation in temperate zones has been investigated primarily in pine plantations in New Zealand, with only a small number of studies in alpine regions (Table 1).

Land management and intensity is another important driver for the SOC dynamics in alpine soils. Livestock grazing, which is the prevalent form of agriculture in the European Alps, has been shown to considerably affect the quantity and quality of SOC in alpine grasslands (Leifeld and Fuhrer, 2009; Seeber and Seeber, 2005), as well as to increase bulk density, runoff and soil erosion (Nguyen et al., 1998). Greenhouse gas (GHG) fluxes tightly coupled to soil biological processes may, in turn, be substantially altered by the LUC or management practices, with a strong feedback on global warming. Given the rapidly changing land use, rising temperatures, and the relatively high amount of readily decomposable C, alpine ecosystems are being exposed to dramatic environmental changes within a very short time period. Studies with focus on C dynamics in alpine regions are, however, scarce to date (Hagedorn et al., 2010) and our knowledge about the implications of LUC or management practices on soil C storage urgently needs to be improved.

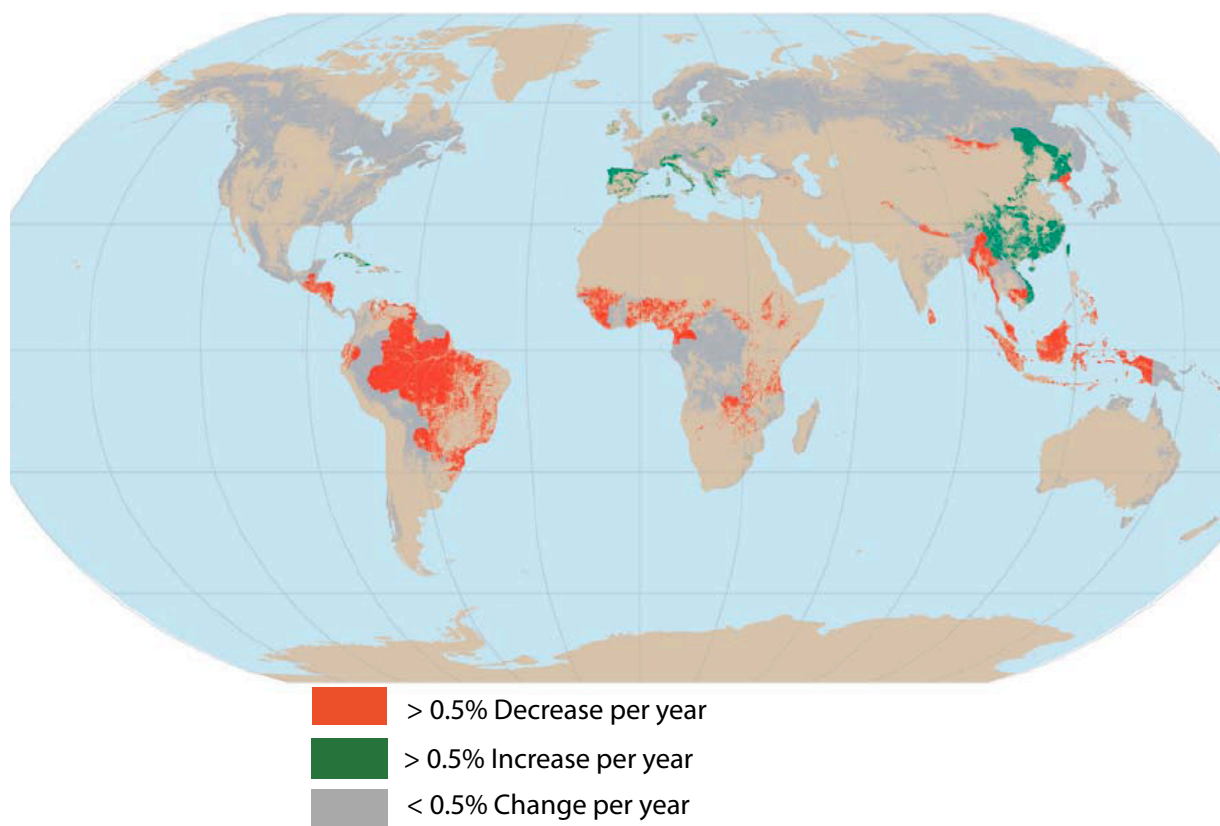


Fig. 1: Net change in forest area between 2000 and 2005. Taken from (Nabuurs et al., 2007)

## **1.2 Drivers of SOC dynamics as affected by land use (change)**

Soil carbon stocks reflect the balance between C input (from root and aboveground biomass and root exudates) and output (respired CO<sub>2</sub> and dissolved organic carbon (DOC)) under steady state conditions. When these fluxes are altered, as by livestock grazing or afforestation, C stocks will adjust until a new equilibrium will be attained. Whether the new C stocks equilibrate at a higher or lower level than prior to the disturbance (i.e. whether C is gained or lost), depends on various factors including (i) quantity and quality of the litter input, (ii) microclimate, (iii), soil physical properties, and (iv) soil microbes.

### **1.2.1 C-input**

C storage and its vertical distribution within the soil column are affected by the amount of litter input and the input pathways (above vs. belowground) (Jackson et al., 1996; Jobbagy and Jackson, 2000), both of which vary strongly between vegetation type. In addition, grass foliage is considered to be more readily decomposable than spruce litter as a result of lower amounts of lignin and higher fractions of N in its composition; this may also lead to changes in quality of the SOC under the respective vegetation type (Guo et al., 2007). Livestock feeding on foliar biomass therefore reduce organic matter (OM) input into soil and can induce shifts in the plant community composition (Yates et al., 2000). Thus, C input is certainly affected by both LUC and grazing in alpine ecosystems, with an expected impact on SOC storage.

### **1.2.2 Microclimate and physical soil properties**

Microclimate exerts a dominant control on SOM mineralization, while soil respiration is positively related to temperature and can be limited by water availability (Yuste et al., 2007). Since microclimatic conditions can be strongly modified by vegetation type (Kellman et al., 2007; Smith and Johnson, 2004), it is likely that afforestation affects soil respiration rates and thus alters the CO<sub>2</sub> fluxes into the atmosphere.

Soil structure plays a central role in the preservation of OM through the physical protection in aggregates (Six et al., 2002), while C mineralization can also be hampered by a limitation of oxygen in compacted soils (Beylich et al., 2010). Since plants have the ability to modify the soil structure through the penetration of roots, the exudation of organic compounds and by their water demand (Angers and Caron, 1998), afforestation may affect soil aggregation. In addition, livestock grazing can lead to severe soil

compaction and enhance soil erosion, particularly on steep alpine slopes (Trimble and Mendel, 1995). On cattle tracks, cattle trampling influences the shape of the soil surface, creating a specific microtopography and leading to increased C mineralization as a result of aggregate disruption (Steffens et al., 2008). Grazing effects on soil physical properties are presumably the major process driving C storage on steep alpine pastures, but very few data are available up until now.

### **1.2.3 Soil microbiology**

Soil microbes maintain their metabolism through the breakdown of organic compounds and are consequently the key drivers of the SOM mineralization. Microbial communities differ among plant species (Berg and Smalla, 2009) and land use (Balser and Firestone, 2005; Macdonald et al., 2009) and can also be modified by agricultural practices (Steenwerth et al., 2002). Plant-microbe interactions play a crucial role in C sequestration (Singh et al., 2004) with fungi-dominated soils purportedly having a higher potential to sequester C (Bailey et al., 2002; Six et al., 2006). Furthermore, it has been shown that the microbial C-use efficiency (the amount of respired CO<sub>2</sub> per unit microbial biomass) increases during succession, implying a more efficient exploitation of the substrate in older ecosystems (Ohtonen et al., 1999). Livestock grazing affects the soil microbes mainly by two different mechanisms: (1) through trampling and (2) through foliar removal. Microbial activity is likely to be limited in compacted soils (Beylich et al., 2010) and microbial biomass has been shown to be negatively correlated with bulk density in a restoration study (McKinley et al., 2005). Herbivory impacts root exudation, nutrient cycling, and plant community composition with substantial consequences for the soil microbes (Bardgett et al., 1998).

Through their fundamental control over biological processes, shifts in the microbial community structure (induced by afforestation or grazing) may have considerable implications on GHG fluxes (Schimel and Gullledge, 1998) and soil C storage (Fontaine and Barot, 2005; Waldrop et al., 2004). However, particular effects of LUC on the soil microbes, with specific reference to C sequestration, are still poorly understood and need further elucidation (De Deyn et al., 2008; Macdonald et al., 2009).

**Table 1** Compilation of studies investigating the effects of grassland afforestation on soil organic carbon in temperate zones. Absolute change in mineral soil C is expressed either as concentration- (mg g<sup>-1</sup>) or stock-change (t ha<sup>-1</sup>) or relative to the stocks under grassland (%). Modified form (Poeplau et al., 2011)

Country	Tree species	Age (year)	Depth (cm)	Change in mineral soil C	Reference
<b>NZ</b>	<b>Conifer</b>	<b>15</b>	<b>10</b>	<b>-49 to +2 mg g<sup>-1</sup></b>	<b>(Alfredsson et al., 1998)</b>
NZ	Pine	10	10	-4 mg g <sup>-1</sup>	(Davis, 1994)
<b>NZ</b>	<b>Various</b>	<b>10-20</b>	<b>10</b>	<b>-4.5 t ha<sup>-1</sup></b>	<b>(Davis and Condron, 2002)</b>
NZ	Pine	13-30	10	-2.7 t ha <sup>-1</sup>	(Giddens et al., 1997)
<b>NZ</b>	<b>Pine</b>	<b>17-19</b>	<b>6</b>	<b>-11.5 mg g<sup>-1</sup></b>	<b>(Groenendijk et al., 2002)</b>
NZ	Pine	20	20	-20 t ha <sup>-1</sup>	(Parfitt et al., 1997)
<b>NZ</b>	<b>Pine</b>	<b>19</b>	<b>20</b>	<b>-30 t ha<sup>-1</sup></b>	<b>(Ross et al., 1999)</b>
NZ	Pine	23-26	10	-5.4 to -23 t ha <sup>-1</sup>	(Scott et al., 1999)
<b>USA</b>	<b>Pine</b>	<b>5-50</b>	<b>38</b>	<b>none</b>	<b>(Scharenbroch et al., 2010)</b>
USA	Deciduous	130	33	+12.8 t ha <sup>-1</sup>	(Martens et al., 2004)
<b>Italy Germany</b>	<b>Spruce</b>	<b>20-110</b>	<b>50</b>	<b>-20%</b>	<b>(Thuille and Schulze, 2006a)</b>
Italy	Spruce	60	10	-20 t ha <sup>-1</sup>	(Thuille et al., 2000)
<b>Italy</b>	<b>Deciduous</b>	<b>75</b>	<b>30</b>	<b>-46 t ha<sup>-1</sup></b>	<b>(Alberti et al., 2008)</b>

### 1.3 Non-CO<sub>2</sub> greenhouse gases

Methane (CH<sub>4</sub>) and nitrous oxide (N<sub>2</sub>O) are the two most important greenhouse gases (GHG) aside from CO<sub>2</sub>, with a combined contribution of 6% (CO<sub>2</sub> contributes to 26%) to the total atmospheric greenhouse effect (Kiehl and Trenberth, 1997). Since pre-industrial times, atmospheric concentrations of CH<sub>4</sub> have more than doubled, but remain constant since the 1990s, while N<sub>2</sub>O concentrations have increased by 16% in the same time period with an ongoing tendency (Denman et al., 2007). Agriculture accounts for 50 and 60% of anthropogenic CH<sub>4</sub> and N<sub>2</sub>O emissions per year, respectively (Smith et al., 2007). Therefore, soils play a major role in the production and uptake of these two GHGs, with land use and land management being an essential part of the anthropogenic impact.

### 1.3.1 Methane

Methane ( $\text{CH}_4$ ) is mainly produced in natural and agricultural waterlogged soils by methanogenic archaea and oxidized in well-aerated upland soils by methanotrophic bacteria. The largest terrestrial sinks for atmospheric  $\text{CH}_4$  are generally found in forest soils and uptake rates often decrease after forest conversion into grassland or arable fields (Hütsch et al., 1994; Smith et al., 2000; Willison et al., 1995). The reduced methanotrophic activity has been attributed to the disturbance of soil physical structure associated with such land-use changes, and to the associated application of mineral nitrogen fertilizers. The disruption of aggregates and compaction of the soil by the trampling livestock might affect the ecological niche of methanotrophs (Boeckx and Van Cleemput, 2001) and limit the diffusive transport of atmospheric  $\text{CH}_4$  into soils (Ball et al., 1997b; Smith et al., 2003). Fertilization of the pasture by cattle faeces might further inhibit  $\text{CH}_4$  oxidation by competitive inhibition of monooxygenase by ammonium (Bodelier and Laanbroek, 2004). As a consequence, it is possible that the rapid forest expansion in the Alps will further enhance soil  $\text{CH}_4$  sink by lowering N fertilization and improving soil structure.

### 1.3.2 Nitrous oxide

In soils,  $\text{N}_2\text{O}$  originates from two different processes involved in the nitrate transformation: nitrification and denitrification. During the oxidation of ammonium to nitrate (nitrification), which is a mainly aerobic process governed by nitrifiers, some N is lost as  $\text{N}_2\text{O}$  to the atmosphere. During denitrification (the reduction of nitrate to  $\text{N}_2$ ),  $\text{N}_2\text{O}$  is a by-product evolving under low redox conditions in presence of labile C (Del Grosso et al., 2000). Denitrification is also assumed to be the major process involved in atmospheric  $\text{N}_2\text{O}$  uptake by soils, mainly under low N and low  $\text{O}_2$  levels (Chapuis-Lardy et al., 2007). Since these processes may all occur in the same soil depending on current environmental conditions,  $\text{N}_2\text{O}$  fluxes are very variable throughout the year and can change within a short period of time. Nevertheless, both vegetation type or land management are likely to have an overruling impact on  $\text{N}_2\text{O}$  fluxes through the modification of soil moisture regime, soil pH and N uptake by the plants (Schaufli et al., 2010). Furthermore, N fertilization by cattle urine is likely to strongly stimulate  $\text{N}_2\text{O}$  fluxes (Luo et al., 2007).



## 2. Objectives

In the European Alps, forest cover is rapidly expanding on abandoned grassland while the remaining land is prevalently used as for livestock grazing. At the same time, alpine soils store a high amount of labile C, which is susceptible to changing environmental conditions. The major aim of my PhD-project was to assess how (1) livestock grazing and (2) afforestation would affect (i) soil C storage, (ii) factors controlling C dynamics, and (iii) greenhouse gas exchange between the soil and the atmosphere.

In particular, the following questions were addressed:

### **Livestock grazing:**

- What are the implications of cattle grazing with respect to microtopography and C dynamics on steep alpine pastures?
- How does cattle trampling affect soil microbes?
- What are the implications on the CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes?

### **Afforestation:**

- Does afforestation with Norway spruce (as a simulation of natural occurring tree encroachment) increase or decrease the soil C sink in alpine ecosystems?
- How does the tree establishment affect the factors influencing soil organic matter decomposition?
- Does afforestation affect CO<sub>2</sub>, CH<sub>4</sub> or N<sub>2</sub>O fluxes and what are the mechanisms involved?

### 3. Materials and Methods

#### 3.1 Study site and sampling design

The study site was selected in order to represent a typical (sub)-alpine ecosystem with spruce encroachment after pasture abandonment. To have more detailed information about land use history and tree age, however, we chose a series of different aged spruce afforestations instead of a successional forest. The spruce chronosequence was established on a south-facing pastoral slope extending from 1450 m a.s.l to 1800 m a.s.l. in the community of Jaun, Canton of Fribourg, Switzerland (7°15'54 E; 46°37'17 N). Mean summer and winter air temperatures are 11.4°C and 0.6°C, respectively; mean annual precipitation averages 1250 mm with a maximum in summer. Soils are *Cambisols* on calcareous bedrock.

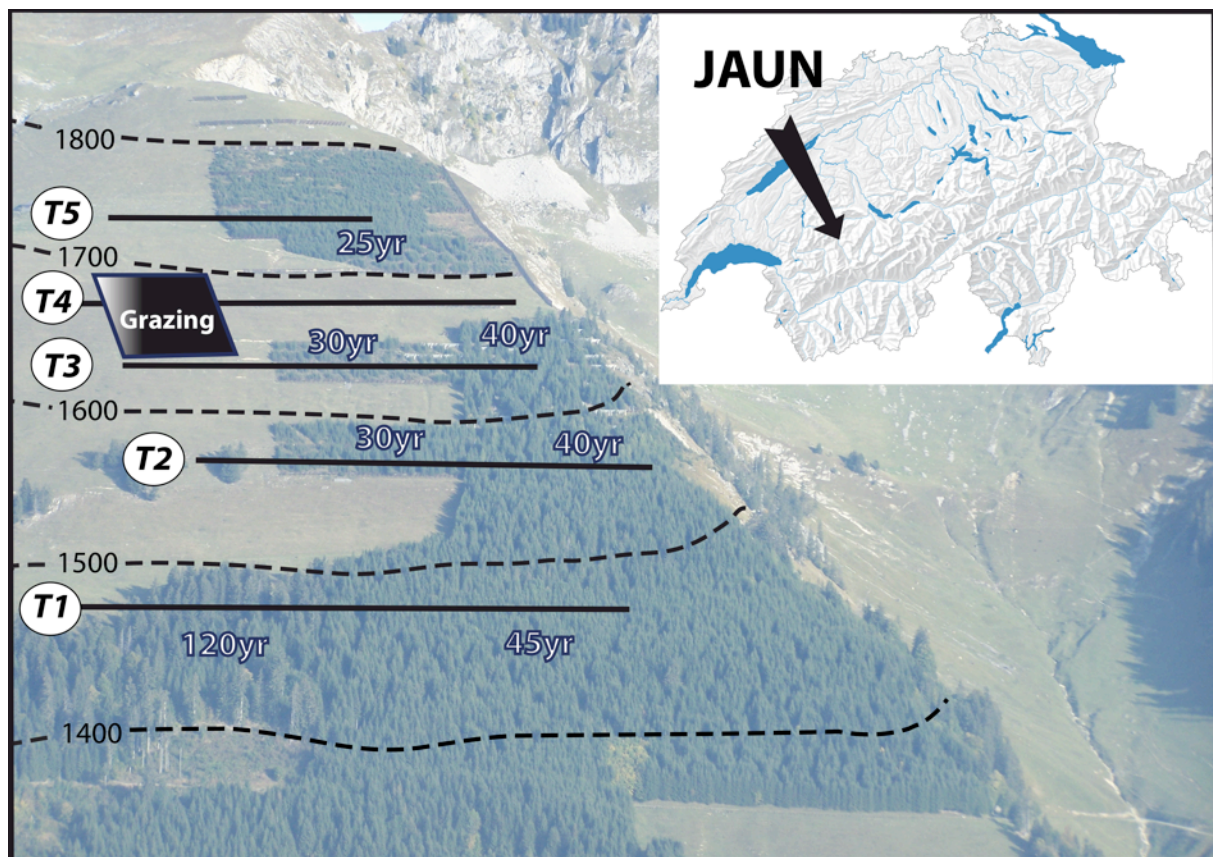


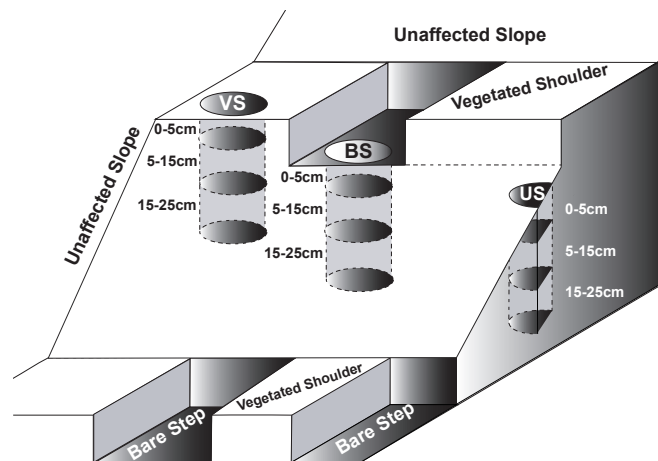
Fig. 2: Study site with transects (T1-T5), along which soils have been sampled. T4 consists of pasture soils only and was used to test the horizontal homogeneity of the slope. The black rhombus displays the pastoral area, where the effects of livestock grazing were assessed

The eastern part of the slope was gradually afforested with Norway spruce (*Picea abies* L.) after severe avalanches in 1956, while the western part remained under pasture. The chronosequence includes stands with an age of 25, 30, 40, and 45 years and a mature spruce forest (older than 120 years (Fig. 2).

Soils were sampled along four transects at different altitudes, ranging from the pasture into the forest stands. Within each differing section of the transect (determined by land use or stand age), five to ten soil pits were dug and sampled at six different depths (0-5, 5-10, 10-20, 20-40, 40-60 and 60-80cm). A fifth transect with 20 soil pits only within the pasture was used to test the homogeneity of the slope (Fig. 1) In addition, the organic layer was sampled quantitatively in all forest stands. The effects of livestock trampling were assessed in the pasture within an area of two hectares. According to the microtopography (bare steps, vegetated shoulders, unaffected slope), three adjacent soil cores (0-25cm) were taken in 20 replicates. The relative proportion of the respective trampling class on the total surface was estimated by aerial photo.



Fig. 3: Modification of the soil surface by the trampling cattle. Terraces (left) and bare steps (right) formed by the repetitive trampling.



**Fig. 4:** Schematic illustration of the soil sampling within the different trampling categories: BS='Bare Step', VS ='Vegetated Shoulder', US ='Unaffected Slope'.

### 3.2 Soil chemical and microbial analyses and gas fluxes

**Table 2** Overview of the collected data and the soil analyses according to the different manuscripts of my thesis. More details about the methodological aspects can be found in the respective papers.

Data/Analyses	Methods/Equipment	References
<b>Paper I (Livestock grazing)</b>		
<i>Soil C and N concentrations</i>	Elemental analyser coupled to IRMS	(Hiltbrunner et al., 2012)
$\delta^{13}\text{C}$ , $\delta^{15}\text{N}$ of SOM		
<i>Soil porosity</i>	Pyknometer mehod	(Blake and Hartge, 1986)
<i>Microbial Phospholipids (PLFA)</i>	Lipid extraction/GC-MS	(Zelles, 1997)
<i>Fungal ergosterol</i>	Ergosterol extraction/HPLC	(Daood et al., 2008)
<i>Soil respiration in situ</i>	Static chambers/GC-FID	(Rochette et al., 1992)
<b>Paper II (Effects of afforestation on GHG-fluxes)</b>		
<i>Fluxes of <math>\text{CH}_4</math> and <math>\text{N}_2\text{O}</math></i>	Static chambers /GC-FID/ECD	(Hartmann et al., 2011)
<i>Soil <math>\text{NH}_4^+</math> and <math>\text{NO}_3^-</math></i>	KCl extraction/ Colorimetrically	(Navone, 1964)
<i>Potential nitrification</i>	Shaken slurry method	(Hart et al., 1994)
<i>Denitrification enzyme activity</i>	acetylene inhibition assay	(Patra et al., 2005)
<i>Soil porosity</i>	Pyknometer mehod	(Blake and Hartge, 1986)
<i>Volumetric soil water content</i>	TDR-probes	
<b>Paper III (Effects of afforestation on C storage)</b>		
<i>C and N conc. of SOM and plants</i>	Elemental analyser coupled to IRMS	(Hiltbrunner et al., 2012)
$\delta^{13}\text{C}$ , $\delta^{15}\text{N}$ of SOM and plants		
<i>Labile C fraction</i>	Soil incubation	
<i>Lignin content of plant material</i>	Wet chemical extraction	(Dence, 1992)
<i>Stone content and root biomass</i>	Wet sieving	
<i>Microbial biomass</i>	Substrate induced respiration	(Cheng and Virginia, 1993)
<i>Soil respiration in situ</i>	Gas sampling/GC-FID	(Rochette et al., 1992)
<i>Soil moisture and temperature</i>	Decagon loggers	
<i>Tree biomass</i>	Allometric functions	(Kaufmann, 2001)
<b>Others (livestock grazing)</b>		
<i>Fluxes of <math>\text{CH}_4</math> and <math>\text{N}_2\text{O}</math></i>	Static chambers /GC-FID/ECD	(Hartmann et al., 2011)
<i>Extractable <math>\text{NH}_4^+</math> and <math>\text{NO}_3^-</math></i>	KCl extraction/Colorimetrically	(Navone, 1964)
<i>Potential nitrification</i>	Shaken slurry method	(Hart et al., 1994)
<i>Denitrification enzyme activity</i>	Acetylene inhibition assay	(Patra et al., 2005)
<b>Others (afforestation)</b>		
<i>Microbial Phospholipids (PLFA)</i>	Lipid extraction/GC-MS	(Zelles, 1997)
<i>Fungal ergosterol</i>	Ergosterol extraction/HPLC	(Daood et al., 2008)
<i>Neutral sugars of SOM and plants</i>	Acid hydrolysis	(Eder et al., 2010)

## 4. Results and discussion

In the following chapter, I will discuss the impact of (1) cattle grazing and (2) spruce afforestation on (i) soil C storage and (ii) the factors driving soil C dynamics as well as on (iii) fluxes of methane and nitrous oxide. Where appropriate, I will compare the relevance of the two processes on C dynamics and greenhouse gas fluxes in this alpine ecosystem.

### 4.1 Soil C stocks

Livestock grazing on this steep alpine slope strongly modified the soil surface of the pastures and thus affected numerous soil properties as shown in paper I. On the cattle tracks, a regular pattern of bare steps and intermediate vegetated shoulders was formed by the repetitive trampling. In between the tracks, the slope was mostly unaffected by the trampling and thus served as a control (Fig. 3-4). Reflecting the microtopography, the C storage was significantly affected by the trampling; in the uppermost 25 cm of the bare step soils approximately  $6 \text{ kg C m}^{-2}$  was stored, while in the vegetated areas the stocks amounted to  $7.5 \text{ kg C m}^{-2}$ . Accordingly, the loss of SOC by trampling accounted for about  $1.5 \text{ kg m}^{-2}$  in the bare steps, corresponding to 20% of the total stock in this layer. At the same time, roughly 10% of the study site was classified as bare step soils. Thus, calculated on an areal basis,  $0.15 \text{ kg m}^{-2}$  or 2% of the total carbon stocks (0–25 cm) were lost through the cattle trampling. This loss is rather small compared to grazing effects in a series of temperate and subtropical South American grasslands, where approximately 10% of the C stocks in the uppermost 30 cm were lost (Pineiro et al., 2009) or in a semi arid Mongolian grassland, where the loss by heavy grazing accounted for 35% of the total stocks in the 0–4cm layer (Steffens et al., 2008).

As discussed in paper III, afforestation with Norway spruce had only transient effects on the C sink of the mineral soils. Approximately  $3 \text{ kg C m}^{-2}$  was lost 40 - 45 years after tree establishment, although this loss was partly compensated by the C accumulating in the organic layer. C pools of the mineral soils under pasture were similar to those found in the old forest ( $\sim 13 \text{ kg m}^{-2}$ ). If the organic layer was included, however, soils in the old forest stored  $\sim 2 \text{ kg m}^{-2}$  more C than the ones under pasture. Our data, therefore, do not support the findings of Thuille and Schulze (2006b), who reported consistently higher C stocks in the mineral soils under sub-alpine meadows than under spruce forests. In

contrast, a recent metadata analysis compiling land use change effects in the temperate zone showed a clear increase of the soil C storage roughly 100 years after grassland afforestation if the organic layer is taken into account, but negligible effects on the mineral soil (Poeplau et al., 2011). Apart from effects of climate, soil texture, tree species, and land use history (Guo and Gifford, 2002; Laganier et al., 2010), different sampling depths may have contributed to the inconsistent results among the studies (Harrison et al., 2011). In any case, SOC-pools on a mass basis were only marginally affected by the land use change as compared to C stored in the tree biomass, which amounted  $\sim 40 \text{ kg m}^{-2}$  in the old forest.

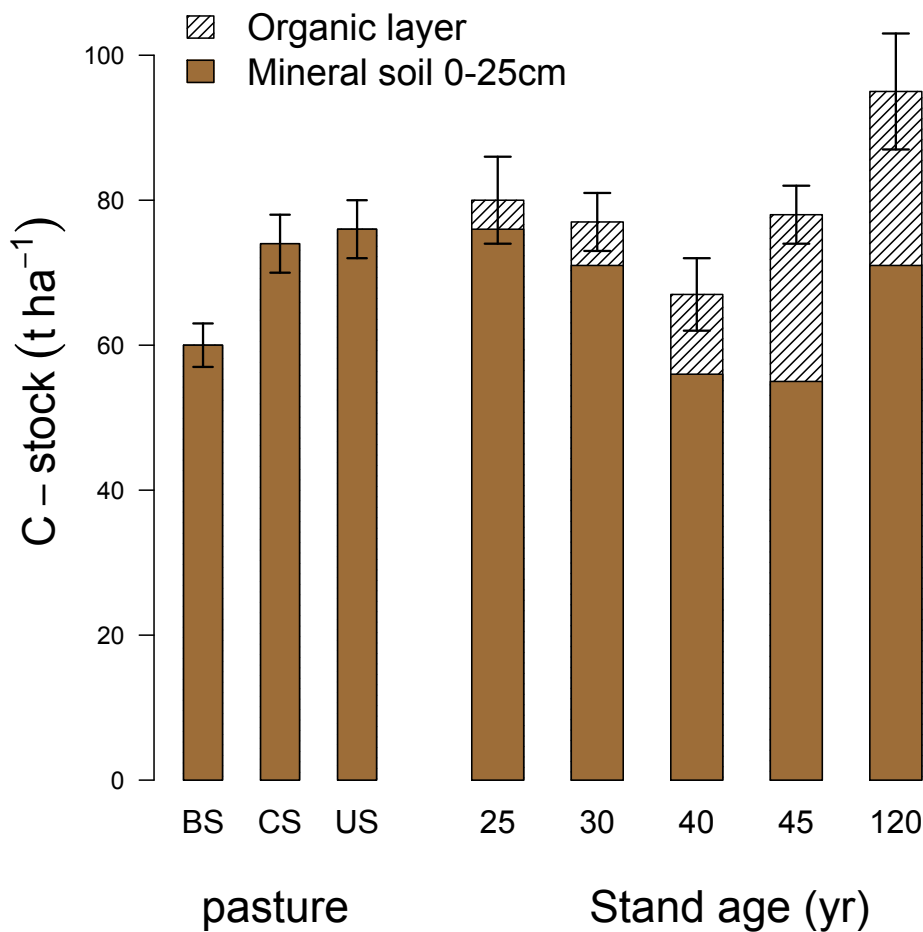


Fig. 5: C stocks in the organic layer and the uppermost 25cm of mineral soils under pasture as affected by the trampling (BS= Bare steps, VS= Vegetated shoulders, US= Unaffected slope) and in the different aged spruce stands.

Comparing the impacts of livestock and afforestation on soil C pools indicates that cattle tracks were moderately depleted in soil C, while spruce establishment slightly increased

the soil C storage (Fig.5). In the mature forest most of the additional C was stored in the organic layer; this makes it vulnerable to decomposition if the environmental conditions are altered after wind throw or bark beetle infestation, for example. The more stabilized C in the mineral soil is, by contrast, much less affected by the land use conversion when trees can grow for over one decade. This important finding has to be considered when choosing adequate rotation lengths and management practices in alpine forest ecosystems. Trampling effects on SOC stocks are comparatively small when calculated on an areal basis since the degraded steps cover only ~10% of the total surface. However, it is reasonable that the steps will further be eroded since they form a perfect channel system for the surface water flow. With this in mind, the C loss is likely to continue, further enhancing the degradation of the slope. In contrast to the SOC dynamics following afforestation with recovering soil C stocks after one decade, cattle grazing on this steep alpine pasture has clear deleterious effects on soil C in the long term.

## **4.2 Effects of land use (change) on C dynamics**

### **4.2.1 Soil organic matter input**

Plant functional traits control the production, composition and input pathways of litter into the soil and thereby have a large impact on the C sequestration (De Deyn et al., 2008). If vegetation is lacking, as on the degraded cattle tracks, soil C supply is dramatically reduced, which leads to decreased C storage in these soils. In addition, bare soils are prone to erosion since they are not protected against the eroding power of raindrops and because soil aggregation is lower in the absence of roots (Angers and Caron, 1998).

If vegetation is changing, C dynamics are likely to be affected by the altered litter input. Paper III demonstrated that after tree plantation  $\delta^{13}\text{C}$  values increased from -27‰ in the uppermost mineral soils under pasture to -25.5 ‰ in the old forest (Fig. 7). The gradual shift in  $^{13}\text{C}$  natural abundance can be interpreted as a clear indication of a replacement of pasture by tree-derived C in SOM with time after afforestation and reflects the altered input pathways and litter chemistry. In grasslands, the soil C is predominantly root derived, while in forests, roots and above ground litter more equally contribute to the C input (Jobbagy and Jackson, 2000). As a result, fine root biomass was



almost 40% lower in the forest than in the pasture soils and needle fall led to the development of an organic layer under the spruce trees. The insufficient incorporation of the needle C into the mineral soil in conjunction with the lower root production may partly explain the C loss from mineral soils as shown by Guo et al. (2007). Furthermore, grass roots and foliage had smaller fractions of lignin (23 and 18%, respectively) and narrower C/N ratios (26 and 49%, respectively) than the spruce roots and needles (lignin fractions of 31 and 30% and C/N ratios of 40 and 70%, respectively). As a result of the lower spruce litter quality, the fraction of labile C in the mineral soil decreased after forest establishment and the C/N ratios increased. This shows that forest establishment lowered the quality of the SOC, which may enhance its stabilization in the long term (De Deyn et al., 2008).

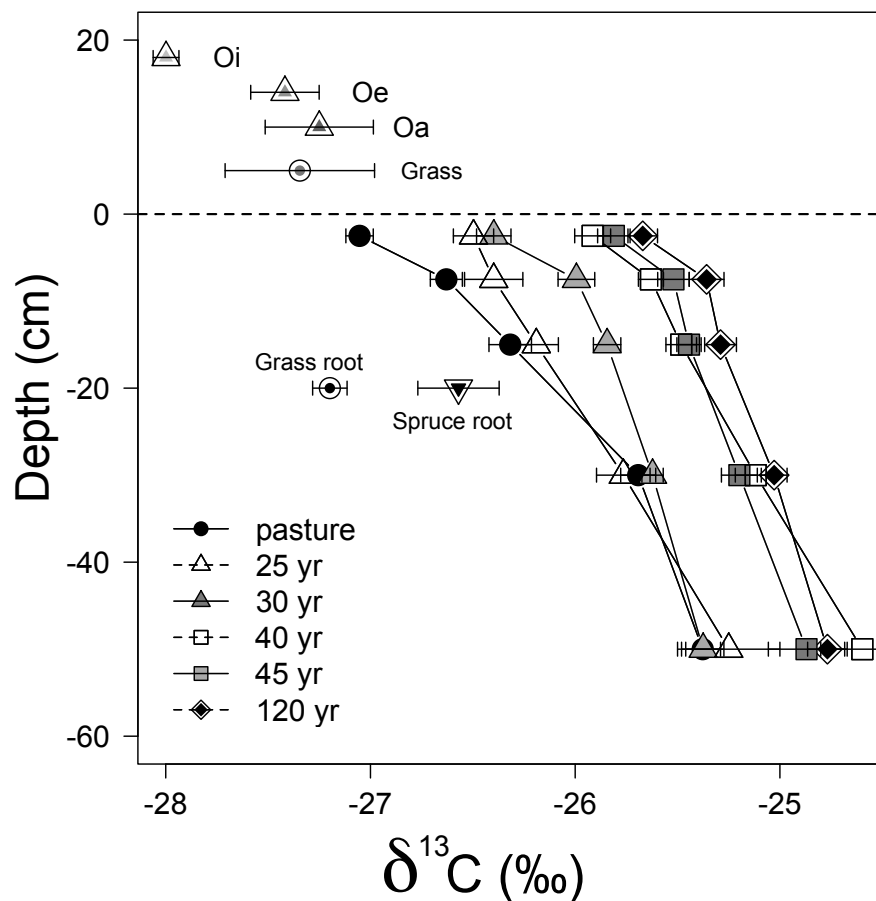


Fig.6: Natural abundance of the  $^{13}\text{C}$  isotope in the organic layers (Oi, Oe, Oa) and the mineral soils (0-50cm) as affected by vegetation and stand age



#### **4.2.2 Microclimate and physical soil properties**

Soil respiration, defined by the combined root and microbial respiration, is positively correlated to soil temperature and can be inhibited by very high and very low soil moisture (Smith et al., 2003); consequently, microclimate is a key factor controlling C mineralization. As discussed in paper III, average soil temperature during the growing season in the spruce stands was about 5°C below the one under pasture, which resulted in significantly lower respiration rates in the forest. But even after correcting for the temperature effect in the statistical analysis, significantly more CO<sub>2</sub> was respired from the pasture than from the forest soils. It is thus likely that accelerated C cycling through the denser grass root system also contributed to the higher soil respiration under pasture. Soil moisture regime, in contrast, had little impact on the total respiration rates *in situ*. In field moist organic layer material, however, C mineralization was strongly suppressed compared to the moistened samples. This suggests that water availability indeed limits litter decomposition on these south-facing slopes during dry summer periods.

While microclimate was altered by the afforestation but not by livestock grazing, the grazing did, however, have strong effects on the soil physical properties as discussed in paper I. Soils of the bare steps had significantly higher bulk densities and lower porosities, although the step soils surprisingly showed little evidence of compaction (bulk density < 1.1 kg dm<sup>-3</sup>); as a result, no deleterious implications on microbial driven processes should be expected under these conditions (Beylich et al., 2010). Gas diffusion, by contrast, may be reduced, since the gas transport through smaller pore volume will be hampered under wet conditions. Further trampling may negatively impact the stability of the bare soils, because aggregate formation is very likely limited by a lack of organic matter (as indicated by the lower C concentrations and the higher aggregate density) (Chaney and Swift, 1984). Afforestation had no consistent effect on bulk density, although the lowest density values were measured in the old forest. Overall, the effects on the physical soil properties were concentrated on a relatively small area, while the major part of the study site was unaffected by the land use (change).

#### **4.2.3 Soil microorganisms**

Soil microorganisms play a crucial role in a large number of processes, such as organic matter turnover, nutrient acquisition, N cycling, C sequestration and aggregate formation (van der Heijden et al., 2008). Changing environmental conditions due to

vegetation replacement or mechanical disturbance by cattle trampling strongly impacts microbial communities (De Deyn et al., 2008; Potthoff et al., 2006; Six et al., 2006) with potential implications on the microbial driven processes. As discussed in paper I, livestock grazing generally reduced the abundance of all microbial groups in the bare steps, resulting in a 40% decrease in soil respiration than in the vegetated soils. The concentrations of bacterial and fungal fatty acids were reduced by 28 and 45%, respectively; as a result, the fungi to bacteria-ratio decreased in the degraded steps (Fig. 8). The most likely mechanisms for this effect, and on the fungi in particular, are the lack of root biomass (mycorrhiza) and litter input (saprophytic fungi) in conjunction with the disruption of the fungal mycelia by the trampling. Since the pronounced decrease of the fungal biomass presumably has a negative impact on aggregate formation (Bossuyt et al., 2001; Rillig et al., 2002), it is likely to further accelerate soil erosion.

The replacement of vegetation associated with land use change is known to modify soil microbial communities (Lauber et al., 2008; Macdonald et al., 2009) as plant species govern the structure and function of the soil microbial communities in their vicinity (Berg and Smalla, 2009). Generally, fungi are the dominant community in forest soils, with large amounts of complex organic substances (e.g. lignin), whilst bacteria are prevailing in fertilized arable soils, metabolizing simpler compounds (de Boer et al., 2005). Fungal and bacterial food webs have different nutrient turnover rates (van der Heijden et al., 2008) and fungal dominated ecosystems may have a higher C sequestration potential (Bailey et al., 2002), although this theory is still under debate (Strickland and Rousk, 2010). In this study, afforestation had surprisingly small effects on the soil microbial abundance and community composition. Neither substrate-induced respiration nor total PLFA in the uppermost mineral soil (0-5cm) significantly differed between vegetation types, whilst fungal:bacterial ratios were even found to decrease slightly in the older spruce stands (Fig. 7). This is in contrast to both our expectations and the majority of other studies which show fungal abundance is highest in forest soils (Bailey et al., 2002; Djukic et al., 2010; Lauber et al., 2008; Macdonald et al., 2009), although the opposite trend has also been reported (Waldrop and Firestone, 2006). How can the small microbial response to the afforestation be explained? Firstly, soil pH and C:N ratios, two well known determinants for the microbial community structure, were only moderately affected by the vegetation type (pH = 5.1 and 4.4; C/N= 11 and 14; for pasture and old forest soils, respectively). Secondly, land use history plays an important

role; it is likely that the effects on the soil microbes would have been more pronounced after forest plantation on an arable field with fertilizer application and tilling. Thirdly, mycorrhizal abundance might be higher in the denser grass root system than in the forest soils, which harbour more saprophytic fungi; a potential substitution of these two fungal groups is not detectable with the PLFA technique that utilises only one fungal biomarker for both ectomycorrhiza and saprophytes. Finally, in contrast to other studies (Djukic et al., 2010), we analysed mineral soils and organic layers separately; while vegetation had little impact on the soil microbes in mineral soil, fungi were much more abundant in the organic layer (Fig. 8), reflecting the high stratification of the fungal communities (Baldrian et al., 2012; Joergensen and Wichern, 2008). Consequently, sampling procedure must be taken into account when comparing between different studies.

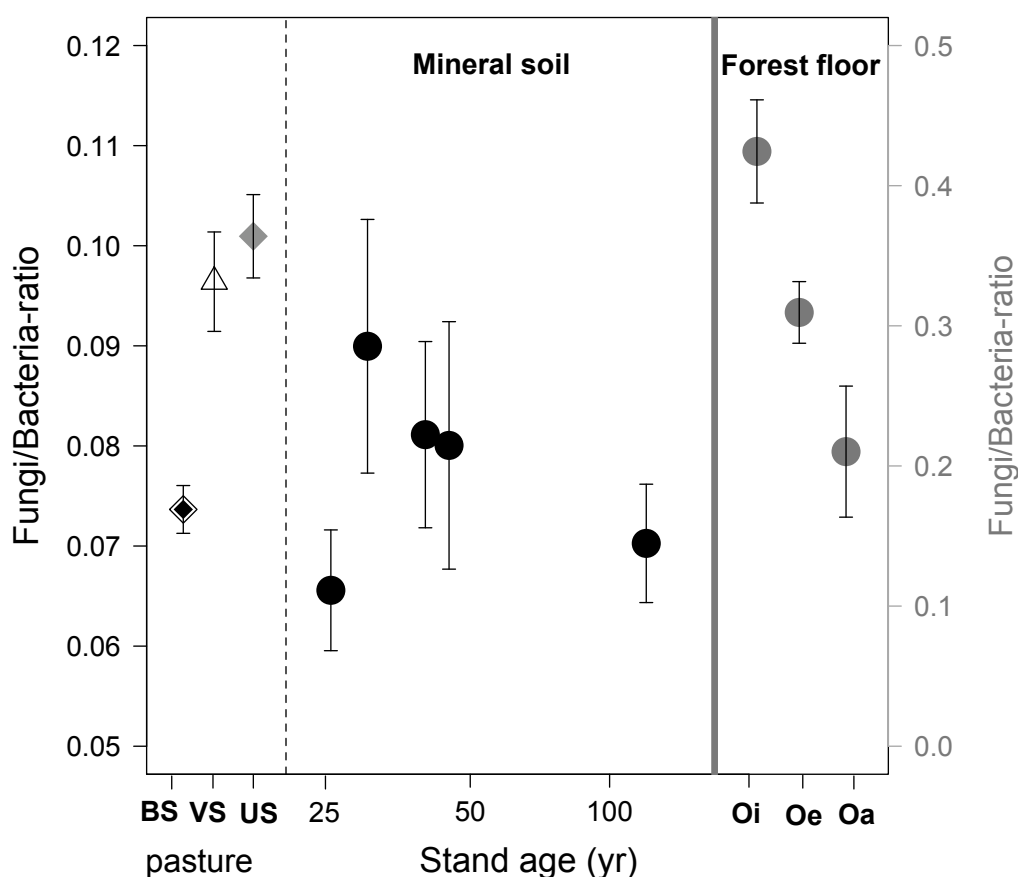


Fig.7: Fungi:Bacteria-ratios in the mineral soils (0-5cm) under pasture as affected by the trampling (BS= Bare steps, VS= Vegetated shoulders, US= Unaffected slope) (left), in the different aged spruce stands (centre) and in the organic layer (right). Note the different scale for the organic layer samples.

### 4.3 Methane and nitrous oxide fluxes

Upland soils are the major biological sink for atmospheric CH<sub>4</sub>, although they may turn into a source for CH<sub>4</sub> under wet conditions (Le Mer and Roger, 2001). In this study, soil CH<sub>4</sub> uptake was the dominant process irrespective of time, land use or trampling effects. There were a few single measurements, however, that showed CH<sub>4</sub> emissions from soils, indicating that methane production (methanogenesis) exceeds CH<sub>4</sub> oxidation (methanotrophy) under extreme conditions. In the bare steps formed by the trampling, CH<sub>4</sub> uptake was more than 50% reduced in comparison to the vegetated areas (Fig. 6); this is presumably the result of (i) limited diffusion of CH<sub>4</sub> into the compacted step soils and (ii) lower abundance of the methanotrophic bacteria due to the trampling. Methane uptake is strongly controlled by soil porosity as gas diffusion, and thus the supply of CH<sub>4</sub> for the methanotrophic bacteria, can be hampered in compacted soils (Ball et al., 1997a); in addition, water filled pore space (WFPS) is higher in compacted soils, which additionally limits gas diffusion. This well-known relationship between soil structure, soil moisture and CH<sub>4</sub> oxidation (Ball et al., 1997a; Smith et al., 2003) was also important in this study. Bare step soils had significantly smaller pore volumes and higher water filled pore spaces ( $P < 0.001$ ), which is the major reason for the smaller CH<sub>4</sub> uptake (or even positive fluxes in a few measurements). Furthermore, microbial biomass was significantly smaller in the degraded bare soils than under vegetation. Although we did not assess the methanotrophic bacteria specifically, it is likely that this relatively sensitive bacterial group was particularly negatively affected by the trampling, which additionally has diminished soil sink capacity.

As with CH<sub>4</sub> fluxes, nitrous oxide emissions were significantly affected by the trampling, with higher emissions from the degraded steps (Fig. 6). The effects of the trampling on N<sub>2</sub>O were less evident, however, which may be explained by the wider range of processes involved in the N<sub>2</sub>O production. Nitrous oxide emissions are generally closely related to the mineral N status of the soils and emerge as a by-product of nitrification and denitrification (Davidson et al., 2000; Morkved et al., 2007; Pihlatie et al., 2004). Since nitrification is an aerobic process and denitrification primarily occurs when O<sub>2</sub> is limiting (Schimel and Gullledge, 1998), the contribution of each process on the N<sub>2</sub>O production changes with environmental conditions, but this may not necessarily be reflected in the net fluxes. Nitrous oxide emissions in this study were weakly affected by the soil mineral N status, which also did not consistently differ between the trampling

categories. Thus, the general impact of livestock on soil N<sub>2</sub>O emissions turned out to be rather small.

Spruce afforestation on the extensively grazed subalpine pasture increased the soil CH<sub>4</sub> sink by a factor of two to three, as discussed in paper II (Fig.6). Soil structure was not affected by the tree establishment, but soil moisture gradually decreased with stand age, which is presumably the key mechanism for the enhanced CH<sub>4</sub> sink following afforestation. Other factors such as soil acidification or increasing NH<sub>4</sub><sup>+</sup> concentrations, which have been shown inhibit CH<sub>4</sub> uptake (King and Schnell, 1994; Reay et al., 2005; Weslien et al., 2009), were of minor relevance in our study. Surprisingly, not even additional urea fertilization (20g N m<sup>-2</sup>) significantly reduced CH<sub>4</sub> uptake, although strong effects on the N dynamics were observed; N<sub>2</sub>O emissions from the fertilized soils, for instance, were significantly increased and N status was improved. Land use conversion, by comparison, had little effects on the N<sub>2</sub>O fluxes; this is in contrast to the results of Christiansen and Gundersen (2011), who measured stimulated N<sub>2</sub>O emissions with increasing age in a series of spruce and oak afforestations. However, considering the high temporal and spatial variability of N<sub>2</sub>O fluxes, more measurements would be needed to appropriately quantify LUC effects on nitrous oxide emissions.

Overall, the impact of grazing and afforestation on the methane uptake were opposing; tree plantation enhanced and cattle trampling decreased the soil sink. In the old forest, CH<sub>4</sub> uptake was more than six times higher than in the bare steps. Given that the forest area expanded 900 km<sup>-2</sup> in the last two decades (Brändli, 2010), and assuming a difference of 1-1.5 μmol CH<sub>4</sub> m<sup>2</sup> h<sup>-1</sup> between pasture and old forest stands during the growing season, land use change increased the soil sink by ~50-70 t CH<sub>4</sub>-C yr<sup>-1</sup> within this time period. When the stimulating effects of cattle trampling and fertilization on methanogenic archaea (Radl et al., 2007) and the CH<sub>4</sub> production from ruminant digestion are taken into account, the positive effect of reforestation on the CH<sub>4</sub> sink in alpine ecosystems is further amplified. On the other hand, the vegetation had no effect on N<sub>2</sub>O emissions, which may be partly explained by the relatively low number of measurements. Since fertilization by the cattle strongly increases N<sub>2</sub>O production, however, the abandonment of the pastures certainly reduces N<sub>2</sub>O emissions by the cessation of the fertilization.

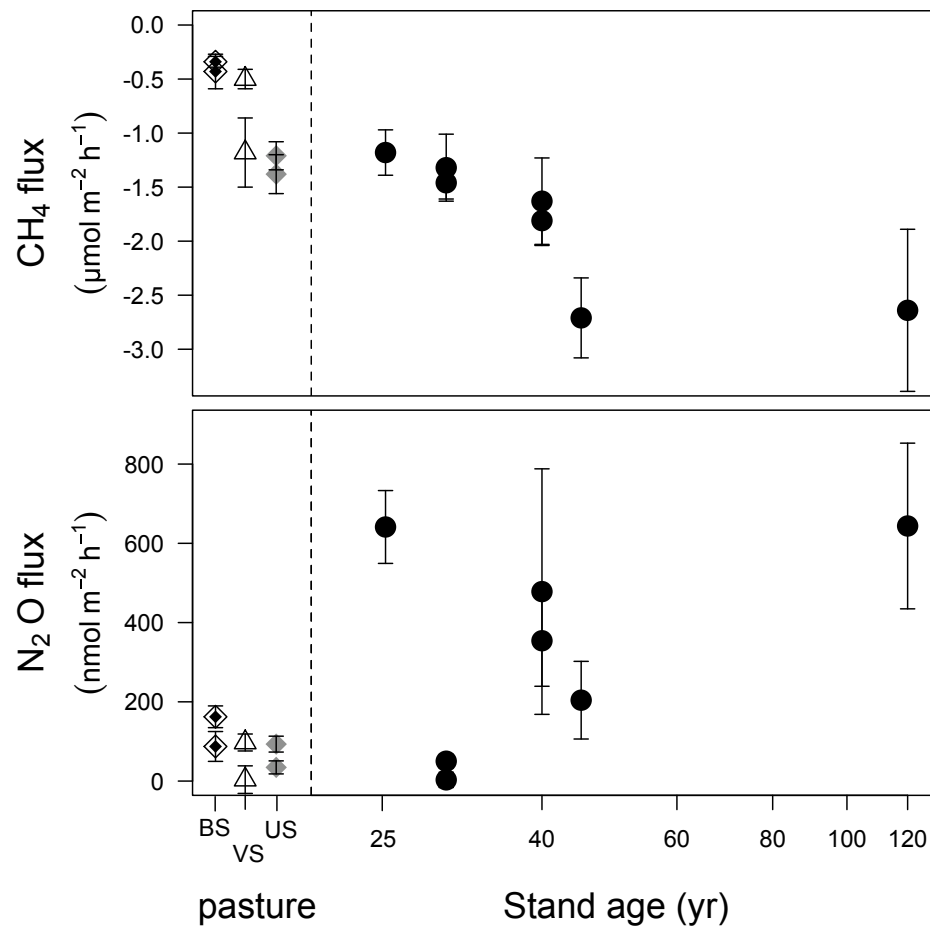


Fig. 8: Fluxes of methane (top) and nitrous oxide (bottom) from pasture soils as affected by the trampling (BS= Bare steps, VS= Vegetated shoulders, US= Unaffected slope) (left) and from soils of the different aged spruce stands (right).

## 5. Conclusions

**Cattle grazing led to the development of bare steps formed by repetitive trampling, which covered 10% of the total surface area.** C stocks in the step soils were 20% lower than in the surrounding vegetated areas. On an areal basis, however, trampling induced C loss amounted to only 2-3% of the total stock in the uppermost 25cm.

**Bare step soils had significantly lower microbial biomass, reflecting the lower C concentrations in the steps.** Fungi in particular were negatively affected by the trampling, which potentially has negative effects on soil aggregation and thus, might further accelerate soil erosion.

**Spruce afforestation increased soil C sink by ~20 t ha<sup>-1</sup> primarily through the accumulation of additional C in the organic layer.** C stocks in the mineral soil transiently decreased in the 40-45 year old spruce stands and increased thereafter to the levels prior to the land use change (~130 t ha<sup>-1</sup>). In comparison to the C stored in tree biomass (~400 t ha<sup>-1</sup> in the >120 years old forest), however, changes in the soil C pools were negligible.

**Grass-derived C in the mineral soils was gradually replaced by spruce derived C, as shown by the positive shift in <sup>13</sup>C natural abundance of the SOM with increasing stand age.** This replacement was accompanied by a decline in SOM quality (increasing C:N ratio and smaller fraction of labile C) as a result of the lower quality (higher C:N ratio and higher lignin concentrations) of spruce than grass litter.

**Soil methane sink increased by a factor between two to three after spruce afforestation as a result of enhanced gas diffusion in the dryer soils of the older spruce stands.** Soil respiration was higher in the pasture than in the forests, with soil temperature as driving factor.

**Soil microbial abundance and community composition was only moderately affected by the vegetation change.** In the mineral soils, the fungal/bacterial ratio (as analysed by PLFA) remained largely unaffected after tree plantation (0.07-0.1). By contrast, in the developing organic layer, fungal fraction in the microbial biomass increased by a factor of between two to four as compared to mineral soil (0.2 – 0.4).

## 6. Research perspectives

During my PhD-project, I investigated a wide spectrum of processes involved in the soil C dynamics and GHG-fluxes as affected by land use (change). As a result, numerous new questions arose, from which I will address the most important three in the following section. **Soil C storage** transiently decreased in the 40 and 45 year old stands, primarily due to C loss from the subsoils. This is in contrast to the general assumption that the effects of LUC are most pronounced in the surface soils. To date, however, the majority of the LUC studies only include topsoil layers, risking an exclusion of stock changes in the deeper soils, where large amounts of relatively stable C are stored. Therefore, investigating subsoil C loss in more detail, and the possible mechanisms involved (priming by the deeper tree roots, enhanced aeration of the soils by the higher water demand of the trees, effects on DOC fluxes), would help to obtain a more holistic understanding about the implications of afforestation on SOC dynamics within the entire soil. Natural  $^{13}\text{C}$  abundance of the SOM in the bulk soils showed a gradual **replacement of grass-derived C by spruce-derived C**. This is remarkable, since only a few studies (e.g. (Gruenzweig et al., 2007)) applied  $\delta^{13}\text{C}$  values to assess effects of land use change on SOC dynamics in a pure C3 plant system, as the one in Jaun. To obtain a more detailed picture, however, bulk soils should further be separated into different fractions by density fractionation. In conjunction with  $^{14}\text{C}$  analysis, C turnover dynamics, and even turnover- times, could be determined more appropriately. **Soil methane uptake** strongly increased after afforestation, with decreasing soil moisture in the older spruce stands as underlying mechanism. In addition, we tested the impact of other well-known drivers of the  $\text{CH}_4$  oxidation, such as soil N status, pH, temperature and soil porosity. Effects on methane oxidizing bacteria, however, have not been analysed thus far, although methanotrophic communities have been shown to be altered by land use conversion (Singh et al., 2007). Therefore, additional investigation of the methanotrophs, either by PLFA-SIP or molecular analysis, would be a promising approach to further elucidate the processes involved in soil  $\text{CH}_4$  uptake.



## References

- Alberti, G., Peressotti, A., Piussi, P., Zerbi, G., 2008. Forest ecosystem carbon accumulation during a secondary succession in the Eastern Prealps of Italy. *Forestry* 81(1), 1-11.
- Alfredsson, H., Condron, L.M., Clarholm, M., Davis, M.R., 1998. Changes in soil acidity and organic matter following the establishment of conifers on former grassland in New Zealand. *Forest Ecology and Management* 112(3), 245-252.
- Amundson, R., 2001. The carbon budget in soils. *Annual Review of Earth and Planetary Sciences* 29, 535-562.
- Angers, D.A., Caron, J., 1998. Plant-induced changes in soil structure: Processes and feedbacks. *Biogeochemistry* 42(1-2), 55-72.
- Bailey, V.L., Smith, J.L., Bolton, H., 2002. Fungal-to-bacterial ratios in soils investigated for enhanced C sequestration. *Soil Biology & Biochemistry* 34(7), 997-1007.
- Baldrian, P., Kolarik, M., Stursova, M., Kopecky, J., Valaskova, V., Vetrovsky, T., Zifcakova, L., Snajdr, J., Ridl, J., Vlcek, C., Voriskova, J., 2012. Active and total microbial communities in forest soil are largely different and highly stratified during decomposition. *Isme Journal* 6(2), 248-258.
- Ball, B.C., Dobbie, K.E., Parker, J.P., Smith, K.A., 1997a. The influence of gas transport and porosity on methane oxidation in soils. *Journal of Geophysical Research-Atmospheres* 102(D19), 23301-23308.
- Ball, B.C., Smith, K.A., Klemetsson, L., Brumme, R., Sitaula, B.K., Hansen, S., Prieme, A., MacDonald, J., Horgan, G.W., 1997b. The influence of soil gas transport properties on methane oxidation in a selection of northern European soils. *Journal of Geophysical Research-Atmospheres* 102(D19), 23309-23317.
- Balser, T.C., Firestone, M.K., 2005. Linking microbial community composition and soil processes in a California annual grassland and mixed-conifer forest. *Biogeochemistry* 73(2), 395-415.
- Bardgett, R.D., Wardle, D.A., Yeates, G.W., 1998. Linking above-ground and below-ground interactions: How plant responses to foliar herbivory influence soil organisms. *Soil Biology & Biochemistry* 30(14), 1867-1878.
- Berg, G., Smalla, K., 2009. Plant species and soil type cooperatively shape the structure and function of microbial communities in the rhizosphere. *Fems Microbiology Ecology* 68(1), 1-13.
- Beylich, A., Oberholzer, H.R., Schrader, S., Hoper, H., Wilke, B.M., 2010. Evaluation of soil compaction effects on soil biota and soil biological processes in soils. *Soil & Tillage Research* 109(2), 133-143.
- Blake, G.R., Hartge, K.H., 1986. Particle Density. *Methods of Soil Analysis, Part I*, In: Klute, A. (ed.): *Physical and Mineralogical Methods*, Amer. Soc. Agron. and Soil Sci. Soc. Amer., Madison (Wisconsin), USA, 363-375.
- Bodelier, P.L.E., Laanbroek, H.J., 2004. Nitrogen as a regulatory factor of methane oxidation in soils and sediments. *Fems Microbiology Ecology* 47(3), 265-277.
- Boeckx, P., Van Cleemput, O., 2001. Estimates of N<sub>2</sub>O and CH<sub>4</sub> fluxes from agricultural lands in various regions in Europe. *Nutrient Cycling in Agroecosystems* 60(1-3), 35-47.
- Bossuyt, H., Denef, K., Six, J., Frey, S.D., Merckx, R., Paustian, K., 2001. Influence of microbial populations and residue quality on aggregate stability. *Applied Soil Ecology* 16(3), 195-208.

- Brändli, U.-B., 2010. Schweizerisches Landesforstinventar. Ergebnisse der dritten Erhebung 2004–2006. Birmensdorf, Eidgenössische Forschungsanstalt für Wald, Schnee und Landschaft WSL. Bern, Bundesamt für Umwelt, BAFU. 312 S.
- Budge, K., Leifeld, J., Hiltbrunner, E., Fuhrer, J., 2011. Alpine grassland soils contain large proportion of labile carbon but indicate long turnover times. *Biogeosciences* 8(7), 1911-1923.
- Chaney, K., Swift, R.S., 1984. The influence of organic-matter on aggregate stability in some british soils. *Journal of Soil Science* 35(2), 223-230.
- Chapuis-Lardy, L., Wrage, N., Metay, A., Chotte, J.L., Bernoux, M., 2007. Soils, a sink for N<sub>2</sub>O? A review. *Global Change Biology* 13(1), 1-17.
- Cheng, W.X., Virginia, R.A., 1993. Measurement of microbial biomass in arctic tundra soils using fumigation extraction and substrate-induced respiration procedures. *Soil Biology & Biochemistry* 25(1), 135-141.
- Christiansen, J.R., Gundersen, P., 2011. Stand age and tree species affect N<sub>2</sub>O and CH<sub>4</sub> exchange from afforested soils. *Biogeosciences* 8(9), 2535-2546.
- Daood, H.G., Korbasz, M., Hamdan, S., Beczner, J., 2008. Simultaneous LC Determination of Ergosterol, Tocopherols and Carotenoids in Foods. *Chromatographia* 68, S137-S140.
- Davidson, E.A., Keller, M., Erickson, H.E., Verchot, L.V., Veldkamp, E., 2000. Testing a conceptual model of soil emissions of nitrous and nitric oxides. *Bioscience* 50(8), 667-680.
- Davis, M.R., 1994. Topsoil properties under tussock grassland and adjoining pine forest in otago, new-zealand. *New Zealand Journal of Agricultural Research* 37(4), 465-469.
- Davis, M.R., Condon, L.M., 2002. Impact of grassland afforestation on soil carbon in New Zealand: a review of paired-site studies. *Australian Journal of Soil Research* 40(4), 675-690.
- de Boer, W., Folman, L.B., Summerbell, R.C., Boddy, L., 2005. Living in a fungal world: impact of fungi on soil bacterial niche development. *Fems Microbiology Reviews* 29(4), 795-811.
- De Deyn, G.B., Cornelissen, J.H.C., Bardgett, R.D., 2008. Plant functional traits and soil carbon sequestration in contrasting biomes. *Ecology Letters* 11(5), 516-531.
- Del Grosso, S.J., Parton, W.J., Mosier, A.R., Ojima, D.S., Kulmala, A.E., Phongpan, S., 2000. General model for N<sub>2</sub>O and N<sub>2</sub> gas emissions from soils due to denitrification. *Global Biogeochemical Cycles* 14(4), 1045-1060.
- Dence, C.W., 1992. The determination of lignin. In: Lin, S.Y. & Dence, C.W. (Eds.) *Methods in lignin chemistry*. Springer-Verlag, Berlin Heidelberg. p. 33–61.
- Denman, K.L., Brasseur, G., Chidthaisong, A., Ciais, P., Cox, P.M., Dickinson, R.E., Hauglustaine, D., Heinze, C., Holland, E., Jacob, D., Lohmann, U., Ramachandran, S., da Silva Dias, P.L., S.C., W., Zhang, X., 2007. Couplings Between Changes in the Climate System and Biogeochemistry. In: *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* [Solomon, S., D. Qin, M. Manning, Z. Chen, M. Marquis, K.B. Averyt, M.Tignor and H.L. Miller (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Djukic, I., Zehetner, F., Mentler, A., Gerzabek, M.H., 2010. Microbial community composition and activity in different Alpine vegetation zones. *Soil Biology & Biochemistry* 42(2), 155-161.
- Eder, E., Spielvogel, S., Koelbl, A., Albert, G., Koegel-Knabner, I., 2010. Analysis of hydrolysable neutral sugars in mineral soils: Improvement of alditol acetylation

- for gas chromatographic separation and measurement. *Organic Geochemistry* 41(6), 580-585.
- Fontaine, S., Barot, S., 2005. Size and functional diversity of microbe populations control plant persistence and long-term soil carbon accumulation. *Ecology Letters* 8(10), 1075-1087.
- Giddens, K.M., Parfitt, R.L., Percival, H.J., 1997. Comparison of some soil properties under *Pinus radiata* and improved pasture. *New Zealand Journal of Agricultural Research* 40(3), 409-416.
- Groenendijk, F.M., Condon, L.M., Rijkse, W.C., 2002. Effects of afforestation on organic carbon, nitrogen and sulfur concentrations in New Zealand hill country soils. *Geoderma* 108(1-2), 91-100.
- Gruenzweig, J.M., Gelfand, I., Fried, Y., Yakir, D., 2007. Biogeochemical factors contributing to enhanced carbon storage following afforestation of a semi-arid shrubland. *Biogeosciences* 4(5), 891-904.
- Guo, L.B., Gifford, R.M., 2002. Soil carbon stocks and land use change: a meta analysis. *Global Change Biology* 8(4), 345-360.
- Guo, L.B., Wang, M., Gifford, R.M., 2007. The change of soil carbon stocks and fine root dynamics after land use change from a native pasture to a pine plantation. *Plant and Soil* 299(1-2), 251-262.
- Hagedorn, F., Mulder, J., Jandl, R., 2010. Mountain soils under a changing climate and land-use. *Biogeochemistry* 97(1), 1-5.
- Harrison, R.B., Footen, P.W., Strahm, B.D., 2011. Deep Soil Horizons: Contribution and Importance to Soil Carbon Pools and in Assessing Whole-Ecosystem Response to Management and Global Change. *Forest Science* 57(1), 67-76.
- Hart, S.C., Stark, J.M., Davidson, E.A., Firestone, M.K., 1994. Nitrogen mineralisation, immobilisation and nitrification. In: Weaver, R. W. et al., (Ed.), *Methods of Soil Analysis, Part 2. Microbiological and Biochemical Properties-SSSA, Vol. Part 2.*, Madison, WI, USA.
- Hartmann, A.A., Buchmann, N., Niklaus, P.A., 2011. A study of soil methane sink regulation in two grasslands exposed to drought and N fertilization. *Plant and Soil* 342(1-2), 265-275.
- Hiltbrunner, D., Schulze, S., Hagedorn, F., Schmidt, M.W.I., Zimmermann, S., 2012. Cattle trampling alters soil properties and changes soil microbial communities in a Swiss sub-alpine pasture. *Geoderma* 170, 369-377.
- Houghton, R.A., 2003. Revised estimates of the annual net flux of carbon to the atmosphere from changes in land use and land management 1850-2000. *Tellus Series B-Chemical and Physical Meteorology* 55(2), 378-390.
- Houghton, R.A., Goodale, C.L., 2004. Effects of land-use change on the carbon balance of terrestrial ecosystems. In: R.S.A.G.P.H.R.A. DeFries (Ed.), *Ecosystems and Land Use Change. Geophysical Monograph Series*, pp. 85-98.
- Hütsch, B.W., Webster, C.P., Powlson, D.S., 1994. Methane oxidation in soil as affected by land-use, soil-ph and n-fertilization. *Soil Biology & Biochemistry* 26(12), 1613-1622.
- Jackson, R.B., Canadell, J., Ehleringer, J.R., Mooney, H.A., Sala, O.E., Schulze, E.D., 1996. A global analysis of root distributions for terrestrial biomes. *Oecologia* 108(3), 389-411.
- Jobbagy, E.G., Jackson, R.B., 2000. The vertical distribution of soil organic carbon and its relation to climate and vegetation. *Ecological Applications* 10(2), 423-436.
- Joergensen, R.G., Wichern, F., 2008. Quantitative assessment of the fungal contribution to microbial tissue in soil. *Soil Biology & Biochemistry* 40(12), 2977-2991.

- Kaufmann, E., 2001. Estimation of standing timber, growth and cut. In: P. Brassel and H. Lischke (Ed.), *Swiss National Forest Inventory: Methods and Models of the Second Assessment*. Swiss Federal Research Institute WSL, Birmensdorf, S. 162-196.
- Kellman, L., Beltrami, H., Risk, D., 2007. Changes in seasonal soil respiration with pasture conversion to forest in Atlantic Canada. *Biogeochemistry* 82(1), 101-109.
- Kiehl, J.T., Trenberth, K.E., 1997. Earth's annual global mean energy budget. *Bulletin of the American Meteorological Society* 78(2), 197-208.
- King, G.M., Schnell, S., 1994. Ammonium and nitrite inhibition of methane oxidation by *Methylobacter albus* bg8 and *Methylosinus trichosporium* ob3b at low methane concentrations. *Applied and Environmental Microbiology* 60(10), 3508-3513.
- Laganiere, J., Angers, D.A., Pare, D., 2010. Carbon accumulation in agricultural soils after afforestation: a meta-analysis. *Global Change Biology* 16(1), 439-453.
- Lauber, C.L., Strickland, M.S., Bradford, M.A., Fierer, N., 2008. The influence of soil properties on the structure of bacterial and fungal communities across land-use types. *Soil Biology & Biochemistry* 40(9), 2407-2415.
- Le Mer, J., Roger, P., 2001. Production, oxidation, emission and consumption of methane by soils: A review. *European Journal of Soil Biology* 37(1), 25-50.
- Leifeld, J., Fuhrer, J., 2009. Long-term management effects on soil organic matter in two cold, high-elevation grasslands: clues from fractionation and radiocarbon dating. *European Journal of Soil Science* 60(2), 230-239.
- Leifeld, J., Zimmermann, M., Fuhrer, J., Conen, F., 2009. Storage and turnover of carbon in grassland soils along an elevation gradient in the Swiss Alps. *Global Change Biology* 15(3), 668-679.
- Luo, J., Ledgard, S.F., Lindsey, S.B., 2007. Nitrous oxide emissions from application of urea on New Zealand pasture. *New Zealand Journal of Agricultural Research* 50(1), 1-11.
- Macdonald, C.A., Thomas, N., Robinson, L., Tate, K.R., Ross, D.J., Dando, J., Singh, B.K., 2009. Physiological, biochemical and molecular responses of the soil microbial community after afforestation of pastures with *Pinus radiata*. *Soil Biology & Biochemistry* 41(8), 1642-1651.
- MacDonald, D., Crabtree, J.R., Wiesinger, G., Dax, T., Stamou, N., Fleury, P., Lazpita, J.G., Gibon, A., 2000. Agricultural abandonment in mountain areas of Europe: Environmental consequences and policy response. *Journal of Environmental Management* 59(1), 47-69.
- Martens, D.A., Reedy, T.E., Lewis, D.T., 2004. Soil organic carbon content and composition of 130-year crop, pasture and forest land-use managements. *Global Change Biology* 10(1), 65-78.
- McKinley, V.L., Peacock, A.D., White, D.C., 2005. Microbial community PLFA and PHB responses to ecosystem restoration in tallgrass prairie soils. *Soil Biology & Biochemistry* 37(10), 1946-1958.
- Morkved, P.T., Dorsch, P., Bakken, L.R., 2007. The N<sub>2</sub>O product ratio of nitrification and its dependence on long-term changes in soil pH. *Soil Biology & Biochemistry* 39(8), 2048-2057.
- Nabuurs, G.J., Masera, O., Andrasko, K., Benitez-Ponce, P., Boer, R., Dutschke, M., Elsiddig, E., Ford-Robertson, J., Frumhoff, P., Karjalainen, T., Krankina, O., Kurz, W.A., Matsumoto, M., Oyhantcabal, W., Ravindranath, N.H., Sanz Sanchez, M.J., Zhang, X., 2007. Forestry. In *Climate Change 2007: Mitigation. Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on*

- Climate Change [B. Metz, O.R. Davidson, P.R. Bosch, R. Dave, L.A. Meyer (eds)], Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Navone, R., 1964. Proposed method for nitrate in potable waters. *J. Am. Wat. Works Ass.* 56, 781-783.
- Nguyen, M.L., Sheath, G.W., Smith, C.M., Cooper, A.B., 1998. Impact of cattle treading on hill land 2. Soil physical properties and contaminant runoff. *New Zealand Journal of Agricultural Research* 41(2), 279-290.
- Ohtonen, R., Fritze, H., Pennanen, T., Jumpponen, A., Trappe, J., 1999. Ecosystem properties and microbial community changes in primary succession on a glacier forefront. *Oecologia* 119(2), 239-246.
- Parfitt, R.L., Percival, H.J., Dahlgren, R.A., Hill, L.F., 1997. Soil and solution chemistry under pasture and radiata pine in New Zealand. *Plant and Soil* 191(2), 279-290.
- Patra, A.K., Abbadie, L., Clays-Josserand, A., Degrange, V., Grayston, S.J., Loiseau, P., Louault, F., Mahmood, S., Nazaret, S., Philippot, L., Poly, E., Prosser, J.I., Richaume, A., Le Roux, X., 2005. Effects of grazing on microbial functional groups involved in soil N dynamics. *Ecological Monographs* 75(1), 65-80.
- Paul, K.I., Polglase, P.J., Nyakuengama, J.G., Khanna, P.K., 2002. Change in soil carbon following afforestation. *Forest Ecology and Management* 168(1-3), 241-257.
- Pihlatie, M., Syvasalo, E., Simojoki, A., Esala, M., Regina, K., 2004. Contribution of nitrification and denitrification to N<sub>2</sub>O production in peat, clay and loamy sand soils under different soil moisture conditions. *Nutrient Cycling in Agroecosystems* 70(2), 135-141.
- Pineiro, G., Paruelo, J.M., Jobbagy, E.G., Jackson, R.B., Oesterheld, M., 2009. Grazing effects on belowground C and N stocks along a network of cattle exclosures in temperate and subtropical grasslands of South America. *Glob. Biogeochem. Cycle* 23.
- Poeplau, C., Don, A., Vesterdal, L., Leifeld, J., Van Wesemael, B., Schumacher, J., Gensior, A., 2011. Temporal dynamics of soil organic carbon after land-use change in the temperate zone - carbon response functions as a model approach. *Global Change Biology* 17(7), 2415-2427.
- Post, W.M., Kwon, K.C., 2000. Soil carbon sequestration and land-use change: processes and potential. *Global Change Biology* 6(3), 317-327.
- Potthoff, M., Steenwerth, K.L., Jackson, L.E., Drenovsky, R.E., Scow, K.M., Joergensen, R.G., 2006. Soil microbial community composition as affected by restoration practices in California grassland. *Soil Biology & Biochemistry* 38(7), 1851-1860.
- Radl, V., Gattinger, A., Chronakova, A., Nemcova, A., Cuhel, J., Simek, M., Munch, J.C., Schlöter, M., Elhottova, D., 2007. Effects of cattle husbandry on abundance and activity of methanogenic archaea in upland soils. *Isme Journal* 1, 443-452.
- Reay, D.S., Nedwell, D.B., McNamara, N., Ineson, P., 2005. Effect of tree species on methane and ammonium oxidation capacity in forest soils. *Soil Biology & Biochemistry* 37(4), 719-730.
- Rillig, M.C., Wright, S.F., Eviner, V.T., 2002. The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: comparing effects of five plant species. *Plant and Soil* 238(2), 325-333.
- Rochette, P., Gregorich, E.G., Desjardins, R.L., 1992. Comparison of static and dynamic closed chambers for measurement of soil respiration under field conditions. *Canadian Journal of Soil Science* 72(4), 605-609.
- Ross, D.J., Tate, K.R., Scott, N.A., Feltham, C.W., 1999. Land-use change: effects on soil carbon, nitrogen and phosphorus pools and fluxes in three adjacent ecosystems. *Soil Biology and Biochemistry* 31(6), 803-813.

- Scharenbroch, B.C., Flores-Mangual, M.L., Lepore, B., Bockheim, J.G., Lowery, B., 2010. Tree Encroachment Impacts Carbon Dynamics in a Sand Prairie in Wisconsin. *Soil Sci Soc Am J* 74(3), 956-968.
- Schaufler, G., Kitzler, B., Schindlbacher, A., Skiba, U., Sutton, M.A., Zechmeister-Boltenstern, S., 2010. Greenhouse gas emissions from European soils under different land use: effects of soil moisture and temperature. *European Journal of Soil Science* 61(5), 683-696.
- Schimel, J.P., Gullledge, J., 1998. Microbial community structure and global trace gases. *Global Change Biology* 4(7), 745-758.
- Scott, N.A., Tate, K.R., Ford-Robertson, J., Giltrap, D.J., Smith, C.T., 1999. Soil carbon storage in plantation forests and pastures: land-use change implications. *Tellus B* 51(2), 326-335.
- Seeber, J., Seeber, G.U.H., 2005. Effects of land-use changes on humus forms on alpine pastureland (Central Alps, Tyrol). *Geoderma* 124(3-4), 215-222.
- Singh, B.K., Millard, P., Whiteley, A.S., Murrell, J.C., 2004. Unravelling rhizosphere-microbial interactions: opportunities and limitations. *Trends in Microbiology* 12(8), 386-393.
- Singh, B.K., Tate, K.R., Kolipaka, G., Hedley, C.B., Macdonald, C.A., Millard, P., Murrell, J.C., 2007. Effect of afforestation and reforestation of pastures on the activity and population dynamics of methanotrophic bacteria. *Applied and Environmental Microbiology* 73(16), 5153-5161.
- Six, J., Feller, C., Denef, K., Ogle, S.M., Sa, J.C.D., Albrecht, A., 2002. Soil organic matter, biota and aggregation in temperate and tropical soils - Effects of no-tillage. *Agronomie* 22(7-8), 755-775.
- Six, J., Frey, S.D., Thiet, R.K., Batten, K.M., 2006. Bacterial and fungal contributions to carbon sequestration in agroecosystems. *Soil Science Society of America Journal* 70(2), 555-569.
- Smith, D.L., Johnson, L., 2004. Vegetation-mediated changes in microclimate reduce soil respiration as woodlands expand into grasslands. *Ecology* 85(12), 3348-3361.
- Smith, K.A., Ball, T., Conen, F., Dobbie, K.E., Massheder, J., Rey, A., 2003. Exchange of greenhouse gases between soil and atmosphere: interactions of soil physical factors and biological processes. *European Journal of Soil Science* 54(4), 779-791.
- Smith, K.A., Dobbie, K.E., Ball, B.C., Bakken, L.R., Sitaula, B.K., Hansen, S., Brumme, R., Borken, W., Christensen, S., Prieme, A., Fowler, D., Macdonald, J.A., Skiba, U., Klemmedtsson, L., Kasimir-Klemmedtsson, A., Degorska, A., Orlanski, P., 2000. Oxidation of atmospheric methane in Northern European soils, comparison with other ecosystems, and uncertainties in the global terrestrial sink. *Global Change Biology* 6(7), 791-803.
- Smith, P., Martino, D., Cai, Z., Gwary, D., Janzen, H., Kumar, P., McCarl, B., Ogle, S., O'Mara, F., Rice, C., Scholes, B., Sirotenko, O., 2007. Agriculture. In *Climate Change 2007: Mitigation. Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* [B. Metz, O.R. Davidson, P.R. Bosch, R. Dave, L.A. Meyer (eds)], Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Steenwerth, K.L., Jackson, L.E., Calderon, F.J., Stromberg, M.R., Scow, K.M., 2002. Soil microbial community composition and land use history in cultivated and grassland ecosystems of coastal California. *Soil Biology & Biochemistry* 34(11), 1599-1611.

- Steffens, M., Kolbl, A., Totsche, K.U., Kogel-Knabner, I., 2008. Grazing effects on soil chemical and physical properties in a semiarid steppe of Inner Mongolia (PR China). *Geoderma* 143(1-2), 63-72.
- Strickland, M.S., Rousk, J., 2010. Considering fungal:bacterial dominance in soils - Methods, controls, and ecosystem implications. *Soil Biology & Biochemistry* 42(9), 1385-1395.
- Tappeiner, U., Tappeiner, G., Hilbert, A., Mattanovich, E., 2003. The EU Agricultural Policy and the Environment. Evaluation of the Alpine Region. Blackwell, Berlin, Germany.
- Tasser, E., Walde, J., Tappeiner, U., Teutsch, A., Nogglar, W., 2007. Land-use changes and natural reforestation in the Eastern Central Alps. *Agriculture Ecosystems & Environment* 118(1-4), 115-129.
- Thuille, A., Buchmann, N., Schulze, E.-D., 2000. Carbon stocks and soil respiration rates during deforestation, grassland use and subsequent Norway spruce afforestation in the Southern Alps, Italy. *Tree Physiol* 20(13), 849-857.
- Thuille, A., Schulze, E.-D., 2006a. Carbon dynamics in successional and afforested spruce stands in Thuringia and the Alps. *Glob. Change Biol.* 12(2), 325-342.
- Thuille, A., Schulze, E.D., 2006b. Carbon dynamics in successional and afforested spruce stands in Thuringia and the Alps. *Global Change Biology* 12(2), 325-342.
- Trimble, S.W., Mendel, A.C., 1995. The cow as a geomorphic agent — A critical review *Geomorphology* 13(1-4), 233-253.
- van der Heijden, M.G.A., Bardgett, R.D., van Straalen, N.M., 2008. The unseen majority: soil microbes as drivers of plant diversity and productivity in terrestrial ecosystems. *Ecology Letters* 11(3), 296-310.
- Waldrop, M.P., Firestone, M.K., 2006. Seasonal dynamics of microbial community composition and function in oak canopy and open grassland soils. *Microbial Ecology* 52(3), 470-479.
- Waldrop, M.P., Zak, D.R., Sinsabaugh, R.L., Gallo, M., Lauber, C., 2004. Nitrogen deposition modifies soil carbon storage through changes in microbial enzymatic activity. *Ecological Applications* 14(4), 1172-1177.
- Weslien, P., Klemetsson, A.K., Borjesson, G., Klemetsson, L., 2009. Strong pH influence on N<sub>2</sub>O and CH<sub>4</sub> fluxes from forested organic soils. *European Journal of Soil Science* 60(3), 311-320.
- Willison, T.W., Webster, C.P., Goulding, K.W.T., Powlson, D.S., 1995. Methane oxidation in temperate soils - effects of land-use and the chemical form of nitrogen-fertilizer. *Chemosphere* 30(3), 539-546.
- Yates, C.J., Norton, D.A., Hobbs, R.J., 2000. Grazing effects on plant cover, soil and microclimate in fragmented woodlands in south-western Australia: implications for restoration. *Austral Ecology* 25(1), 36-47.
- Yuste, J.C., Baldocchi, D.D., Gershenson, A., Goldstein, A., Misson, L., Wong, S., 2007. Microbial soil respiration and its dependency on carbon inputs, soil temperature and moisture. *Global Change Biology* 13(9), 2018-2035.
- Zelles, L., 1997. Phospholipid fatty acid profiles in selected members of soil microbial communities. *Chemosphere* 35(1-2), 275-294.

## Part B - Publications



## Paper I

# **Cattle trampling alters soil properties and changes soil microbial communities in a Swiss sub-alpine pasture**

David Hiltbrunner <sup>ab\*</sup>, Sebastian Schulze <sup>a</sup>, Frank Hagedorn <sup>a</sup>, Michael W.I. Schmidt <sup>b</sup> and Stephan Zimmmermann <sup>a</sup>

<sup>a</sup> Swiss Federal Institute of Forest, Snow and Landscape Research (WSL), Zürcherstr. 111, CH-8903 Birmensdorf, Switzerland

<sup>b</sup> Department of Geography, University of Zurich, Winterthurerstr. 190, CH-8057 Zurich, Switzerland

Corresponding author. Tel: +41 44 7392 490, fax: +41 44 7392 215, *E-mail address:* [david.hiltbrunner@wsl.ch](mailto:david.hiltbrunner@wsl.ch) (D. Hiltbrunner)

**Published in Geoderma 170 (2012) 369–377**



# Cattle trampling alters soil properties and changes soil microbial communities in a Swiss sub-alpine pasture

David Hiltbrunner<sup>a,b,\*</sup>, Sebastian Schulze<sup>a</sup>, Frank Hagedorn<sup>a</sup>,  
Michael W.I. Schmidt<sup>b</sup>, Stephan Zimmermann<sup>a</sup>

<sup>a</sup> Swiss Federal Institute of Forest, Snow and Landscape Research (WSL), Zürcherstr. 111, CH-8903 Birmensdorf, Switzerland

<sup>b</sup> Department of Geography, University of Zurich, Winterthurerstr. 190, CH-8057 Zurich, Switzerland

## ARTICLE INFO

### Article history:

Received 30 May 2011

Received in revised form 20 October 2011

Accepted 25 November 2011

Available online 29 December 2011

### Keywords:

Alpine pastures

Livestock trampling

Soil erosion

Soil carbon

Microbial communities

## ABSTRACT

Stock farming plays an important role in the agriculture of alpine regions although deleterious effects on the soils are most pronounced here. We investigated the effects of cattle trampling on soil physical, chemical and microbial properties in a Swiss sub-alpine pasture. About 10% of the study site was bare of vegetation as a result of repeated cattle trampling and the bulk density of these bare steps was 20% higher than of the soils unaffected by trampling. In the upper 25 cm, soil organic carbon (SOC) concentrations and total SOC stocks were 35% and 20% respectively lower than on the vegetated slope. As compared with the vegetated slope, topsoils of the bare steps featured narrower C:N-ratios and were more enriched in the <sup>15</sup>N isotope, with typical values of deeper soil layers. This indicates that bare soils primarily evolved by erosion and not by a compaction, which might, together with the reduced litter input, explain the lower SOC contents. The abundances of soil microbes, estimated by the concentrations of phospholipid fatty acid (PLFA), were 30% smaller in the bare soils than in the vegetated areas. This depletion was most pronounced for fungi as expressed in the lower concentrations of the fatty acid 18:2ω6.9 (45%) and ergosterol (50%). The lower fungal abundance very likely has negative consequences for the stability of the bare soils, since fungi play an important role in the formation of soil aggregates. In summary, our results show that cattle trampling decreases soil carbon storage and alters soil microbial community structure.

© 2011 Elsevier B.V. All rights reserved.

## 1. Introduction

In mountainous regions stock farming plays an important role in agricultural land use since tillage is restricted due to steep slopes and a harsh climate. However, particularly in these regions livestock has the largest deleterious effects on soils (Trimble and Mendel, 1995) as overgrazing induces erosion by reducing the vegetation cover and infiltration rates (Gifford and Hawkins, 1978; Hamza and Anderson, 2005; Meeuwig, 1970). Nguyen et al. (1998) registered that cattle treading increased the transport of suspended solids, nitrogen and phosphorus from soils on steep slopes. Also, Wu and Tiessen (2002) found a significant loss of soil organic carbon (SOC) and total nitrogen (TN) from heavily degraded pastures in a Chinese alpine grassland. Additionally, soil microbial communities are likely to be altered by erosion with a smaller abundance of Gram negative bacteria as being reported after deforestation on the Loess plateau in China (Hamer et al., 2009). This, in turn, can influence the decomposition of soil organic matter (SOM) and the aggregate stability of these

soils, especially when fungi are affected (Bossuyt et al., 2001; Six et al., 2006).

Although the impact of grazing on soil properties has been widely assessed (Greenwood and McKenzie, 2001; Schuman et al., 1999; Steffens et al., 2008; Warren et al., 1986), studies taking the small-scale spatial soil heterogeneity into account are scarce (Csotonyi and Addicott, 2004; Golluscio et al., 2009). Cattle tracks, typically formed by repetitive trampling, are a common phenomenon in alpine pastures. The surface of the tracks is strongly modified by the trampling, leading to a specific micro-topography consisting of vegetated areas and bare steps. There is no study known to us assessing the formation of steps and its impact on the functioning of soils. It is not clear how the steps have formed, if they result from soil compaction or erosion. Moreover, the steps are likely to have an altered SOM dynamics as they are free of vegetation and denser than the surrounding soil. In conjunction with the different soil properties, this will also affect the composition and function of microbial communities. Given that the area of grasslands is rather large in the Alps (e.g. 537,801 ha in Switzerland corresponding to 35% of the agricultural land (BFS, 2005)) and also in other alpine regions across the world, small-scale variability of soil properties can accumulate to fairly large overall effects.

The aim of this study was (1) to elucidate the processes leading to steps in steep alpine grasslands and (2) to quantify the effects of

\* Corresponding author at: Swiss Federal Institute of Forest, Snow and Landscape Research (WSL), Zürcherstr. 111, CH-8903 Birmensdorf, Switzerland. Tel.: +41 44 7392 490; fax: +41 44 7392 215.

E-mail address: [david.hiltbrunner@wsl.ch](mailto:david.hiltbrunner@wsl.ch) (D. Hiltbrunner).

cattle trampling on physical, chemical and microbial properties of the soils. Our approach was to use stable isotope signatures of soils to gain insight into the formation of steps, to quantify soil C and N stocks, and to assess the effects of trampling on the soil microbial communities by analysing phospholipid fatty acids (PLFA). We hypothesized that (i) steps result from a combination of soil compaction and erosion; that (ii) the steps are depleted in SOM due to erosion and the lacking C input from vegetation; and that (iii) microbial biomass would decrease and shift towards less Gram negative bacteria in the step soils as a result of the smaller C availability.

## 2. Materials and methods

### 2.1. Study area and soil sampling

The study was conducted at a sub-alpine pasture in the Canton of Fribourg, Switzerland (7°15'54 E; 46°37'17 N). The site is located on a south-facing slope in an altitude between 1550 m a.s.l and 1650 m a.s.l. The mean summer and winter temperatures are 11.4 °C and 0.6 °C, respectively and mean annual precipitation is 1250 mm with a maximum during summer. Soils are Cambisols on calcareous bedrock with a stone content smaller than 5% in the analysed soil layer. The site was used as a pasture for at least 150 years, but most probably for centuries. To date, the pasture is used for summer grazing (June–September) with a livestock density of about 4 cattle ha<sup>-1</sup>.

To assess the impact of animal trampling on soil properties, we subdivided the almost horizontal cattle tracks into two categories according to the microtopography: 'Bare Steps' (BS) formed by intensive trampling and 'Vegetated Shoulders' (VS) in between the steps. 'Unaffected Slope' (US) between two tracks were unaffected by trampling and served as a control (Fig. 1; Fig. S1; Fig. S2).

For the further assessment of the trampling impact, we generated two different datasets. To obtain a larger scale spatial distribution of the soil SOC and TN concentrations and stocks, four blocks (A–D; 25 m × 25 m) were delineated across the whole site (200 m × 100 m) and five sampling plots within each block were chosen randomly (Fig. 2). From each plot, one soil core (50 mm diam.) per category was taken ( $n=60$ ) and subdivided into 0–5 cm,

5–15 cm and 15–25 cm increments for chemical analyses. In addition, another 60 soil cores (20 replicates per category) were taken to determine the soil bulk density.

To get a deeper insight into the category specific microbiology and activity, two additional blocks were fenced and in each block 15 chambers (five per category) were installed to measure soil respiration. After the last measurement, two soil subsamples (0–5 cm) were taken from each chamber. The subsamples used for microbial and chemical analysis were immediately put into a cool box and transported to a –20 °C refrigeration room within hours. The subsample for physical soil properties was taken with a stainless steel cylinder (0–5 cm) and a known volume.

### 2.2. Spatial proportion of the trampling categories

To observe the spatial organization of cattle tracks a georeferenced, pre-processed multi channel RGB aerial photo with a spatial resolution of 25 cm was loaded into a GIS using ArcGIS Desktop version 9.3 (ESRI Inc. 2009). All cattle tracks of the study site (2 ha, 200 m × 100 m) were visually vectorized as a continuous line feature and subsequently buffered with the average cattle track width producing a polygon feature representing the total area and location of cattle tracks in the study area (Fig. 2). The average width of the tracks was estimated by measuring the width of 90 consecutive bare steps and 90 vegetated shoulders along two randomly chosen transects. To further subdivide the area of the tracks into bare steps and vegetated shoulders, in total 160 photos of a wooden frame (50 cm × 50 cm) laid on the tracks were taken. Using Adobe photoshop (Adobe Systems Inc. 2009) the pictures were geometrically corrected and the pixels representing bare soil or vegetation were separated from each other. The percentage of the two classes was determined with the histogram function (Klassen et al., 2003). In order to validate the automatized cover estimation, 20 pictures were manually categorized using a superimposed 1 cm<sup>2</sup> grid.

### 2.3. Soil physical and chemical analysis

To calculate the respective stocks, concentrations of soil organic carbon and total nitrogen were measured and bulk densities of the soils were determined. The natural abundance of <sup>15</sup>N was used to

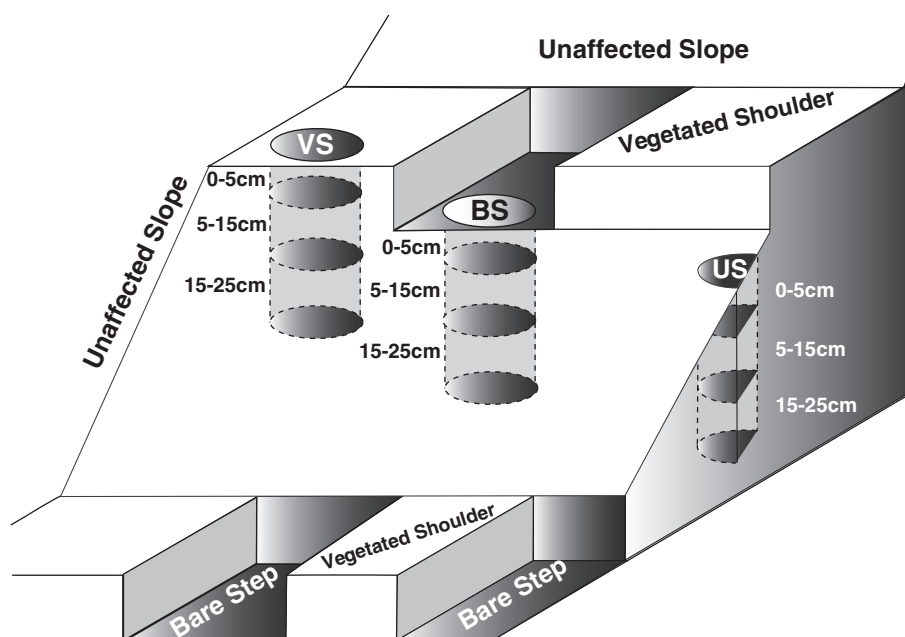
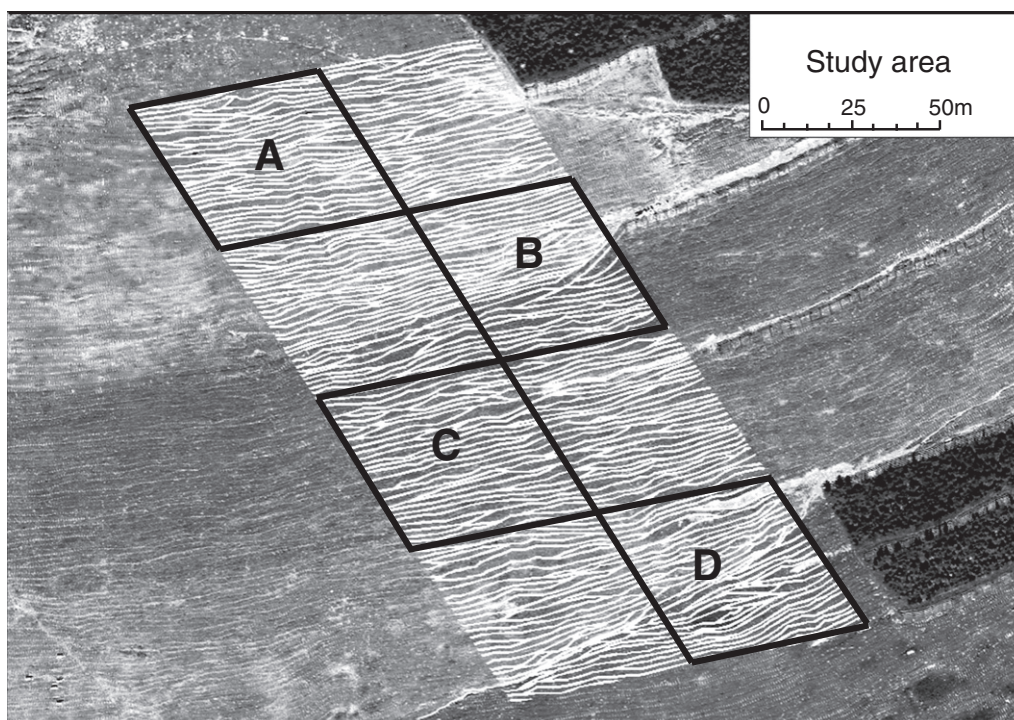


Fig. 1. Schematic illustration of the soil sampling within the different trampling categories: BS = 'Bare Step', VS = 'Vegetated Shoulder', US = 'Unaffected Slope'.



**Fig. 2.** Aerial photograph of the study site with cattle tracks (white lines) and blocks for the soil sampling (A–D).

estimate whether the steps were formed by erosion or compaction, because deviations from a 'natural'  $^{15}\text{N}$  enrichment with SOM aging and thus, with soil depth allows to identify if soil material was removed through erosion. Bulk soil density was calculated with the mass of the oven dried ( $105^\circ\text{C}$ ) soil weight divided by the volume of the cylinder. Particle density was determined by the pycnometer-method (Blake et al., 1986) and total porosity was calculated as one minus the ratio between bulk density and particle density. Prior to chemical analysis, soil samples were oven-dried ( $60^\circ\text{C}$ ) and sieved  $<2\text{ mm}$ . Soil pH was measured potentiometrically in  $0.01\text{ M CaCl}_2$  with a solid/extractant ratio of 1:2. SOC- and TN concentrations and the nitrogen isotopic ratio ( $\delta^{15}\text{N}$ ) values of the soils were determined from ground subsamples with an automated elemental analyser–continuous flow isotope ratio mass spectrometer (Euro-EA, Hekatech GmbH, Germany, interfaced with a Delta-V Advanced IRMS, Thermo GmbH, Germany).  $\delta^{15}\text{N}$  was reported in per mil (‰) relative to atmospheric  $\text{N}_2$ . Peach leaves (NIST-1547), previously validated against several IAEA standards were used as a reference ( $1.337 \pm 0.087\text{‰}$ ,  $n=29$ ). Since the bedrock of the site is calcareous, soil samples with  $\text{pH} > 6$  were fumigated with HCl prior to the analysis in order to dissolve carbonates (Walthert et al., 2010).

SOC and TN stocks were calculated by multiplying SOC and TN concentrations with the soil bulk density and the thickness of each depth class. As trampling considerably affected soil bulk densities, we additionally calculated the SOC stocks for a normalized soil mass as described by Ellert and Bettany (1995) and Steffens et al. (2008). The unaffected slope was used as reference and the respective soil mass per  $\text{m}^2$  and depth increment (0–5 cm, 5–15 cm, 15–25 cm) was calculated. The resulting masses were then multiplied with the carbon concentrations of the three trampling categories. Bulk densities were corrected for a stone content of 5% for stock calculations.

#### 2.4. Soil respiration

Soil respiration rates were measured to estimate the biological activity of the soils.  $\text{CO}_2$  efflux from soil was measured using a static chamber technique. Per block ( $n=2$ ), five PVC-chambers per

category were stuck into the soil and the volume of each chamber was determined individually. The chambers were installed two months before the first measurement in order to let the soil recover from physical disturbance. For the respiration measurements, the chambers were tightly closed and gas samples were taken with a syringe through a septum in the cover plate after 0, 10, and 20 min. Gas samples were analysed for  $\text{CO}_2$  concentrations with a gas chromatograph (Agilent 7890, fitted to a flame ionisation detector (FID)) using a Porapak Q packed column. Soil respiration rates were calculated from the slope of the  $\text{CO}_2$  concentrations. Measurements were carried out at five dates (10.08./13.08./16.08./03.09 and 01.10. 2010).

#### 2.5. Lipid extraction and PLFA analysis

Soil microbial community compositions were characterized by analysing concentrations of phospholipid fatty acids (PLFA) in soils (Frostegard et al., 2011). Microbial fatty acids were extracted from 1 g of fresh soil with a monophasic mixture (1:2:0.8 v/v/v) of chloroform, methanol and phosphate buffer (Bligh and Dyer, 1959; Zelles, 1997). For separating the organic and the water phase, accessory water and chloroform was added. The lipids were fractionated into neutral lipids, glycolipids and phospholipids on silicic acid columns, impregnated with ammonium acetate. The phospholipid fatty acids were methylated at  $60^\circ\text{C}$  for 2 h using trimethylchlorosilane and methanol (1:9 v/v) (Thiel et al., 2001). 19:0 was added as an internal standard prior to the GC/MS measurement. For PLFA detection, an Agilent GC–mass spectrometry (MS) system (HP 6890 N Plus gas chromatograph connected to a 5973 N MSD detector) with a 50 m capillary column (Agilent 128–5552 DB5–5MS) was used. The PLFAs were identified by comparing the retention times of the samples with those of a standard mixture (bacterial acid methyl esters, Supelco Inc.) and by inspection of mass spectra. Standard nomenclature was used to describe the fatty acids. First number refers to number of the C-atoms, followed by the number of double bonds and their position ( $\omega$ ). The prefixes “i” and “a” refer to iso- and anteiso-branched fatty acids. The prefix “cy” indicates cyclopropane fatty acids. 18:2 $\omega$ 6,9 was used as a biomarker for fungi, i15:0, a15:0,



i16:0, i17:0, a17:0 for Gram positive bacteria, 16:1 $\omega$ 5, 16:1 $\omega$ 7, cy17:0, 18:1 $\omega$ 7 and cy19:0 for Gram negative bacteria (Zelles, 1997; Zelles, 1999) and 10Me16:0 and 10Me18:0 for actinomycetes (Federle and Megusar, 1986). Total PLFA concentration (nmol PLFA per gram dry soil (nmol g<sup>-1</sup>)) was used as an index of the amount of living microbial biomass and the fungal/bacterial ratio (18:2 $\omega$ 6.9/Gram positive + Gram negative + 17:0 + 15:0) as an indicator for the microbial composition.

## 2.6. Ergosterol extraction

Ergosterol was extracted according to Daoud et al. (2008) with slight modifications. 1 g of moist soil was saponified for 30 min under reflux in 10 ml of a methanol/ethanol/KOH (5:2.5:1; v/v/w) mixture. The product was filtered with an ethanol rinse and after adding deionised water the ergosterol was extracted with 30 ml of n-hexane. The hexane-phase was dried over NaSO<sub>4</sub> and subsequently evaporated under vacuum at 40 °C. Before the injection into the HPLC column (Dionex, Purospher Star RP-18, diam. 3  $\mu$ m) the samples were redissolved in 1 ml of LC-grade methanol. The UV-detector was set at 282 nm.

## 2.7. Statistical analysis

Analysis of variance (ANOVA) was conducted to test the effect of animal trampling on soil physical, chemical and microbial properties using a randomized block design. Data that did not conform to the assumptions of ANOVA were log or square root transformed prior to analysis. Multiple comparisons of the means were performed with TukeyHSD Post Hoc test. Elemental stocks were calculated by the rules of error propagation. Linear regression was used to test the relationship between physico-chemical properties and microbial variables. For all statistical analyses the level of significance was  $p < 0.05$ . Soil respiration rates were analysed as average effluxes per chamber (mean of 5 dates) to avoid temporal pseudo-replication. The relationship between PLFA data (mol%) of the different samples and physico-chemical factors (i.e. pH, porosity, water content, C:N-ratio and  $\delta^{15}\text{N}$  values) was tested performing redundancy analysis (RDA), which is a linear canonical ordination method based on principle component analysis. RDA constraints the ordination axes to be linear combinations of the environmental variables (i.e. physico-chemical factors) and thus allows a direct comparison between multivariate data (PLFA) and environmental variables (Terbraak and Prentice, 1988). RDA was preferred over canonical correspondence analysis (cca) since the PLFA data showed a linear rather than a unimodal response

(Terbraak and Šmilauer, 2002). Environmental variables and species (PLFA) are presented as vectors in the RDA-plot and their position and length indicate direction and size of increase. Species and environmental vectors pointing in the same direction, are assumed to be well correlated and increasing length of the vector indicates increasing confidence in the correlation (Terbraak and Prentice, 1988). All statistical analyses were performed with the R 2.10.1 (<http://www.r-project.org>) software package and the vegan library (Oksanen et al., 2007) was used for RDA analysis.

## 3. Results

### 3.1. Spatial proportion of the trampling categories

Cattle tracks made a contribution of  $0.87 \pm 0.11$  ha to the study area (2 ha), while  $1.13 \pm 0.11$  ha were unaffected by the trampling. This corresponded to a total track length of 12.4 km with an average track width of  $72 \text{ cm} \pm 9.67 \text{ cm}$ . On the tracks, the bare steps covered  $0.21 \pm 0.01$  ha and the vegetated shoulders  $0.66 \pm 0.01$  ha. Overall, unaffected slopes accounted for 57%, vegetated shoulders for 33% and bare steps for 10% of the total area.

### 3.2. Soil properties

Across the whole site, bulk density in the upper 0–25 cm of the BS was significantly ( $p < 0.001$ ) higher compared to VS and US while the latter did not differ significantly. Soil pH values were in a rather narrow range between 4.7 and 5.3 and no clear trend of an increasing pH with soil depth was observable. However, pH-values in the BS soils were slightly higher throughout the whole soil profile compared to the other categories (Table 1).

In the two blocks, which were used for microbial community assessment, bulk density values (0–5 cm) of the bare steps were also significantly higher than in the other two categories (Table 2). Soils of the bare steps had also higher particle densities and significantly lower soil porosities as compared to the vegetated categories (Table 2).

### 3.3. SOC, TN, $\delta^{15}\text{N}$ and stocks

SOC and TN concentrations decreased with soil depth in all categories (Fig. 4; Table 1). Within the soil profile, SOC decreased between 44% and 53%, while the decrease for TN was less pronounced (27%–42%). This resulted in narrower C:N-ratios in the deeper soils than in the surface soils (Table 1). Trampling affected SOC and TN

**Table 1**  
Soil properties, C-, and N-stocks within the different trampling- and depth classes (with standard errors) and the whole soil profile (bold numbers). Different letters in the same column indicate statistical significant differences between the categories according to Tukey HSD ( $p < 0.05$ ).

Category	Depth (cm)	Concentrations								Stocks (0–25 cm)							
		TN [mg g <sup>−1</sup> ]		C/N		δ <sup>15</sup> N (‰)		pH (CaCl <sub>2</sub> )		Bulk density [g cm <sup>−3</sup> ]		SOC <sup>b</sup> [kg m <sup>−2</sup> ]		TN <sup>b</sup> [kg m <sup>−2</sup> ]		SOC <sup>c</sup> [kg m <sup>−2</sup> ]	
Bare Steps (10%) <sup>a</sup>	0–5	3.86	(0.1)	10.1	(0.28)	2.79	(0.13)	5.3	(0.09)	0.88	(0.04)	1.4	(0.2)	0.15	(0.02)	1.1	(0.2)
	5–15	3.16	(0.2)	9.1	(0.46)	3.88	(0.22)	4.9	(0.09)	1.12	(0.03)	2.8	(0.2)	0.31	(0.02)	2.2	(0.1)
	15–25	2.35	(0.1)	8.4	(0.62)	5.20	(0.21)	5.2	(0.12)	1.12	(0.02)	1.8	(0.2)	0.23	(0.02)	1.7	(0.1)
	0–25	2.98 A	(0.1)	9.0 A	(0.28)	4.21 A	(0.17)	5.1 A	(0.11)	1.07 A	(0.02)	6.0	(0.3)	0.69	(0.03)	5.0	(0.3)
Vegetated Shoulders (33%) <sup>a</sup>	0–5	5.06	(0.1)	12.2	(0.27)	1.85	(0.16)	4.8	(0.08)	0.74	(0.04)	1.9	(0.3)	0.15	(0.02)	1.8	(0.3)
	5–15	3.92	(0.1)	10.8	(0.54)	3.03	(0.18)	4.7	(0.08)	0.85	(0.03)	3.0	(0.2)	0.28	(0.02)	3.2	(0.2)
	15–25	2.86	(0.1)	10.7	(0.86)	4.25	(0.20)	4.8	(0.10)	0.99	(0.03)	2.5	(0.1)	0.25	(0.02)	2.7	(0.2)
	0–25	3.72 B	(0.1)	11.1 B	(0.35)	3.28 B	(0.16)	4.8 A	(0.09)	0.88 B	(0.02)	7.4	(0.4)	0.68	(0.03)	7.7	(0.4)
Unaffected Slope (57%) <sup>a</sup>	0–5	4.58	(0.1)	12.0	(0.29)	2.20	(0.16)	5.0	(0.07)	0.71	(0.03)	1.6	(0.2)	0.13	(0.02)	1.6	(0.3)
	5–15	3.88	(0.1)	10.7	(0.23)	3.00	(0.16)	4.8	(0.08)	0.91	(0.02)	3.2	(0.2)	0.30	(0.02)	3.2	(0.2)
	15–25	3.34	(0.2)	9.7	(0.30)	4.19	(0.20)	4.9	(0.11)	1.06	(0.03)	2.8	(0.2)	0.31	(0.01)	2.8	(0.2)
	0–25	3.80 B	(0.1)	10.5 B	(0.20)	3.32 B	(0.15)	4.9 A	(0.08)	0.93 B	(0.02)	7.6	(0.4)	0.74	(0.03)	7.6	(0.4)

<sup>a</sup> Proportion of the trampling category on the total area.

<sup>b</sup> Stocks per m<sup>2</sup>.

<sup>c</sup> Stocks per m<sup>2</sup> calculated for an equivalent soil mass.

**Table 2**

Soil properties and PLFA concentrations [nmol g<sup>-1</sup> dry soil] of the soil respiration plots (with standard errors). Different letters in the same row indicate statistical significant differences according to Tukey HSD ( $p < 0.05$ ).

	Category					
	Bare steps		Vegetated shoulders		Unaffected slope	
Bulk density [g cm <sup>-3</sup> ]	1.01 a	(0.04)	0.84 b	(0.07)	0.83 b	(0.03)
Particle density [g cm <sup>-3</sup> ]	2.43 a	(0.03)	2.38 b	(0.02)	2.39 ab	(0.02)
Porosity (%)	0.58 a	(0.029)	0.65 b	(0.03)	0.65 b	(0.01)
pH (CaCl <sub>2</sub> )	5.6 a	(0.27)	5.1 ab	(0.27)	5.0 b	(0.16)
C [mg g <sup>-1</sup> ]	43.1 a	(5.4)	56.4 b	(6.9)	49.7 ab	(2.2)
N [mg g <sup>-1</sup> ]	4.00 a	(0.36)	4.64 b	(0.36)	4.37 ab	(0.16)
C:N	10.6 a	(0.42)	12.0 b	(0.54)	11.4 ab	(0.18)
$\delta^{15}\text{N}$ (‰)	2.82 a	(0.42)	1.86 b	(0.27)	2.17 b	(0.19)
Soil resp. (μmol m <sup>-2</sup> s <sup>-1</sup> )	2.03 a	(0.16)	3.47 b	(0.17)	3.35 b	(0.16)
Ergosterol (μg g <sup>-1</sup> dry soil)	4.73 a	(1.03)	10.1 b	(1.96)	8.98 b	(1.08)
PLFA						
Total <sup>a</sup>	385 a	(27.2)	534 b	(30.6)	538 b	(7.77)
Bacteria <sup>a</sup>	210 a	(13.9)	290 b	(16.9)	291 b	(5.5)
Gram positive <sup>a</sup>	84 a	(5.4)	124 b	(8.3)	124 b	(3.5)
Gram negative <sup>a</sup>	118 a	(8.4)	155 b	(8.3)	157 b	(3.6)
Actinomycetes <sup>a</sup>	28.3 a	(1.3)	39.7 b	(2.5)	37.8 b	(1.1)
Fungi:Bacteria	0.070 a	(0.002)	0.094 b	(0.005)	0.094 b	(0.004)
Fungi <sup>a</sup>	15.2 a	(1.5)	27.6 b	(2.1)	27.6 b	(0.99)

<sup>a</sup> Sum of PLFA representative for a distinct microbial group [nmol g<sup>-1</sup> dry soil].

concentrations: the upper 0–25 cm of the BS soils had about 35% and 20% smaller concentrations of SOC and TN than soils on the vegetated slope, respectively ( $p < 0.001$ ).

SOC concentrations of the uppermost 0–5 cm of the BS soils corresponded to the 5–15 cm layer of VS and US soils, which were on the same surface level (Fig. 1; Fig. 4). TN concentrations showed the same trend, however, the variance in the N data was larger and the differences between categories and depth classes were smaller. The  $\delta^{15}\text{N}$  values increased with soil depth in all categories and the values of one depth class of the BS soil corresponded to the ones of the deeper depth class in VS and US soils (Table 1; Fig. 3). The uppermost 25 cm of the US soils stored about 7.6 kg m<sup>-2</sup> organic carbon and 0.74 kg m<sup>-2</sup> nitrogen. The BS soils of the same depth contained only 6 kg m<sup>-2</sup> SOC and 0.69 kg m<sup>-2</sup> TN (Table 1). If a normalized

soil mass was used for the stock calculation, the difference between the steps and the vegetated categories was even more pronounced (Table 1).

### 3.4. Soil respiration and microbiology

Soil respiration in the BS soils was about 40% reduced compared to VS and US (Table 2). Respiration rates were negatively correlated to bulk density ( $R^2 = 0.29$ ;  $p < 0.01$ ),  $\delta^{15}\text{N}$  values ( $R^2 = 0.23$ ;  $p < 0.01$ ) and positively to total PLFA ( $R^2 = 0.58$ ;  $p < 0.001$ ).

PLFA analysis revealed a 28% lower bacterial and 45% lower fungal abundance in BS soils compared to the vegetated areas. As a result, the fungi:bacteria-ratio was significantly lower in the bare steps ( $p < 0.001$ ). The decrease was also reflected in the concentrations of the fungal biomarker ergosterol, which were about 50% lower in the steps. In contrast, no differences between the vegetated categories VS and US could be observed in the PLFA concentrations, while the ergosterol concentrations were slightly higher in the soils of the vegetated shoulders (Table 2). Total PLFA and ergosterol concentrations were positively correlated to SOC ( $R^2 = 0.43$  and  $R^2 = 0.64$ , respectively;  $p < 0.001$  for both) and negatively to bulk density ( $R^2 = 0.50$  and  $R^2 = 0.41$ , respectively;  $p < 0.001$  for both). Moreover, the fungi:bacteria-ratio increased with wider C:N-ratios of the soil organic matter ( $R^2 = 0.25$ ;  $p < 0.01$ ). Both fungal markers, ergosterol and the fatty acid 18:2ω6.9 were closely correlated either in absolute and relative values of the fatty acid ( $R^2 = 0.55$  and  $R^2 = 0.57$  respectively;  $p < 0.001$  for both).

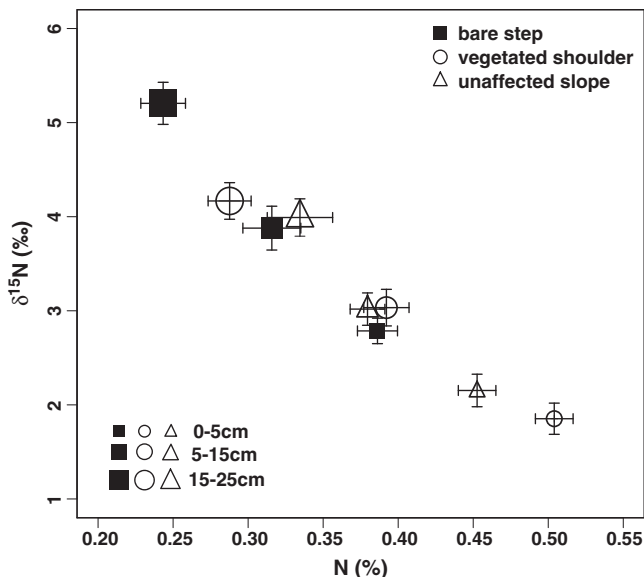
Redundancy analysis (RDA) showed that 48% of the variance in the microbial community matrix could be related to environmental variables (proportion of constraint inertia on the total inertia), with the first two axes explaining nearly 90% of this variation. RDA 1 was related to soil pH while RDA 2 was correlated to the quality of soil organic matter (C:N-ratio and  $\delta^{15}\text{N}$  values). Block 1 and block 2 separated along RDA 2, reflecting the different C:N-ratios (11.9 block 1 and 10.7 block 2, respectively) and  $\delta^{15}\text{N}$  values (1.8 block 1 and 2.8 block 2, respectively) (Fig. 5).

## 4. Discussion

### 4.1. Step formation: erosion or compaction?

Vegetation free 'bare' steps are a common feature of steep alpine grasslands. They have evolved either by soil compaction or erosion or the combination both. The contribution of these two processes can only be roughly quantified since they are closely linked and very likely site specific. Mechanical forces of the hooves degrade vegetation until the steps are bare. In addition, hooves compact soils and reduce soil porosity. Compaction is favoured through the lack of vegetation, since roots provide a filamentous network, which resists compressing forces (Soane, 1990). The unprotected, bare soil surface in the steps is potentially prone to erosion particularly in mountainous regions with high precipitation and steep slopes. Soil compaction decreases infiltration rates and thus leads to an increased surface runoff, which in turn results in higher soil erosion (Trimble and Mendel, 1995; Warren et al., 1986). Despite the tight coupling of the compaction and the erosion processes, we tried to unravel them by comparing physical soil properties of the different trampling classes and by applying the analysis of stable  $^{15}\text{N}$  isotopes.

The 20% higher bulk densities and concomitantly lower soil porosities imply a certain compaction of the bare step soils. However, taking into account the lower soil surface level of the bare steps compared to the vegetated areas (Fig. 1), the higher bulk densities in the steps could also be the result of an erosive loss of the topsoil. The latter assumption is supported by the depth distribution of  $^{15}\text{N}$  in the profiles. In all categories, the  $^{15}\text{N}$  abundance increased with soil depth most likely as a result of increasing age of SOM with



**Fig. 3.** N-concentrations vs.  $^{15}\text{N}$  isotopes indicate the discrimination of the  $^{15}\text{N}$  isotope with soil depth in the different trampling categories.

depth (Conen et al., 2008). Discrimination processes during SOM turnover lead to a loss of  $^{15}\text{N}$ -depleted forms of nitrogen ( $\text{NO}_3$ ,  $\text{N}_2\text{O}$  etc.) while the heavier  $^{15}\text{N}$  isotope accumulates in soils (Amundson et al., 2003; Billings and Richter, 2006; Nadelhoffer and Fry, 1988). If soil compaction would have been the dominant process for the step formation, we would expect that the topsoils of the bare steps would have similar  $\delta^{15}\text{N}$  than the vegetated shoulders as  $^{15}\text{N}$ -depleted topsoils would have been compressed by the trampling into the deeper soil. However, the  $\delta^{15}\text{N}$  values in the steps of surface soils (0–5 cm) in the steps corresponded to those at 5–15 cm depth in the vegetated areas (Fig. 3, Table 1). Consequently, the depth profile of  $\delta^{15}\text{N}$  was capped, which can only be explained by a loss of topsoil material and hence, by erosion. Alternatively, the different  $^{15}\text{N}$  enrichment with soil depth in the steps and surrounding soil could have resulted from a different microbial nitrogen cycling. Though, the degree of the  $^{15}\text{N}$  discrimination, as expressed by the discrimination factor ( $\ln(N)$  vs.  $\delta^{15}\text{N}$ , (Nadelhoffer and Fry, 1988)), showed no significant differences between the trampling categories (Fig. 3), suggesting that microbial nitrogen turnover remained largely unaffected by trampling. We therefore conclude that erosion and not compaction was the dominant process for the step formation.

#### 4.2. Implications for C dynamics

In the uppermost soil (0–5 cm) of the bare steps, soil carbon concentrations were about 35% smaller than in the vegetated soils. This depletion was also apparent in the deeper soils of the bare steps (5–15 cm, 15–25 cm), where carbon concentrations were about 25% lower as compared to the corresponding depth classes of the vegetated plots (Fig. 4). The decrease in soil C is most probably the result of three different processes, (I) erosion of the unprotected bare soils, (II) reduced C input due to the lack of vegetation and (III) soil aggregate disruption through trampling. In the bare steps, physical protection and aggregate stability of the soils are reduced. When the soil surface is not protected by vegetation, it is entirely exposed to the eroding power of raindrops and the steps form a perfect channel system for overland water-flow. Hence, SOM-rich topsoil had been eroded over decades and the remaining soils were depleted in organic C and N with typical contents of deeper soil horizons. A second reason for the lower C concentration and stocks in the bare steps is the reduced biomass input. In a shortgrass steppe, Kelly et al. (1996) reported 28%

less SOC at locations with little or no plant input for about 45 years and a plant removal study within the Rothamsted experiments showed a loss of 27% soil carbon after 40 years (Jenkinson and Rayner, 1977). Besides the reduced SOM input, the protection of SOM is negatively impacted by the trampling. Organic matter occluded in soil aggregates is often inaccessible for microbes and thus protected against decomposition (Six et al., 2002). Treading disrupts these aggregates and formerly protected organic material might decompose rapidly. In a semi-arid steppe, Steffens et al. (2008) found a significant negative correlation between SOC and bulk density as a result of grazing intensity. They speculated that physical disruption of soil aggregates, together with wind erosion were the likely reasons for this loss of SOM. In agreement, in a semi-arid grassland, the amount of stable aggregates in cattle tracks was three times smaller than in ungrazed areas (Zhou et al., 2010). At our site on a steep slope, trampling effects are highly concentrated to a relatively small area, which has even more deleterious implications on soil aggregate structure. Since the site has been grazed for at least 150 years, it is likely that most of the formerly protected carbon has been mineralized or eroded. Lower soil respiration rates in the bare steps (Table 2) are presumably the result of this degradation. Although soil compaction can suppress C and N mineralization and retard the carbon loss of soils, this is not likely to happen at our site since bulk density is quite low for pastures. In a loamy sand soil, De Neve and Hofman (2000) found that a reduction of mineralization occurred only at bulk densities  $\geq 1.5 \text{ g cm}^{-3}$ , which is much higher than the  $1.1 \text{ g cm}^{-3}$  in the soils of the bare steps. More likely, lower microbial abundance together with an almost absent autotrophic respiration accounted for the reduced soil respiration in the steps, reflecting the degraded soil properties here.

Not only the concentrations but also the stocks of C were affected by the trampling. In the uppermost 0–25 cm of the bare step soils, C storage was about  $6 \text{ kg m}^{-2}$  while in the vegetated areas the stocks ranged from  $7.4 \text{ kg m}^{-2}$  (VS) to  $7.6 \text{ kg m}^{-2}$  (US). Accordingly, the loss of SOC by trampling accounted for about  $1.5 \text{ kg m}^{-2}$  in BS, which is 20% of the total stock in this layer (Table 1). This loss was even higher (2.5 kg; 30%) when an equivalent soil mass was used for the stock calculations (Table 1), which might be more appropriate in this study, since trampling significantly compacted the step soils.

About 10% of the study site was classified as bare step soils. Thus, calculated on an areal basis, about  $0.15 \text{ kg m}^{-2}$  or 2% ( $0.25 \text{ kg}$  or 3.5%; equivalent soil mass) of the total carbon stocks (0–25 cm) were lost due the cattle trampling. This loss is not negligible, however, compared to other sites it is rather small. In a semi-arid Mongolian grassland Steffens et al. (2008) estimated that heavy grazing for 35 years resulted in 34% lower C stocks of the 0–4 cm layer as compared to ungrazed sites. In contrast to their study, however, there were no ungrazed enclosures in our experiment.

Although our data show clear evidence that SOC is lost from the bare steps through erosion, there is still an on-going debate about the fate of the eroded organic material (Harden et al., 2008; Lal, 2005; Lal et al., 2004; Van Oost et al., 2007). There are only few field studies, which have investigated to which amount SOC is relocated, buried or mineralized during the erosion process. Analysing  $^{137}\text{Cs}$ - and  $^{13}\text{C}$ -isotopes from eroded soils in a Swiss sub-alpine grassland under similar site conditions like in our study, Alewell et al. (2009) concluded that a considerable amount of the eroded SOC has been degraded during detachment and transport. However, they stated that the results are very site specific and cannot be generalized. Therefore, more research would be needed to elucidate the fate of SOC lost through erosion from cattle tracks.

#### 4.3. Cattle trampling affects soil microorganisms

The significantly different soil physical properties and SOM concentrations of the steps were also reflected in the microbial

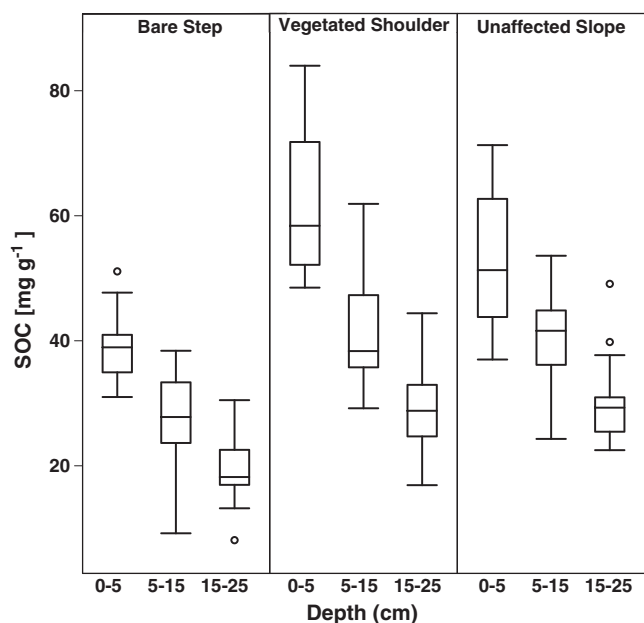


Fig. 4. SOC-concentrations across trampling categories and soil depths.





## 5. Summary and conclusions

Our results in a subalpine pasture showed that cattle trampling strongly affected the physical, chemical and microbial properties of soils. After decades of grazing, soil surface exhibited a distinct micro-topographical pattern, which is a typical feature of alpine pastures. The trampling led to denser soils and the  $\delta^{15}\text{N}$  depth patterns indicated that substantial amounts of the uppermost 10 cm of the vegetation-free steps were lost through erosion. The bare steps (0–25 cm) had 35% smaller soil C concentrations and 20% smaller soil C stocks than the vegetated areas. As a result, the biomass of soil microbes was 30% lower in the bare steps. This decrease was most pronounced for fungi leading to lower fungal to bacterial ratios in the bare steps. Together with the absence of plant roots, the reduction of fungi, which strongly contribute to the formation of soil aggregates, might further accelerate soil erosion in the bare steps.

The strong change in soil properties by the trampling presumably has a strong impact on many soil functions. Greenhouse gas fluxes, for instance, are likely to be altered with lower porosity as a driving factor. Thus, to obtain a more holistic picture of all the relevant processes, more research is needed.

Supplementary materials related to this article can be found online at doi: [10.1016/j.geoderma.2011.11.026](https://doi.org/10.1016/j.geoderma.2011.11.026).

## Acknowledgements

We gratefully thank P. A. Niklaus for the measurement of the gas samples and M. Hilf for the measurement of the PLFA samples (University of Zurich). We also thank M. Walser, R. Köchli and O. Schramm for the field assistance, G.D. Lieberherr for assistance in the laboratory and A. Schlumpf for the HPLC analysis. This study was funded by the COST Action 639 (BurnOut) and the Swiss Federal Office for the Environment (FOEN).

## References

- Alewell, C., Schaub, M., Conen, F., 2009. A method to detect soil carbon degradation during soil erosion. *Biogeosciences* 6 (11), 2541–2547.
- Amundson, R., Austin, A.T., Schuur, E.A.G., Yoo, K., Matzek, V., Kendall, C., Uebersax, A., Brenner, D., Baisden, W.T., 2003. Global patterns of the isotopic composition of soil and plant nitrogen. *Global Biogeochemical Cycles* 17 (1).
- Baath, E., Anderson, T.H., 2003. Comparison of soil fungal/bacterial ratios in a pH gradient using physiological and PLFA-based techniques. *Soil Biology and Biochemistry* 35 (7), 955–963.
- Beylich, A., Oberholzer, H.R., Schrader, S., Hoper, H., Wilke, B.M., 2010. Evaluation of soil compaction effects on soil biota and soil biological processes in soils. *Soil and Tillage Research* 109 (2), 133–143.
- Bundesamt für Statistik (BFS), 2005. Arealstatistik Schweiz Zahlen Fakten Analysen 2005.
- Billings, S.A., Richter, D.D., 2006. Changes in stable isotopic signatures of soil nitrogen and carbon during 40 years of forest development. *Oecologia* 148 (2), 325–333.
- Blake, G.R., Hartge, K.H., Klute, A.E., 1986. Particle Density. In: Klute, A. (Ed.), *Methods of Soil Analysis, Part I. Physical and Mineralogical Methods*, pp. 363–375. American Society of Agronomy, Inc., and Soil Science Society of America. Madison (Wisconsin), USA.
- Bligh, E.G., Dyer, W.J., 1959. A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology* 37 (8), 911–917.
- Bossuyt, H., Deneff, K., Six, J., Frey, S.D., Merckx, R., Paustian, K., 2001. Influence of microbial populations and residue quality on aggregate stability. *Applied Soil Ecology* 16 (3), 195–208.
- Bowen, R.M., Harper, S.H.T., 1990. Decomposition of wheat straw and related compounds by fungi isolated from straw in arable soils. *Soil Biology and Biochemistry* 22 (3), 393–399.
- Busse, M.D., Beattie, S.E., Powers, R.F., Sanchez, F.G., Tiarks, A.E., 2006. Microbial community responses in forest mineral soil to compaction, organic matter removal, and vegetation control. *Canadian Journal of Forest Research* 36 (3), 577–588.
- Calderon, F.J., Jackson, L.E., Scow, K.M., Rolston, D.E., 2000. Microbial responses to simulated tillage in cultivated and uncultivated soils. *Soil Biology and Biochemistry* 32 (11–12), 1547–1559.
- Conen, F., Zimmermann, M., Leifeld, J., Seth, B., Alewell, C., 2008. Relative stability of soil carbon revealed by shifts in delta N-15 and C:N ratio. *Biogeosciences* 5 (1), 123–128.
- Csotonyi, J.T., Addicott, J.F., 2004. Influence of trampling-induced microtopography on growth of the soil crust bryophyte *Ceratodon purpureus* in Jasper National Park. *Canadian Journal of Botany* 82 (9), 1382–1392.
- Daoud, H.G., Korbász, M., Hamdan, S., Beczner, J., 2008. Simultaneous LC determination of ergosterol, tocopherols and carotenoids in foods. *Chromatographia* 68, 137–140.
- De Neve, S., Hofman, G., 2000. Influence of soil compaction on carbon and nitrogen mineralization of soil organic matter and crop residues. *Biology and Fertility of Soils* 30 (5–6), 544–549.
- Dick, R.P., Myrold, D.D., Kerle, E.A., 1988. Microbial biomass and soil enzyme activities in compacted and rehabilitated skid trail soils. *Soil Science Society of America Journal* 52 (2), 512–516.
- Ellert, B.H., Bettany, J.R., 1995. Calculation of organic matter and nutrients stored in soils under contrasting management regimes. *Canadian Journal of Soil Science* 75 (4), 529–538.
- Federle, T.W., Megusar, F.G.M.E., 1986. *Microbial distribution in soil - new techniques*. Slovene Society for Microbiology, Ljubljana, pp. 493–498.
- Fierer, N., Schimel, J.P., Holden, P.A., 2003. Variations in microbial community composition through two soil depth profiles. *Soil Biology and Biochemistry* 35 (1), 167–176.
- Frostegard, A., Tunlid, A., Baath, E., 2011. Use and misuse of PLFA measurements in soils. *Soil Biology and Biochemistry* 43 (8), 1621–1625.
- Gifford, G.F., Hawkins, R.H., 1978. Hydrologic impact of grazing on infiltration: a critical review. *Water Resources Research* 14 (2), 305–313.
- Golluscio, R.A., Austin, A.T., Martinez, G.C.G., Gonzalez-Polo, M., Sala, O.E., Jackson, R.B., 2009. Sheep grazing decreases organic carbon and nitrogen pools in the Patagonian Steppe: combination of direct and indirect effects. *Ecosystems* 12 (4), 686–697.
- Greenwood, K.L., McKenzie, B.M., 2001. Grazing effects on soil physical properties and the consequences for pastures: a review. *Australian Journal of Experimental Agriculture* 41 (8), 1231–1250.
- Griffiths, B.S., Ritz, K., Ebbelwhite, N., Dobson, G., 1999. Soil microbial community structure: effects of substrate loading rates. *Soil Biology and Biochemistry* 31 (1), 145–153.
- Guckert, J.B., Hood, M.A., White, D.C., 1986. Phospholipid ester-linked fatty acid profile changes during nutrient deprivation of *Vibrio cholerae*: increases in the trans/cis ratio and proportions of cyclopropyl fatty acids. *Applied and Environmental Microbiology* 52 (4), 794–801.
- Hamer, U., Makeschin, F., An, S., Zheng, F.L., 2009. Microbial activity and community structure in degraded soils on the Loess Plateau of China. *Journal of Plant Nutrition and Soil Science* 172 (1), 118–126.
- Hamza, M.A., Anderson, W.K., 2005. Soil compaction in cropping systems – a review of the nature, causes and possible solutions. *Soil Tillage Research* 82 (2), 121–145.
- Harden, J.W., Berhe, A.A., Torn, M., Harte, J., Liu, S., Stallard, R.F., 2008. Soil erosion: Data say C sink. *Science* 320 (5873), 178–179.
- Ibekwe, A.M., Kennedy, A.C., Halvorson, J.J., Yang, C.H., 2007. Characterization of developing microbial communities in Mount St. Helens pyroclastic substrate. *Soil Biology and Biochemistry* 39 (10), 2496–2507.
- Jenkinson, D.S., Rayner, J.H., 1977. The turnover of soil organic matter in some of the Rothamsted classical experiments. *Soil Science* 123 (5), 298–305.
- Kaur, A., Chaudhary, A., Choudhary, R., Kaushik, R., 2005. Phospholipid fatty acid – a bioindicator of environment monitoring and assessment in soil ecosystem. *Current Science* 89 (7), 1103–1112.
- Kelly, R.H., Burke, I.C., Lauenroth, W.K., 1996. Soil organic matter and nutrient availability responses to reduced plant inputs in shortgrass steppe. *Ecology* 77 (8), 2516–2527.
- Kieft, T.L., Ringelberg, D.B., White, D.C., 1994. Changes in ester-linked phospholipid fatty acid profiles of subsurface bacteria during starvation and desiccation in a porous medium. *Applied and Environmental Microbiology* 60 (9), 3292–3299.
- Klassen, S.P., Ritchie, G., Frantz, J.M., Pinnock, D., Bugbee, B., 2003. Real-time imaging of ground cover: relationships with radiation capture, canopy photosynthesis, and daily growth rate. In: Van Toai, T., Major, D., McDonald, M., Schepers, J., Tarpley, L. (Eds.), *Digital imaging and spectral techniques: applications to precision agriculture and crop physiology*. Am. Soc. Agronomy special publication No. 66. Madison, WI, pp. 3–15.
- Lal, R., 2005. Soil erosion and carbon dynamics. *Soil Tillage Research* 81, 137–142.
- Lal, R., Griffin, M., Apt, J., Lave, L., Morgan, M.G., 2004. *Ecology - Managing soil carbon*. Science 304 (5669) 393–393.
- Li, C.H., Ma, B.L., Zhang, T.Q., 2002. Soil bulk density effects on soil microbial populations and enzyme activities during the growth of maize (*Zea mays* L.) planted in large pots under field exposure. *Canadian Journal of Soil Science* 82 (2), 147–154.
- McKinley, V.L., Peacock, A.D., White, D.C., 2005. Microbial community PLFA and PHB responses to ecosystem restoration in tallgrass prairie soils. *Soil Biology and Biochemistry* 37 (10), 1946–1958.
- Meeuwij, R.O., 1970. Infiltration and soil erosion as influenced by vegetation and soil in northern Utah. *Journal of Range Management* 23 (3), 185–188.
- Nadelhoffer, K.J., Fry, B., 1988. Controls on natural nitrogen-15 and carbon-13 abundances in forest soil organic matter. *Soil Science Society of America Journal* 52 (6), 1633–1640.
- Nguyen, M.L., Sheath, G.W., Smith, C.M., Cooper, A.B., 1998. Impact of cattle treading on hill land 2. Soil physical properties and contaminant runoff. *New Zealand Journal of Agricultural Research* 41 (2), 279–290.
- O'Dea, M.E., 2007. Fungal mitigation of soil erosion following burning in a semi-arid Arizona savanna. *Geoderma* 138 (1–2), 79–85.
- Oksanen, J., Kindt, R., Legendre, P., O'Hara, B., Stevens, M.H.H., 2007. *Vegan: Community Ecology Package R package version 1.8-8*. <http://r-forge.r-project.org/projects/vegan/>.
- Potthoff, M., Steenwerth, K.L., Jackson, L.E., Drenovsky, R.E., Scow, K.M., Joergensen, R.G., 2006. Soil microbial community composition as affected by restoration practices in California grassland. *Soil Biology and Biochemistry* 38 (7), 1851–1860.
- Renger, M., 1970. Über den Einfluss der Dränung auf das Gefüge und die Wasserdurchlässigkeit bindiger Böden. *Mitteilung Deutsch Bodenkundl Gesellschaft* 11, 23–28.
- Rillig, M.C., Wright, S.F., Eviner, V.T., 2002. The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: comparing effects of five plant species. *Plant and Soil* 238 (2), 325–333.

- Ritz, K., Young, I.M., 2004. Interactions between soil structure and fungi. *Mycologist* 18, 52–59.
- Schuman, G.E., Reeder, J.D., Manley, J.T., Hart, R.H., Manley, W.A., 1999. Impact of grazing management on the carbon and nitrogen balance of a mixed-grass rangeland. *Ecological Applications* 9 (1), 65–71.
- Shestak, C.J., Busse, M.D., 2005. Compaction alters physical but not biological indices of soil health. *Soil Science Society of America Journal* 69 (1), 236–246.
- Six, J., Conant, R.T., Paul, E.A., Paustian, K., 2002. Stabilization mechanisms of soil organic matter: implications for C-saturation of soils. *Plant and Soil* 241 (2), 155–176.
- Six, J., Frey, S.D., Thiet, R.K., Batten, K.M., 2006. Bacterial and fungal contributions to carbon sequestration in agroecosystems. *Soil Science Society of America Journal* 70 (2), 555–569.
- Soane, B.D., 1990. The role of organic matter in soil compactibility: a review of some practical aspects. *Soil Tillage Research* 16 (1–2), 179–201.
- Steenwerth, K.L., Jackson, L.E., Calderon, F.J., Stromberg, M.R., Scow, K.M., 2002. Soil microbial community composition and land use history in cultivated and grassland ecosystems of coastal California. *Soil Biology and Biochemistry* 34 (11), 1599–1611.
- Steffens, M., Kolbl, A., Totsche, K.U., Kogel-Knabner, I., 2008. Grazing effects on soil chemical and physical properties in a semiarid steppe of Inner Mongolia (PR China). *Geoderma* 143 (1–2), 63–72.
- Terbraak, C.J.F., Prentice, I.C., 1988. A theory of gradient analysis. *Advances in Ecological Research* 18, 271–317.
- Terbraak, C.J.F., Šmilauer, P., 2002. CANOCO reference manual and CanoDraw for Windows user's guide: software for canonical community ordination (ver. 4.5). Microcomputer Power, Ithaca, NY, USA.
- Thiel, V., Peckmann, J., Richnow, H.H., Luth, U., Reitner, J., Michaelis, W., 2001. Molecular signals for anaerobic methane oxidation in Black Sea seep carbonates and a microbial mat. *Marine Chemistry* 73 (2), 97–112.
- Trimble, S.W., Mendel, A.C., 1995. The cow as a geomorphic agent — A critical review. *Geomorphology* 13 (1–4), 233–253.
- Van Oost, K., Quine, T.A., Govers, G., De Gryze, S., Six, J., Harden, J.W., Ritchie, J.C., McCarty, G.W., Heckrath, G., Kosmas, C., Giraldez, J.V., da Silva, J.R.M., Merckx, R., 2007. The impact of agricultural soil erosion on the global carbon cycle. *Science* 318, 626–629.
- Walthert, L., Graf, U., Kammer, A., Luster, J., Pezzotta, D., Zimmermann, S., Hagedorn, F., 2010. Determination of organic and inorganic carbon, delta C-13, and nitrogen in soils containing carbonates after acid fumigation with HCl. *Journal of Plant Nutrition and Soil Science* 173 (2), 207–216.
- Warren, S.D., Thurow, T.L., Blackburn, W.H., Garza, N.E., 1986. The influence of livestock trampling under intensive rotation grazing on soil hydrologic characteristics. *Journal of Range Management* 39 (6), 491–495.
- Wu, R.G., Tiessen, H., 2002. Effect of land use on soil degradation in alpine grassland soil, China. *Journal of the Soil Science Society of America* 66 (5), 1648–1655.
- Zelles, L., 1997. Phospholipid fatty acid profiles in selected members of soil microbial communities. *Chemosphere* 35 (1–2), 275–294.
- Zelles, L., 1999. Fatty acid patterns of phospholipids and lipopolysaccharides in the characterisation of microbial communities in soil: a review. *Biology and Fertility of Soils* 29 (2), 111–129.
- Zhou, Z.C., Gan, Z.T., Shangguan, Z.P., Dong, Z.B., 2010. Effects of grazing on soil physical properties and soil erodibility in semiarid grassland of the Northern Loess Plateau (China). *Catena* 82 (2), 87–91.

---

## Paper II

# Increasing soil methane sink along a 120-year afforestation chronosequence is driven by soil moisture

David Hiltbrunner<sup>1,3\*</sup>, Stephan Zimmermann<sup>1</sup>, Saeed Karbin<sup>2</sup>, Frank Hagedorn<sup>1</sup> and Pascal A. Niklaus<sup>2</sup>

<sup>1</sup>) Swiss Federal Institute of Forest, Snow and Landscape Research (WSL), CH-8903 Birmensdorf, Switzerland

<sup>2</sup>) Institute of Evolutionary Biology and Environmental Studies, University of Zürich, CH-8057 Zürich, Switzerland

<sup>3</sup>) Department of Geography, University of Zürich, CH-8057 Zürich, Switzerland

\*Corresponding author. Tel: +41 44 7392 490, fax: +41 44 7392 215, *E-mail address*: [david.hiltbrunner@wsl.ch](mailto:david.hiltbrunner@wsl.ch) (D. Hiltbrunner)

Accepted in Global Change Biology: DOI: 10.1111/j.1365-2486.2012.02798.x

---

## Abstract

Upland soils are important sinks for atmospheric methane ( $\text{CH}_4$ ), a process essentially driven by methanotrophic bacteria. Soil  $\text{CH}_4$  uptake often depends on land use, with afforestation generally increasing the soil  $\text{CH}_4$  sink. However, the mechanisms driving these changes are not well understood to date.

We measured soil  $\text{CH}_4$  and  $\text{N}_2\text{O}$  fluxes along an afforestation chronosequence with Norway spruce (*Picea abies* L.) established on an extensively grazed subalpine pasture. Our experimental design included forest stands with ages ranging from 25 to >120 years and included a factorial cattle urine addition treatment to test for the sensitivity of soil  $\text{CH}_4$  uptake to N application. Mean  $\text{CH}_4$  uptake significantly increased with stand age on all sampling dates. In contrast,  $\text{CH}_4$  oxidation by sieved soils incubated in the laboratory did not show a similar age-dependency.

Soil  $\text{CH}_4$  uptake was unrelated to soil N status (but cattle urine additions stimulated  $\text{N}_2\text{O}$  emission). Our data indicated that soil  $\text{CH}_4$  uptake in older forest stands was driven by reduced soil water content, which resulted in a facilitated diffusion of atmospheric  $\text{CH}_4$  into soils. The lower soil moisture likely resulted from increased interception and/or evapotranspiration in the older forest stands. This mechanism contrasts alternative explanations focusing on nitrogen dynamics or the composition of methanotrophic communities, although these factors also might be at play. Our findings further imply that the current dramatic increase in forested area increases  $\text{CH}_4$  uptake in alpine regions.

Keywords: methane oxidation, nitrous oxide, afforestation, chronosequence, Norway spruce, alpine regions, soil moisture regime, fertilization

---

## Introduction

Methane (CH<sub>4</sub>) is produced in water-logged soils by methanogenic archaea (Boone *et al.*, 1993). In contrast, well-aerated upland soils are the most important biological sink for atmospheric CH<sub>4</sub> (IPCC, 2007). Soil CH<sub>4</sub> uptake is essentially driven by the oxidation of CH<sub>4</sub> by soil methanotrophic bacteria. In many soils, both processes – methanogenesis and CH<sub>4</sub> oxidation – take place concurrently, with the soil acting as a net source or sink, depending on which process dominates.

The largest terrestrial sinks for atmospheric CH<sub>4</sub> are generally found in forest soils. When forests are converted into grassland or arable fields, soil CH<sub>4</sub> uptake generally decreases (Hütsch *et al.*, 1994, Smith *et al.*, 2000, Willison *et al.*, 1995). Many investigations have attributed this decrease in methanotrophic activity to the disturbance of soil physical structure associated with such land-use changes, and to the application of mineral nitrogen fertilizers. Physical disturbances of the soils through ploughing disrupts aggregates, which might affect the ecological niche of methanotrophs (Boeckx & Van Cleemput, 2001), especially in coarse-textured soils (Hütsch, 1998). The use of heavy machinery on cultivated land also compacts soils, thereby restricting diffusive transport of atmospheric CH<sub>4</sub> into soils (Ball *et al.*, 1997b, Smith *et al.*, 2003). Fertilization of agricultural fields, in particular with ammonium-based fertilizers, has been shown to inhibit CH<sub>4</sub> oxidation (Gulledge *et al.*, 1997, Jassal *et al.*, 2011, King & Schnell, 1994, Whalen, 2000); however, positive effects of N fertilization also have been reported (Bodelier & Laanbroek, 2004).

Interestingly, when cultivated land is abandoned, CH<sub>4</sub> oxidation reverts only very slowly to pre-cultivation levels. Paired-site studies have demonstrated that this process can take many years (Priemé *et al.*, 1997, Smith *et al.*, 2000) but it is not well understood to date why the increase in soil CH<sub>4</sub> uptake is so slow. One factor involved might be the very low growth rates of methanotrophic bacteria thriving on atmospheric CH<sub>4</sub> (King, 1997, Menyailo *et al.*, 2008, Priemé *et al.*, 1996). Another reason may be that the original soil structure is restored only after many years (Hütsch, 1998, Priemé *et al.*, 1997, Regina *et al.*, 2007, Smith *et al.*, 2000). However, not many studies on the recovery of the soil CH<sub>4</sub> sink are available, presumably because not many such chronosequences have been established.

Across Europe and N-America, large areas of land have been abandoned for socio-economic reasons. In the European mountains, woody plant encroachment in abandoned grasslands is widespread (FAO, 2001). In Switzerland, the forest cover in the Alps increased by 900 km<sup>2</sup> between 1984 and 2005, which corresponds to a 15% increase in total forested area in this region (Brändli, 2010). Whether and to what extent soil CH<sub>4</sub> uptake increases under these

conditions is unclear. Subalpine pastures have generally only moderately been grazed, with little nutrient inputs and they have never been tilled. Thus, the loss of methanotrophic activity when these pastures have been established has probably been smaller than in intensified low-land pastures and arable fields (Christiansen & Gundersen, 2011, Peichl *et al.*, 2010, Priemé *et al.*, 1997). As a consequence, the increase in soil CH<sub>4</sub> uptake after afforestation might also be smaller.

In our study, we have measured soil CH<sub>4</sub> uptake and potential CH<sub>4</sub> oxidation along a chronosequence of Norway spruce afforestations spanning more than 120 years. All forest plots are located in an extensively grazed subalpine pasture. To test the sensitivity of soil CH<sub>4</sub> uptake to nitrogen additions, we further established a N-fertilizer treatment (cattle urine) in all chronosequence plots. Our aims were (i) to test for effects of afforestation on the soil CH<sub>4</sub> sink, focusing in particular on the temporal dynamics of these changes; and (ii) to test how these changes were related to changes in soil physical properties and nitrogen status.

## Materials and Methods

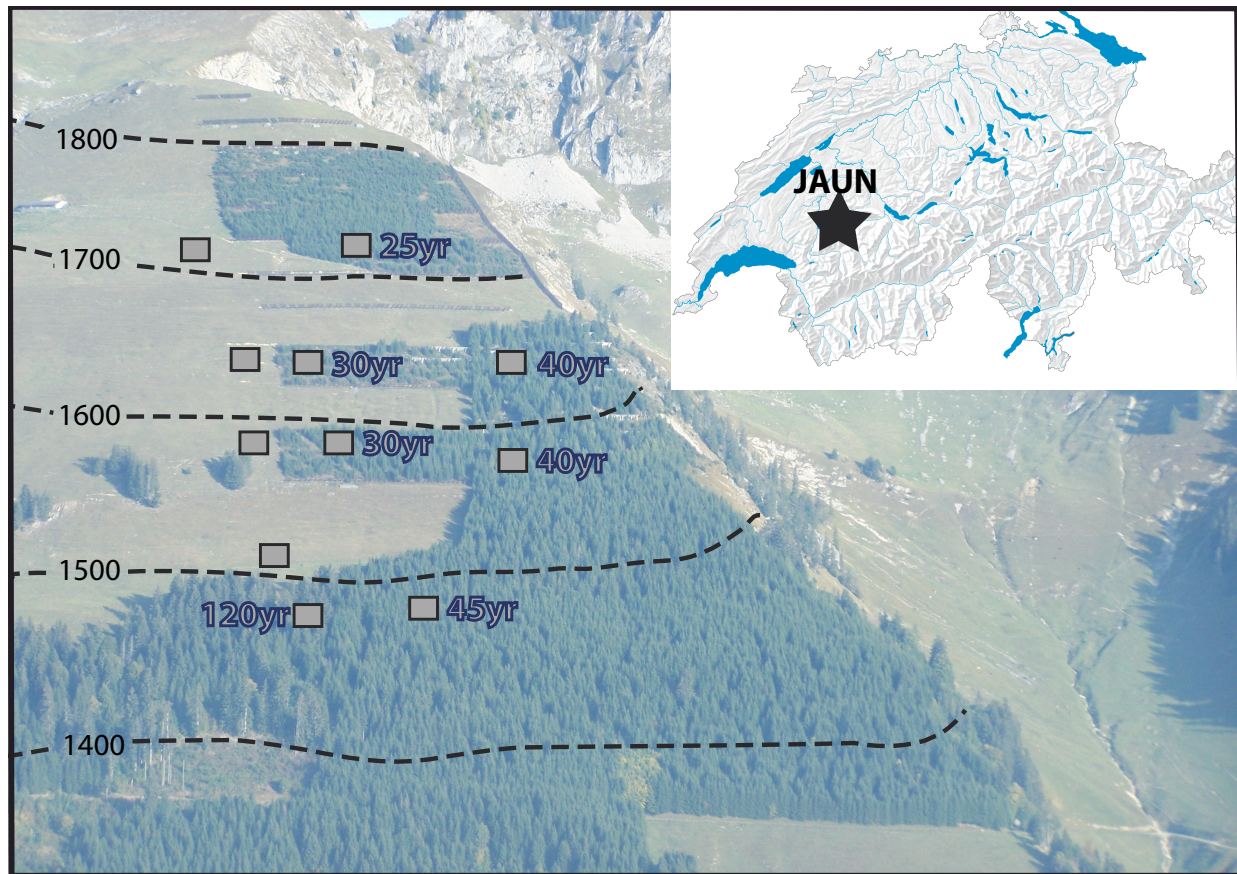
### Study site and experimental design

The present study was conducted in a sub-alpine region in the Canton of Fribourg, Switzerland (7°15'54 E; 46°37'17 N), on a south-facing slope extending from 1450 m a.s.l to 1700 m a.s.l. This slope has been used as pasture for the last 150 years; no land-use records are available prior to this period, but it seems likely that the slope has been under pasture for several centuries. Mean summer and winter air temperatures are 11.4°C and 0.6°C, respectively; mean annual precipitation averages 1250 mm with a maximum in summer. Soils are Cambisols on calcareous bedrock.

After severe avalanches in 1956, an area of about 15 ha on the eastern part of the slope was gradually afforested with Norway spruce (*Picea abies* L.), while the western part remained as a pasture (Fig. 1). Separate patches of forest were planted on different dates, resulting in stands 25, 30, 40, 45, and >120 years old. We established one 15x15m plot in each forest patch, plus an additional four similarly-sized plots in the adjacent pasture. Within each plot, four pairs of subplots were established. One randomly-selected subplot per pair was treated with synthetic cattle urine, while the other one served as unfertilized reference. The synthetic cattle urine was prepared according to Fraser *et al.* (1994) and contained urea as the main N source plus glycine representing the amino acid fraction in the cattle urine, potassium bicarbonate, potassium bromide, potassium chloride and potassium sulphate. The synthetic urine solution was applied to the subplots at a rate of 20 g N m<sup>-2</sup> (as 3.35 L m<sup>-2</sup> aqueous



solution) on August 12, 2010. The same amount of water was added to the unfertilized control plots.



**Fig. 1:** Photograph of the study site showing the plots where the  $\text{CH}_4$  and  $\text{N}_2\text{O}$  fluxes have been measured (grey squares) with the ages of the respective stands.

### Soil-atmosphere $\text{CH}_4$ and $\text{N}_2\text{O}$ fluxes

Soil-atmosphere fluxes of  $\text{CH}_4$  and  $\text{N}_2\text{O}$  were measured using static chambers. On May 17, 2010, a 32 cm diameter  $\times$  30 cm tall static chamber was lowered 20 cm into the soil of each subplot and remained there until the end of the growing season. The chambers were placed at some distance from the tree stems to avoid coarse roots. The remaining headspace volume of each chamber was determined by measuring its height above-ground at several locations within the chamber. Soil-atmosphere trace gas fluxes were determined on July 17, August 10, 13 and 20, September 3, and October 1, 2010, by closing the chamber with a gas-tight lid and sampling the headspace through a septum after 5, 20, and 35 minutes. The headspace samples were injected into pre-evacuated exetainers and analysed for  $\text{CH}_4$  and  $\text{N}_2\text{O}$  concentrations using a gas chromatograph (Agilent 7890 fitted with a flame ionisation ( $\text{CH}_4$ ) and an electron-capture detector ( $\text{N}_2\text{O}$ ), Agilent Technologies Inc., Santa Clara, CA, USA).  $\text{CH}_4$  and  $\text{N}_2\text{O}$  flux

rates were calculated by linear regression of measured concentrations against sampling time. Estimates with regression coefficients  $r^2 < 0.8$  were excluded except when fluxes were close to zero.

Soil surface temperature (0-2 cm) and volumetric water content (0-15 cm) were measured concomitantly with the gas measurement using a thermometer and time domain reflectometry (TDR) probes (TRIME-FM, IMKO, Ettlingen, Germany).

We further measured potential CH<sub>4</sub> oxidation rates of sieved soil under standardized laboratory conditions. On September 28, 2011, two soil cores were taken from each subplot and divided into the 0-5, 5-10, 10-15, and 15-20 cm depth layer of the mineral soil horizon. The soil fractions were sieved (2 mm mesh size), and fresh soil equivalent to 100 g dry weight placed into gas-tight jars fitted with a septum. The soils were equilibrated at 20°C overnight; then, the jars were aerated for 30 min, closed again, and CH<sub>4</sub> oxidation rates determined by measuring headspace CH<sub>4</sub> concentrations after 5, 125 and 425 minutes. These incubations were conducted under atmospheric CH<sub>4</sub> concentrations, i.e. no extra CH<sub>4</sub> was injected into the headspace.

### Soil bulk density and porosity

On November 8, 2011, three soil cylinders of 10.8 cm diameter × 11 cm depth were collected per plot. Bulk soil density was estimated by dividing the mass of the dried soil (105°C) by the volume of the cylinder. Particle density was determined by the pycnometer-method (Blake & Hartge, 1986). Total porosity was calculated as 1-(bulk density/particle density). Soil texture was determined with the pipette method according to Gee and Bauder (1986).

### Soil acidity and mineral N concentrations

Three weeks after the application of synthetic cattle urine, four soil samples (2 cm diameter × 5 cm depth) were collected in each subplot. The soils were sieved, roots removed, and soil pH measured potentiometrically in a dried (60°C) aliquote suspended in 0.01 M CaCl<sub>2</sub> at a soil:extractant ratio of 1:2. Ammonium (NH<sub>4</sub><sup>+</sup>) and nitrate (NO<sub>3</sub><sup>-</sup>) were extracted from 10 g fresh soil with 100 mL 1M KCl in an overhead shaker (1.5h). Extracts were filtered (0790<sup>1/2</sup>, Whatman International, Maidstone, England) and NH<sub>4</sub><sup>+</sup> concentrations were measured colorimetrically by automated flow injection analysis (Perkin Elmer UV/ VIS Spectrometer Lambda 2S, Waltham, MA, USA). Nitrate was determined colorimetrically at 210 nm (Varian Cary 50, Palo Alto CA, USA) as difference in absorbance between non-reduced and reduced (using H<sub>2</sub>SO<sub>4</sub> and copperized zinc) extracts (Navone, 1964).



---

## Potential nitrification and denitrification

Potential nitrification (PN) was determined by the shaken slurry method (Hart *et al.*, 1994). Briefly, 10 g sieved fresh soil was suspended in 90 mL 1mM phosphate buffer adjusted to pH 7.0. Ammonium sulphate (140 mg N kg<sup>-1</sup> soil) was added and the slurry incubated at 25 °C on an orbital shaker. Aliquots of 10 mL were taken after 1, 4, 18 and 22 h. These aliquots were immediately mixed with 15mL 2.5M KCl to stop nitrification, centrifuged, and the supernatant analysed for NO<sub>3</sub><sup>-</sup> as described above. Potential nitrification rates were calculated by linear regression of NO<sub>3</sub><sup>-</sup> concentration against time.

Denitrifying enzyme activity (DEA), which shows the denitrification potential under excess substrate availability, was determined by the application of the acetylene inhibition assay (Patra *et al.*, 2005, Smith & Tiedje, 1979). Fresh sieved soil samples equivalent to 5g dry weight were placed in 125 mL plasma flasks and the headspace replaced by a 90:10 mixture of helium:acetylene. The flasks were incubated at 26°C and, after 1h, an aqueous solution containing KNO<sub>3</sub>, glucose and glutamic acid added. N<sub>2</sub>O concentrations in the headspace were analysed after 60, 90 and 120 min as described above. N<sub>2</sub>O production per unit time (DEA) was estimated by linear regression.

## Tree aboveground biomass

Tree aboveground biomass in each plot was calculated using allometric relations depending on stem diameter at breast height, tree height (Kaufmann, 2001) and basal area per ground area. The diameter of all trees was measured in two areas 25 to 100 m<sup>2</sup> in size in stands up to 30 years old. In the older afforestations, trees were measured in a single large area of 250 to 600 m<sup>2</sup> to account for the bigger size and lower density of trees found there. In addition, the heights of 5-10 single trees per area were measured.

Table 1: Soil properties and above ground tree-biomass in the pasture and the different afforestations with (standard errors) representing the different plots per age.

Land use	pH (CaCl <sub>2</sub> )		Bulk density [g cm <sup>-3</sup> ]		Porosity [%]		Clay content [%]		Tree biomass [t ha <sup>-1</sup> ]	
Pasture	4.9	(0.1)	0.83	(0.04)	65	(1.4)	55	(1)	-	-
Afforestation 25y	4.8	-	0.73	-	69	-	57	-	157	-
Afforestation 30y	4.9	(0.3)	0.79	(0.06)	67	(1.5)	55	(4)	140	(3)
Afforestation 40y	4.2	(0.0)	0.91	(0.05)	63	(1.4)	36	(12)	277	(38)
Afforestation 45y	3.9	-	0.76	-	68	-	51	-	263	-
Afforestation 120y	4.8	-	0.70	-	69	-	49	-	579	-

## Statistical analysis

We analysed our data by fitting mixed-effects models by maximum likelihood (ASReml 3.0, VSN International, UK; Gilmour *et al.*, 2009). The model included the sequential fixed effects altitude (elevation in m a.s.l.), land use (forest vs. meadow), forest stand age, fertilization, and the interactions of fertilization with land use and stand age. The effect of stand age was fitted as a log-linear contrast (1 df, testing for effects of log(age)) followed by a term testing for the deviation from log-linearity (3 df, age fitted as categorical term). The significance of the fixed effects was determined using Wald statistics. Reflecting the structure of the experiment, the model included the nested random effects plot, subplot, and static chamber. Altitude and stand age were partly confounded in our study, with higher average forest stand age at the bottom of the slope, and younger forest patches dominating the top of the slope. We therefore fitted a second model in which the terms for altitude and stand age were interchanged; testing for effects of age after accounting for altitude underestimates the age-effect, while age-effects potentially include an altitude-component when fitted first. Effects with  $P < 0.05$  were considered statistically significant, effects with  $0.05 < P < 0.1$  as marginally significant.

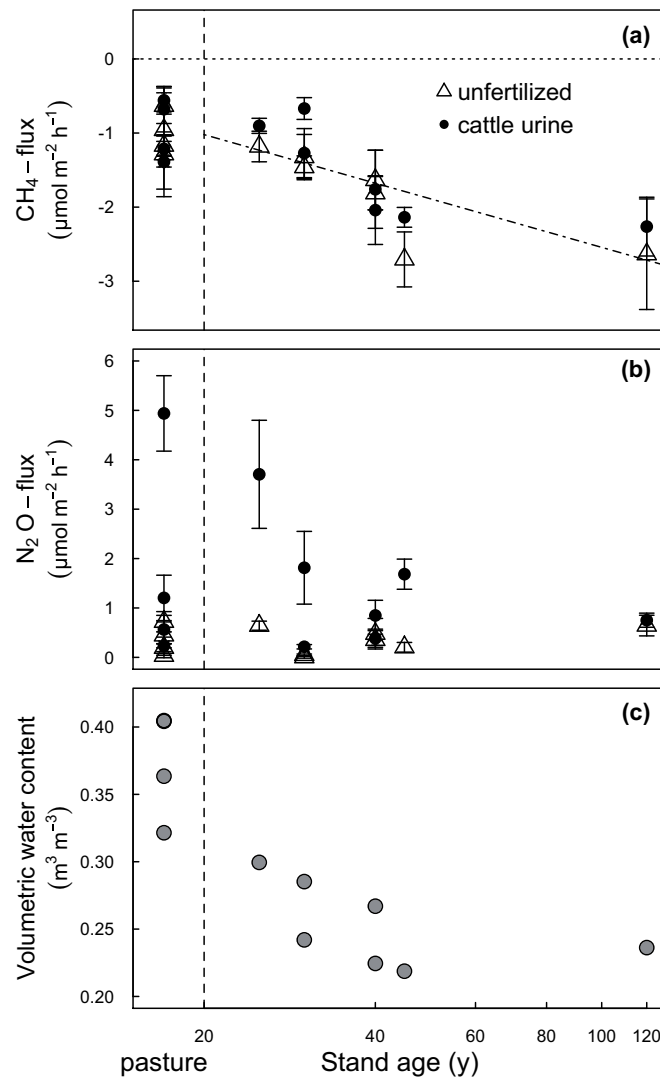
---

## Results

### Soil bulk density, porosity, and water content

Average bulk density of the soils (0-10 cm) showed no consistent trend with land use and stand age. The densities varied between 0.7 and 0.9 g cm<sup>-3</sup>, with the highest values in the 40 years old afforestations (0.91 g cm<sup>-3</sup>) and the lowest ones in the old forest (0.70 g cm<sup>-3</sup>) (Table 1). These findings were confirmed by measuring an additional 65 soil cores sampled across the whole site; these did not show a statistically significant effect of stand age or land use on bulk density (data not shown). In accordance, soil porosities were in a rather narrow range (63-69%) and also did not depend on land use or stand age (Table 1).

Microclimate greatly differed between the two land use types. During the growing season, surface soils of the forest stands were on average 5°C cooler than the pasture soils, which exceeds the temperature lapse rate across 250 m in altitude of 1.5°C. Soil moisture varied within the pasture, but this variation was not related to altitude. In fact, pasture soils at the top and the bottom of the slope had approximately equal soil water contents of 0.40 m<sup>3</sup> m<sup>-3</sup> when averaged over the six sampling dates (Fig. 2c). A general trend, however, was that soil moisture significantly decreased with stand age on all except one date, with variable levels of significance (P<0.05 to P<0.001). The measurement period encompassed a wide range of climatic conditions resulting with rather dry (14-26% volumetric water content; July 17, 2010) and wet soils (24-57%; October 3, 2010). Forest soils were drier than pasture soils on all dates. Reflecting soil moisture, water-filled pore space (WFPS) also decreased along the chronosequence (Fig. 5).

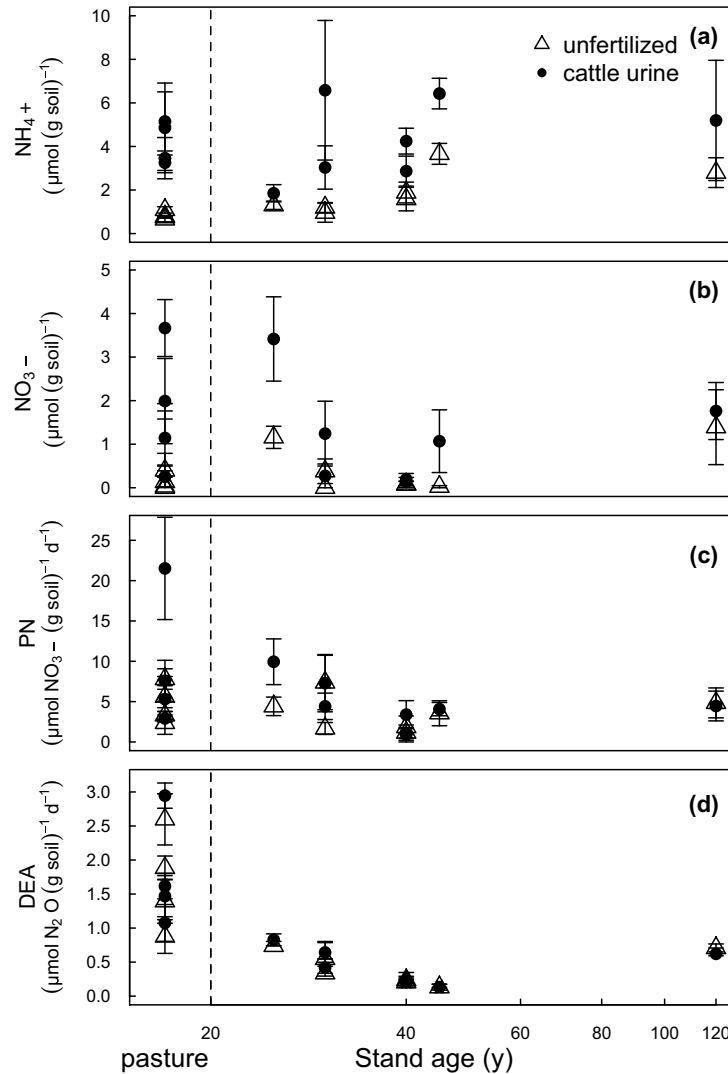


**Fig. 2:** Fluxes of CH<sub>4</sub> (a) and N<sub>2</sub>O (b) in dependence of forest stand age and cattle urine application (negative values indicate uptake from the atmosphere). Error bars are standard errors based on  $n=4$  subplots per forest or pasture plot. Cattle urine addition did not affect volumetric soil water content (c), so that data of fertilized and unfertilized subplots were combined

### Soil methane uptake

Soil CH<sub>4</sub> uptake was higher under forest than under pasture ( $P<0.001$  for effects of land use). Soil CH<sub>4</sub> uptake significantly increased with stand age, with the log-linear component ( $P<0.001$ ) explaining twice as much variance as the term testing for deviations from log-linearity ( $P<0.05$ ). Stand age explained less variance when fitted after accounting for altitude ( $P=0.01$  for  $\log(\text{age})$  and  $P=0.08$  for the deviation from log-linearity). Reflecting the partially confounding influence of age and altitude, the effect of altitude was significant at  $P<0.001$

when fitted before age, but explained ca. 8 times less variance and was at the border to significance ( $P=0.05$ ) when fitted after age.

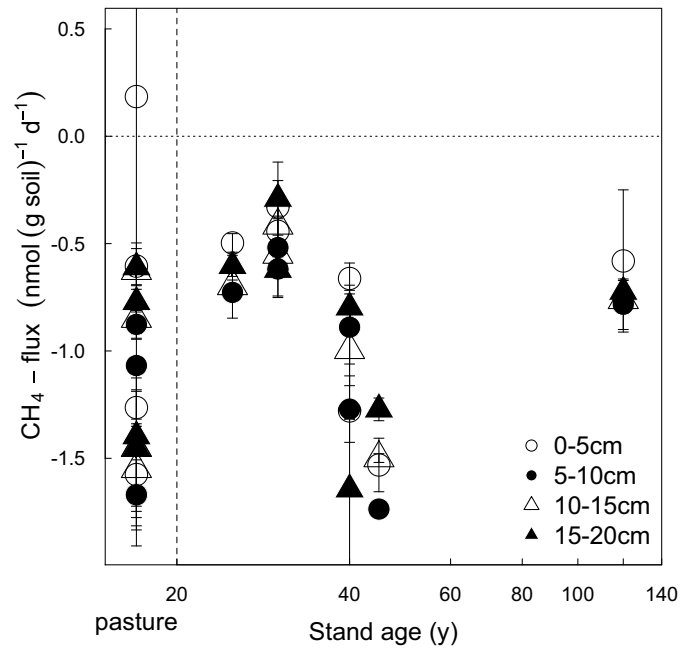


**Fig. 3:** Soil extractable  $\text{NH}_4^+$  (a) and  $\text{NO}_3^-$  (b), potential nitrification rates (PN) (c), and denitrification enzyme activity (DEA) (d) in dependence of forest stand age and cattle urine application. All data refer to the to 5cm of mineral soil. Error bars represent standard errors.

WFPS was significantly negatively related to soil  $\text{CH}_4$  uptake ( $P<0.001$ ; Fig. 5), explaining more than 70% of the variance accounted for by the fixed effects contained in the model. Effects of  $\log(\text{age})$  explained only half as much variance and were less significant ( $P<0.05$ ) when fitted after WFPS, suggesting that at least part of the observed age-effect was due to altered soil moisture. Soil  $\text{NH}_4^+$  and  $\text{NO}_3^-$  (fitted as  $\log([\text{NH}_4^+])$  and  $\sqrt{[\text{NO}_3^-]}$ ) did not explain significant fractions of the variation in  $\text{CH}_4$  fluxes.

Cattle urine addition exerted only little effect on CH<sub>4</sub> oxidation; when data for the different sampling dates were tested individually, a decrease in soil CH<sub>4</sub> uptake of 20% was found one day after fertiliser addition ( $P=0.02$ ). Averaged over all sampling dates, effects of cattle urine addition were no longer statistically significant (-11%, n.s.).

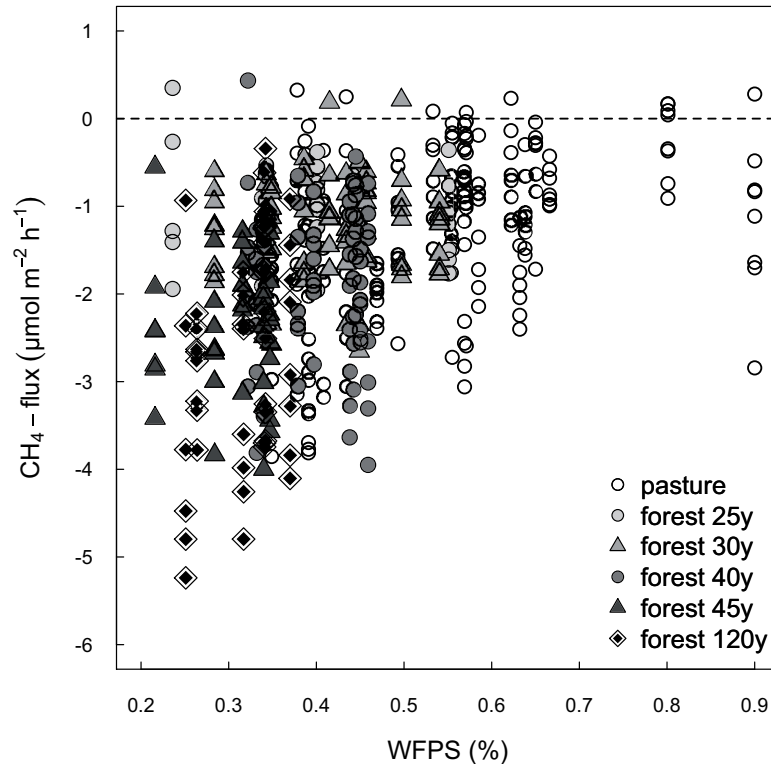
Interestingly, the CH<sub>4</sub> uptake of sieved soils incubated in the laboratory did not reveal any systematic effect of age (Fig. 4), but effects of soil moisture remained significant at  $P<0.001$ .



**Fig. 4:** CH<sub>4</sub> oxidation of the sieved soils in the laboratory in dependence of land use and stand age at different depths. negative values indicate uptake from the atmosphere. Error bars represent standard errors of the means.

### Soil N<sub>2</sub>O emissions and N cycling

In the absence of cattle urine, N<sub>2</sub>O fluxes did not change with age. Cattle urine increased N<sub>2</sub>O emissions from soils in the younger forest stands. This resulted in a significant overall effects of  $\log(\text{age})$  ( $P<0.001$ ) and a significant interaction between  $\log(\text{age})$  and cattle urine application ( $P<0.05$ , respectively, Fig. 2b). Soil extractable NH<sub>4</sub><sup>+</sup> increased with stand age ( $P<0.05$ ), while NO<sub>3</sub><sup>-</sup> did not show such an effect. Potential nitrification did not depend on stand age but increased with cattle urine addition ( $P<0.01$ ). Denitrification enzyme activity did not respond either to stand age or to cattle urine addition. Soil acidity generally decreased with stand age, but the oldest stand had pH similar to the youngest stand; this resulted in no effect of  $\log(\text{age})$ , but a significant deviation from linearity ( $P<0.001$ ).



**Fig. 5:** Soil  $\text{CH}_4$  fluxes in dependence of water filled pore space (WFPS) and stand age (negative values indicate uptake from the atmosphere).

## Discussion

Our results show that  $\text{CH}_4$  oxidation in subalpine soils increased by a factor of two to three after conversion from pasture to forest.  $\text{CH}_4$  oxidation increased with stand age on all sampling dates, spanning a wide range of climatic conditions, emphasizing that this effect is robust.

In contrast to  $\text{CH}_4$  fluxes,  $\text{N}_2\text{O}$  emissions showed no similar change with stand age, at least as long as no cattle urine was added (Fig. 2b). The main driver of  $\text{N}_2\text{O}$  emissions was the mineral N status of the soils, but particularly concentrations of  $\text{NO}_3^-$  were not related to stand age. The primary objective of the  $\text{N}_2\text{O}$  flux measurements was not to assess  $\text{N}_2\text{O}$  fluxes in detail (which would require far more measurements), but to obtain an indicator of the ecosystem's N status and its dependency on age and fertilizer application.

In the 120 year-old old subalpine forest, soil  $\text{CH}_4$  oxidation had reached rates comparable to the range published for temperate coniferous forests (Jang *et al.*, 2006, Peichl *et al.*, 2010, Smith *et al.*, 2000), although much higher rates have been reported for some forests (Ishizuka

*et al.*, 2000, Price *et al.*, 2004). However, it remains unclear whether further increases in CH<sub>4</sub> oxidation can be expected in the future when stand age exceeds 120 years.

Stand age and altitude were not orthogonal in our study. The oldest and the 45 years old forest were at the lower end of the slope (1450m a.s.l.), while the youngest (25 years old) stand was at the upper end at 1700m a.s.l. CH<sub>4</sub> uptake is relatively insensitive to temperature (Smith *et al.*, 2003), and therefore not likely to be affected by the relatively short altitudinal gradient of 250m. Nevertheless, vegetation period, plant growth and the biological activity might be higher at the lower end of the slope and effects of altitude could therefore be confounded with effects of stand age. We argue, however, that this is unlikely in our study, for several reasons. First, and most importantly, the effect of stand age remained statistically significant after adjusting for altitude. Second, the effect of altitude was not statistically significant in the reference grassland ( $P=0.09$ ), although there was a slight trend towards increased CH<sub>4</sub> oxidation rates at lower elevation. Third, if the effect of altitude was to increase productivity and the length of the growing season, then one might argue that the forest stands at the bottom of the slope are even older on a biomass or “degree-days” scale. In this case, altitude would increase the “effective age” of the older stands more than the one of the younger stands, and thus alter the conclusions. To test this hypothesis, we analysed CH<sub>4</sub> oxidation as function of stand biomass (which is a proxy for biomass), and obtained similar results ( $P<0.001$ ).

Stand-age effects found in the few afforestation studies available to date are ambiguous. While the majority of the studies observed a slow increase of CH<sub>4</sub> uptake after tree establishment (McNamara *et al.*, 2008, Peichl *et al.*, 2010, Priemé *et al.*, 1997, Singh *et al.*, 2007), some found no age effect (Ball *et al.*, 2007), or an age-effect which depended on tree species (Christiansen & Gundersen, 2011). However, the mechanisms driving this change remain uncertain. One possibility might be that the populations of methanotrophic bacteria require decades to increase due to slow growth rates under atmospheric CH<sub>4</sub> concentrations (King, 1997, Menyailo *et al.*, 2008, Priemé *et al.*, 1996). In addition, the change from herbaceous to tree cover may induce shifts in methanotrophic community structure. In New Zealand, Singh *et al.* (2009) related higher soil CH<sub>4</sub> uptake in pine afforestation compared to pastures to a higher activity of type II methanotrophs which are thought to oxidize atmospheric CH<sub>4</sub> in soils (Knief *et al.*, 2006). We did not measure methanotrophic community structure; however, the systematic effect of stand age was lost when CH<sub>4</sub> oxidation rates were measured on sieved soils, i.e. when diffusive limitations by soil horizons were eliminated. This suggests that the potential to oxidize CH<sub>4</sub> was similar at all sites, independent of age, although our comparison of incubation and field measurements clearly



has some limitations. In particular, laboratory experiments only reflect the oxidation potential of the incubated soil layer, excluding processes lower in the soil column, e.g. methanogenesis. Discrepancies between laboratory incubations and *in situ* measurements were also reported e.g. by (Reay *et al.*, 2005), who measured considerable CH<sub>4</sub> uptake in sieved grassland soils incubated in the laboratory while the same soils were net sources of CH<sub>4</sub> under field conditions (Nedwell *et al.*, 2003). We argue that different diffusive limitations are the most likely explanation for the discrepancy between CH<sub>4</sub> uptake in laboratory incubations and *in situ*. Soil gas diffusivity is controlled by pore network structure and water-filled pore space. In our study, CH<sub>4</sub> uptake decreased with water-filled pores space, a phenomenon commonly found (Ball *et al.*, 1997a, Bowden *et al.*, 1998, Dörr *et al.*, 1993). Soil bulk density and porosity varied only little (60%-70%) among plots, with no systematic effect of stand age. Soil bulk density was fairly low even in the pasture plots, mainly because the cattle moved on specific tracks, leaving the major part of the pastures unaffected by trampling (Hiltbrunner *et al.*, 2012).

Why did soil moisture decrease with forest stand age? Evapotranspiration and interception often increase with forest age (Farley *et al.*, 2005). Moreover, the organic layer under spruce trees shelters the underlying mineral soil from rainfall (Borken & Beese, 2006). In our study, a gradual accumulation of the organic layer, reaching a thickness of 4 to 10 cm in the two oldest stands, might have contributed to drier mineral soils in the older forest plots. In conjunction with increased water retention by the forest stand, this might have promoted CH<sub>4</sub> diffusion into soils, which in turn enhanced CH<sub>4</sub> uptake with forest development.

High NH<sub>4</sub><sup>+</sup> concentrations can inhibit soil CH<sub>4</sub> oxidation in many ecosystems (Gulledge *et al.*, 1997, King & Schnell, 1994, Le Mer & Roger, 2001, Smith *et al.*, 2000, Steudler *et al.*, 1989). In our study, soil extractable NH<sub>4</sub><sup>+</sup> increased with stand age, with no evidence of an inhibition of CH<sub>4</sub> uptake. Similarly, Tate *et al.* (2007) also did not find a significant relationship between extractable NH<sub>4</sub><sup>+</sup> and soil CH<sub>4</sub> oxidation in a land-use change study in New Zealand, despite relatively high soil NH<sub>4</sub><sup>+</sup> concentrations. The cattle urine application in our study also did not substantially suppress soil CH<sub>4</sub> uptake, despite relatively large amounts added and resulting in increased contents of extractable NH<sub>4</sub><sup>+</sup> and increased associated N<sub>2</sub>O emissions in the following two months. However, the fertilization effects were largest in the younger stands, raising the possibility that rapid N uptake by more N-limited old forest stands and their soils protected methanotrophs against effects of NH<sub>4</sub><sup>+</sup>.

CH<sub>4</sub> oxidation often decreases with soils acidification, either due to direct effects of soil pH, or due to reduced nitrification rates and therefore increased soil NH<sub>4</sub><sup>+</sup> concentrations (Stiehl-

Braun *et al.*, 2011, Weslien *et al.*, 2009). Although soil pH differed between forest plots in our study, these changes did not explain the patterns observed in soil CH<sub>4</sub> uptake. Furthermore, soil pH changes spanned only one single pH unit.

Currently, forest cover is increasing rapidly in the European Alps. Our data can be combined with estimates of land use change to arrive at an educated guess of the order of magnitude by which soil CH<sub>4</sub> uptake may increase as consequence of land abandonment. We base our calculation on Switzerland, but expect similar changes in other European alpine areas. Forest cover increased by more than 90 000 ha between 1984 and 2005 in the Swiss Alps (Brändli, 2010), which is equivalent to as much as 8% per decade. The Swiss alpine forests are dominated by conifers, covering 75-85% of the total forested area, with Norway spruce being by far the most abundant species (Brändli, 2010). We assume that (1) the investigated forest stands are reasonably representative of the new forest area, (2) our flux measurements are a good estimate of soil CH<sub>4</sub> uptake for the snow-free period (May to October), and (3) the difference in soil CH<sub>4</sub> uptake between pasture and 45 years old stands reflects the anticipated changes ( $\Delta = 1.0$  to  $1.5 \mu\text{mol CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ ). Combining these data yields an increase in soil CH<sub>4</sub> uptake in the order of  $\sim 0.5 - 0.8 \text{ kg CH}_4\text{-C ha}^{-1} \text{ yr}^{-1}$ , or  $\sim 50\text{-}70 \text{ t CH}_4\text{-C}$  for the entire 90'000 ha area. Soil CH<sub>4</sub> uptake has been estimated at  $\sim 6000 \text{ t CH}_4\text{-C yr}^{-1}$  for Switzerland (Minonzio *et al.*, 1998). However, this figure is associated with a large uncertainty (minimum  $\sim 1000 \text{ t}$ , maximum  $\sim 18000 \text{ t CH}_4 \text{ yr}^{-1}$ ) mainly due to a lack of data for forest soil CH<sub>4</sub> uptake. Our data thus suggest that the ongoing forest expansion in alpine areas increases the Swiss soil CH<sub>4</sub> sink by up to a few percent per decade.

In summary, our study shows increases in soil CH<sub>4</sub> uptake by a factor of two to three after conversion from subalpine pasture to forest. Our data indicates that the most likely reason for this change were shifts in the soil moisture balance due to increased interception and higher evapotranspiration in older forest stands. As a consequence, water-filled pore space decreased and the diffusion of atmospheric CH<sub>4</sub> into soils was facilitated. This mechanism contrasts alternative mechanisms suggested, including altered soil N status, altered soil structure, or shifts in the methanotrophic community structure (Christiansen & Gundersen, 2011, Priemé *et al.*, 1997, Singh *et al.*, 2007).

## Acknowledgements

We gratefully thank R. Köchli, S. Fuchs, and O. Schramm for field assistance and G.D. Lieberherr and D. Christen for assistance in the laboratory. We also thank four anonymous reviewers for their comments, which helped to improve this manuscript.

---

This study was funded by the COST Action 639 (BurnOut) and the Swiss Federal Office for the Environment (FOEN).

## References

- Ball BC, Dobbie KE, Parker JP, Smith KA (1997a) The influence of gas transport and porosity on methane oxidation in soils. *Journal of Geophysical Research-Atmospheres*, **102**, 23301-23308.
- Ball BC, Smith KA, Klemetsson L et al. (1997b) The influence of soil gas transport properties on methane oxidation in a selection of northern European soils. *Journal of Geophysical Research-Atmospheres*, **102**, 23309-23317.
- Ball T, Smith KA, Moncrieff JB (2007) Effect of stand age on greenhouse gas fluxes from a Sitka spruce (*Picea sitchensis* (Bong.) Carr.) chronosequence on a peaty gley soil. *Global Change Biology*, **13**, 2128-2142.
- Blake GR, Hartge KH (1986) Particle Density. *Methods of Soil Analysis, Part I*, In: Klute, A. (ed.): *Physical and Mineralogical Methods*, Amer. Soc. Agron. and Soil Sci. Soc. Amer., Madison (Wisconsin), USA, 363-375.
- Bodelier PLE, Laanbroek HJ (2004) Nitrogen as a regulatory factor of methane oxidation in soils and sediments. *Fems Microbiology Ecology*, **47**, 265-277.
- Boeckx P, Van Cleemput O (2001) Estimates of N<sub>2</sub>O and CH<sub>4</sub> fluxes from agricultural lands in various regions in Europe. *Nutrient Cycling in Agroecosystems*, **60**, 35-47.
- Boone DR, Whitman WB, Rouviere P (1993) Diversity and taxonomy of methanogens.
- Borken W, Beese F (2006) Methane and nitrous oxide fluxes of soils in pure and mixed stands of European beech and Norway spruce. *European Journal of Soil Science*, **57**, 617-625.
- Bowden RD, Newkirk KM, Rullo GM (1998) Carbon dioxide and methane fluxes by a forest soil under laboratory-controlled moisture and temperature conditions. *Soil Biology & Biochemistry*, **30**, 1591-1597.
- Brändli U-B (2010) Schweizerisches Landesforstinventar. Ergebnisse der dritten Erhebung 2004-2006. Eidgenössische Forschungsanstalt für Wald, Schnee und Landschaft WSL, Birmensdorf, Bundesamt für Umwelt, BAFU. Bern
- Christiansen JR, Gundersen P (2011) Stand age and tree species affect N<sub>2</sub>O and CH<sub>4</sub> exchange from afforested soils. *Biogeosciences*, **8**, 2535-2546.
- Dörr H, Katruff L, Levin I (1993) Soil texture parameterization of the methane uptake in aerated soils. *Chemosphere*, **26**, 697-713.
- Farley KA, Jobbagy EG, Jackson RB (2005) Effects of afforestation on water yield: a global synthesis with implications for policy. *Global Change Biology*, **11**, 1565-1576.

- 
- Fraser PM, Cameron KC, Sherlock RR (1994) Lysimeter study of the fate of nitrogen in animal urine returns to irrigated pasture. *European Journal of Soil Science*, **45**, 439-447.
- Gee GW, Bauder JW (1986) Particle size analysis. In: Klute, A. (ed.), *Methods of Soil Analysis. Part 1. Physical and Mineralogical Methods* 386-411, Am. Soc. Agron., Madison, WI.
- Gilmour AR, Gogel BJ, Cullis BR, Thompson R (2009) *ASReml User Guide Release 3.0* VSN International Ltd, Hemel Hempstead, HP1 1ES, UK.
- Gulledge J, Doyle AP, Schimel JP (1997) Different  $\text{NH}_4^+$ -inhibition patterns of soil  $\text{CH}_4$  consumption: A result of distinct  $\text{CH}_4$ -oxidizer populations across sites? *Soil Biology & Biochemistry*, **29**, 13-21.
- Hart SC, Stark JM, Davidson EA, Firestone MK (1994) Nitrogen mineralisation, immobilisation and nitrification. In: Weaver, R. W. et al., (Ed.), *Methods of Soil Analysis, Part 2. Microbiological and Biochemical Properties-SSSA, Vol. Part 2.*, Madison, WI, USA.
- Hiltbrunner D, Schulze S, Hagedorn F, Schmidt MWI, Zimmermann S (2012) Cattle trampling alters soil properties and changes soil microbial communities in a Swiss sub-alpine pasture. *Geoderma*, **170**, 369-377.
- Hütsch BW (1998) Methane oxidation in arable soil as inhibited by ammonium, nitrite, and organic manure with respect to soil pH. *Biology and Fertility of Soils*, **28**, 27-35.
- Hütsch BW, Webster CP, Powlson DS (1994) Methane oxidation in soil as affected by land-use, soil-pH and N-fertilization. *Soil Biology & Biochemistry*, **26**, 1613-1622.
- IPCC (2007): *Climate Change 2007: The Physical Science Basis Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, (eds. Solomon, S., D. Qin, M. Manning, Z. Chen, M. Marquis, K.B. Averyt, M. Tignor and H.L. Miller). 996 pp. Cambridge University Press, Cambridge, United Kingdom and New York, USA
- Ishizuka S, Sakata T, Ishizuka K (2000) Methane oxidation in Japanese forest soils. *Soil Biology & Biochemistry*, **32**, 769-777.
- Jang I, Lee S, Hong J-H, Kang H (2006) Methane oxidation rates in forest soils and their controlling variables: a review and a case study in Korea. *Ecological Research*, **21**, 849-854.
- Jassal RS, Black TA, Roy R, Ethier G (2011) Effect of nitrogen fertilization on soil  $\text{CH}_4$  and  $\text{N}_2\text{O}$  fluxes, and soil and bole respiration. *Geoderma*, **162**, 182-186.
- Kaufmann E (2001) Estimation of standing timber, growth and cut. In: P. Brassel and H. Lischke (Ed.), *Swiss National Forest Inventory: Methods and Models of the Second Assessment*. Swiss Federal Research Institute WSL, Birmensdorf, S. 162-196.
- King GM (1997) Responses of atmospheric methane consumption by soils to global climate change. *Global Change Biology*, **3**, 351-362.
- King GM, Schnell S (1994) Ammonium and nitrite inhibition of methane oxidation by *methylobacter-albus* BG8 and *methylosinus-trichosporium* OB3b at low methane concentrations. *Applied and Environmental Microbiology*, **60**, 3508-3513.

- 
- Knief C, Kolb S, Bodelier PLE, Lipski A, Dunfield PF (2006) The active methanotrophic community in hydromorphic soils changes in response to changing methane concentration. *Environmental Microbiology*, **8**, 321-333.
- Le Mer J, Roger P (2001) Production, oxidation, emission and consumption of methane by soils: A review. *European Journal of Soil Biology*, **37**, 25-50.
- McNamara NP, Black HJJ, Pearce TG, Reay DS, Ineson P (2008) The influence of afforestation and tree species on soil methane fluxes from shallow organic soils at the UK Gisburn Forest Experiment. *Soil Use and Management*, **24**, 1-7.
- Menyailo OV, Hungate BA, Abraham W-R, Conrad R (2008) Changing land use reduces soil CH<sub>4</sub> uptake by altering biomass and activity but not composition of high-affinity methanotrophs. *Global Change Biology*, **14**, 2405-2419.
- Minonzio G, Grub A, Fuhrer J (1998) Methan-Emissionen der schweizerischen Landwirtschaft. *Schriftenreihe Umwelt Nr. 298*. Bundesamt für Umwelt, Wald und Landschaft (BUWAL), Bern.
- Navone R (1964) Proposed method for nitrate in potable waters. *J. Am. Wat. Works Ass.*, **56**, 781-783.
- Nedwell DB, Murrell JC, Ineson P, Reay DS, Radajewski S, McNamara N, Morris S (2003) Microbiological basis of land use impact on the soil methane sink: Molecular and functional analysis. In: *Genes in the environment*. (eds Hails RS, Beringer JE, Godfray HC J) pp 15-166, Blackwell, London
- Patra AK, Abbadie L, Clays-Josserand A et al. (2005) Effects of grazing on microbial functional groups involved in soil N dynamics. *Ecological Monographs*, **75**, 65-80.
- Peichl M, Arain MA, Ullah S, Moore TR (2010) Carbon dioxide, methane, and nitrous oxide exchanges in an age-sequence of temperate pine forests. *Global Change Biology*, **16**, 2198-2212.
- Price SJ, Sherlock RR, Kelliher FM, McSeveny TM, Tate KR, Condon LM (2004) Pristine New Zealand forest soil is a strong methane sink. *Global Change Biology*, **10**, 16-26.
- Priemé A, Christensen S, Dobbie KE, Smith KA (1997) Slow increase in rate of methane oxidation in soils with time following land use change from arable agriculture to woodland. *Soil Biology & Biochemistry*, **29**, 1269-1273.
- Priemé A, Sitaula JIB, Klemedtsson AK, Bakken LR (1996) Extraction of methane-oxidizing bacteria from soil particles. *Fems Microbiology Ecology*, **21**, 59-68.
- Reay DS, Nedwell DB, McNamara N, Ineson P (2005) Effect of tree species on methane and ammonium oxidation capacity in forest soils. *Soil Biology & Biochemistry*, **37**, 719-730.
- Regina K, Pihlatie M, Esala M, Alakukku L (2007) Methane fluxes on boreal arable soils. *Agriculture Ecosystems & Environment*, **119**, 346-352.

- 
- Singh BK, Tate KR, Kolipaka G, Hedley CB, Macdonald CA, Millard P, Murrell JC (2007) Effect of afforestation and reforestation of pastures on the activity and population dynamics of methanotrophic bacteria. *Applied and Environmental Microbiology*, **73**, 5153-5161.
- Singh BK, Tate KR, Ross DJ et al. (2009) Soil methane oxidation and methanotroph responses to afforestation of pastures with *Pinus radiata* stands. *Soil Biology & Biochemistry*, **41**, 2196-2205.
- Smith KA, Ball T, Conen F, Dobbie KE, Massheder J, Rey A (2003) Exchange of greenhouse gases between soil and atmosphere: interactions of soil physical factors and biological processes. *European Journal of Soil Science*, **54**, 779-791.
- Smith KA, Dobbie KE, Ball BC et al. (2000) Oxidation of atmospheric methane in Northern European soils, comparison with other ecosystems, and uncertainties in the global terrestrial sink. *Global Change Biology*, **6**, 791-803.
- Smith MS, Tiedje JM (1979) Phases of denitrification following oxygen depletion in soil. *Soil Biology & Biochemistry*, **11**, 261-267.
- Steudler PA, Bowden RD, Melillo JM, Aber JD (1989) Influence of nitrogen-fertilization on methane uptake in temperate forest soils. *Nature*, **341**, 314-316.
- Stiehl-Braun PA, Powlson DS, Poulton PR, Nildaus PA (2011) Effects of N fertilizers and liming on the micro-scale distribution of soil methane assimilation in the long-term Park Grass experiment at Rothamsted. *Soil Biology & Biochemistry*, **43**, 1034-1041.
- Tate KR, Ross DJ, Saggarr S, Hedley CB, Dando J, Singh BK, Lambie SM (2007) Methane uptake in soils from *Pinus radiata* plantations, a reverting shrubland and adjacent pastures: Effects of land-use change, and soil texture, water and mineral nitrogen. *Soil Biology & Biochemistry*, **39**, 1437-1449.
- Weslien P, Klemedtsson AK, Borjesson G, Klemedtsson L (2009) Strong pH influence on  $N_2O$  and  $CH_4$  fluxes from forested organic soils. *European Journal of Soil Science*, **60**, 311-320.
- Whalen SC (2000) Influence of N and non-N salts on atmospheric methane oxidation by upland boreal forest and tundra soils. *Biology and Fertility of Soils*, **31**, 279-287.
- Willison TW, Webster CP, Goulding KWT, Powlson DS (1995) Methane oxidation in temperate soils - effects of land-use and the chemical form of nitrogen-fertilizer. *Chemosphere*, **30**, 539-546.

---

## Paper III

# **Afforestation with Norway spruce on a subalpine pasture alters carbon dynamics but only moderately affects soil carbon storage**

David Hiltbrunner<sup>1,2</sup>, Stephan Zimmermann<sup>1</sup>, and Frank Hagedorn<sup>1\*</sup>

<sup>1</sup>) Swiss Federal Institute of Forest, Snow and Landscape Research (WSL), CH-8903 Birmensdorf, Switzerland

<sup>2</sup>) Department of Geography, University of Zürich, CH-8057 Zürich, Switzerland

\*Corresponding author. Tel: +41 44 7392 463, fax: +41 44 7392 215, *E-mail address*: [frank.hagedorn@wsl.ch](mailto:frank.hagedorn@wsl.ch) (F. Hagedorn)

**Submitted to Biogeochemistry: September 2012**

---

## Abstract

In alpine regions, land-use is currently changing with a strong reforestation of abandoned grasslands. Here, we studied the effects of afforestation with Norway spruce (*Picea abies* L.) on an extensively grazed subalpine pasture in Switzerland on SOC cycling and storage. Along a 120-year long chronosequence with spruce stands of 25, 30, 40, 45, and >120 years and adjacent pastures, we measured tree biomass, SOC stocks down to the bedrock, natural  $^{13}\text{C}$  abundances, and litter quality. To unravel controls on SOC cycling, we have monitored microclimatic conditions and quantified SOC decomposability under standardized conditions as well as soil respiration *in situ*.

Stocks of SOC were only moderately affected by the afforestation: in the mineral soil, SOC stocks transiently decreased after tree establishment, reaching a minimum 40 to 45 years after afforestation (-25%) and increased thereafter. Soils of the mature spruce forest stored the largest amount of SOC, 13% more than the pasture soils, mainly due to the accumulation of an organic layer (23 t C ha<sup>-1</sup>). By comparison, C accumulated in the tree biomass exceeded the SOC pool by a factor of three in the old forest.

In contrast to the small impact on C storage, afforestation strongly influenced the composition and quality of the SOM. With increasing stand age,  $\delta^{13}\text{C}$  values of the SOM became consistently more positive, which can be interpreted as a gradual replacement of grass- by spruce derived C. Fine roots of spruce were enriched in  $^{13}\text{C}$ , in lignin and had a higher C/N ratio in comparison to grass roots. As a consequence, SOM quality as indicated by the lower fraction of readily decomposable (labile) C and higher C:N ratios declined after the land-use change. Furthermore, spruce plantation induced a less favorable microclimate with the average soil temperature during the growing season being 5°C lower in the spruce stands than in the pasture. *In situ* soil respiration was approximately 50% lower after the land use conversion, which we primarily attribute to the colder conditions and the lower SOM quality, but also to drier soils and to a decreased fine root biomass (-40%). In summary, afforestation on subalpine pastures only moderately affected SOC storage as compared to the large C sink in tree biomass. In contrast, SOC cycling rates strongly decreased as a result of a less favorable microclimate, a lower C input by roots, and a lower litter quality.



---

## 1. Introduction

Land use and land use change (LUC) are among the most important factors governing the carbon (C) fluxes between the terrestrial biosphere and the atmosphere and thus determining whether an ecosystem is a net source or sink for atmospheric CO<sub>2</sub>. While the tropical deforestation contributes to the rising CO<sub>2</sub> level in the atmosphere, reforestation of abandoned agricultural areas on the Northern hemisphere, by contrast, increases the C sequestration (Houghton, 2003). The net C uptake in these regions primarily results from agriculture extensification in conjunction with land use changes. Particularly in the European Alps, large areas of pasture have been abandoned in the last couple of decades for socio-economic reasons (MacDonald et al., 2000; Tappeiner et al., 2003). As a result, the forest cover strongly expanded due to woody plant encroachment (Hagedorn et al., 2010c; Tasser et al., 2007). In Switzerland, for example, the forested area in the Alps increased by 8% between 1995 and 2006, inducing an accumulation of tree biomass of 15 million m<sup>3</sup> (Brändli, 2010). The strong forest expansion is expected to continue particularly on marginal land of alpine regions due to the declining public support of agriculture and the decreasing attractiveness of farming (Bolliger et al., 2008). As a consequence, C storage in biomass is anticipated to increase significantly on a national scale.

As compared to C storage in biomass, the effects of reforestation on soil organic carbon (SOC) are much more ambiguous. Generally, afforestation is assumed to lead to an initial loss of SOC due to a higher C mineralization associated with soil disturbance at planting (Jandl et al., 2007). Thereafter, C stocks are expected to increase slowly until C input and mineralization equilibrates (Thuille & Schulze, 2006). Whether the soil C stocks achieve the level previous to the land use change is uncertain and depends on different factors like climate, soil properties, tree species, or land use history (Guo and Gifford, 2002; Paul et al., 2002; Post and Kwon, 2000). In alpine regions, with soils having relatively high amounts of labile C (Budge et al., 2011; Leifeld et al., 2009), LUC is likely to have a strong impact on soil C dynamics, but particularly in these regions the database is poor. A recent metadata analysis compiling LUC effects in the temperate zone revealed no clear trend in SOC stock changes in the mineral soils after afforestation of grasslands, which was partly attributed to the high variability among the studies (Poeplau et al., 2011). Moreover, since more than 80% of the studies only included surface soils, effects on subsoil were mostly neglected, which could lead to inaccurate estimates (Harrison et

---

al., 2011). A further constraint in some of the studies is the distribution of chronosequences over rather large areas including a high inherent heterogeneity. On a regional to countrywide scale, land use may have historically been selected according to the soil type, confounding the effects of vegetation type and land use history on soil C storage (Wiesmeier et al., 2012).

While most of the studies mainly focused on the total C stock changes, only a few assessments have addressed the underlying mechanism for the observed changes (Scott et al., 2006; Thuille and Schulze, 2006). On the one hand, input pathways and litter composition considerably differ among plant species, with potential implications on soil organic matter quality and C sequestration (De Deyn et al., 2008). In addition, vegetation type affects soil microorganisms (Berg and Smalla, 2009; Macdonald et al., 2009), which govern the decay of organic matter and thus regulate C mineralization. On the other hand, afforestation can modify microclimatic conditions towards less favorable conditions for organic matter decomposition (Kellman et al., 2007; Smith and Johnson, 2004). Whether afforestation leads to an increase or decrease of the SOC stocks strongly depends on the interaction of these processes and their modification by the LUC.

In the present study, we wanted to investigate (1) how SOC-cycling and storage in relation to tree biomass changes along a 120-year old chronosequence of Norway spruce; and (2) to estimate which mechanisms drive the changes in SOC dynamics; in particular, how the afforestation has affected microclimatic conditions and how the altered input of plant litter has affected SOM quality.

The whole chronosequence including 25, 30, 40, 45, and more than 120-year old forest stands, was established on an extensively grazed subalpine pasture within a rather small area on an even slope, providing homogeneous soil conditions within the whole study site. Here we quantified SOC-stocks and SOM quality in relation to land use and stand age. Furthermore, we measured soil respiration *in situ* and C mineralization under standardized conditions to disentangle controls of C cycling and storage as affected by the vegetation change. We hypothesized that (i) soils would loose C, primarily during the first years after the planting as a result of soil disturbance and a declining C input by roots in the upper mineral soil; that (ii) the C-stocks would re-increase in the older stands mainly due to an accumulation of a forest floor; that (iii) the afforestation would lead to a 'colder' microclimate with less favorable conditions for C mineralization; and that (iv) SOM quality would decline.

---

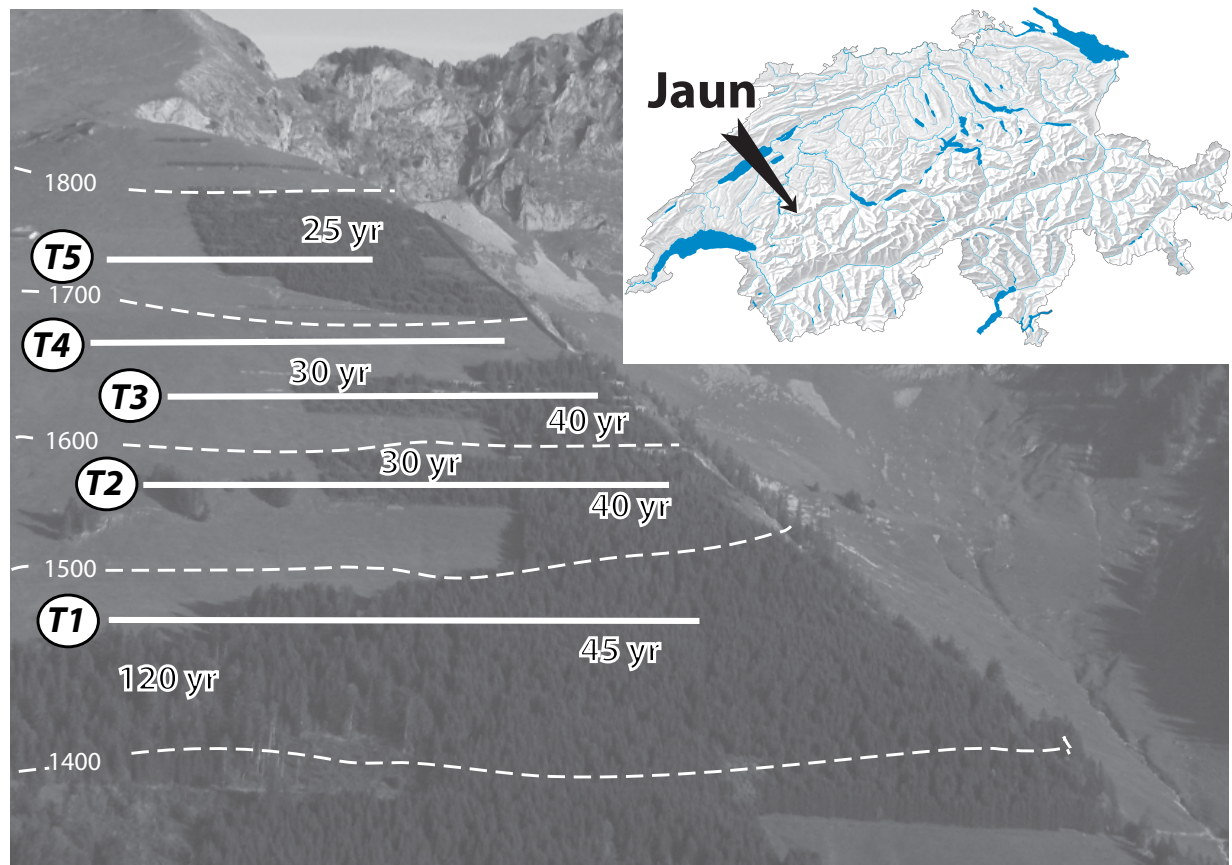
## 2. Materials and Methods

### 2.1 Study site and soil sampling

The study was conducted in a sub-alpine region in the Canton of Fribourg, Switzerland (7°15'54 E; 46°37'17 N), on a south-facing slope reaching from 1450 m a.s.l to 1800 m a.s.l.. Mean summer and winter air temperatures are 11.4°C and 0.6°C, respectively; mean annual precipitation averages 1250 mm with a maximum in summer.

The entire slope has been under pasture for at least 150 years, but most likely it was used for cattle grazing for several centuries. After severe avalanches in 1956, the eastern part of the slope was gradually afforested with Norway spruce (*Picea abies* L.), while the western part remained as a pasture (Fig. 1). Since the trees were planted within a time span of two decades, the afforestations differ in age (25, 30, 40, and 45 years old). In addition, a mature spruce forest (older than 120 years), representing a permanent forest ecosystem, was used as a control (Fig. 1). We selected the stands based on the forest operating plan and determined the precise stand age by dendrochronological analysis. Soils across the whole slope are Eutric Cambisols on calcareous bedrock. While in the pasture an organic layer was lacking, soils of the afforstations had an Oi- and Oe horizon and the mature forest additionally had an Oa horizon. The mineral soil was characterized by an approximately 20cm thick A-horizon with pH values between 4 and 5.5, followed by a B-horizon, partly featuring manganese concretions and iron mottles below 40 cm. The C horizon starts at ~60 cm depth and contained calcareous bedrock material.

Soils were sampled along four transects at different altitudes, ranging from the pasture into the forest stands (n=4 blocks). Each section of the transect differing in land use or stand age was regarded as a plot (n=11) and in each plot, 5 or 10 soil pits were dug with a distance of 10m between each other. A fifth transect at 1680 m a.s.l. with 20 soil pits only within the pasture was used to test the homogeneity of soils along the slope (Fig. 1). In each soil pit, soil morphology and stone content were estimated visually. Soil samples for chemical analyses were taken at six different depths (0-5, 5-10, 10-20, 20-40, 40-60 and 60-80cm). Bulk density was determined at two depths (0-10cm and 20-30cm) in half of the soil pits using steel cylinders with one liter volume. In an additional seven soil pits across the whole site, bulk density was measured down to the bedrock in triplicates. The organic layer was sampled on areal basis with a steel frame (0.25 x 0.25m) for each organic horizon individually (n = 8-20 per plot).



**Fig.1:** Study site with the different transects (T1-T5) along which soils have been sampled. Pasture transect T4 was used to test horizontal homogeneity of the slope

## 2.2 Soil physical analyses, stone content, and root biomass

Bulk soil density was calculated with the mass of the oven dried (105°C) soil weight divided by the volume of the cylinder. From a subset of the soil cores stone content (0-10 cm, 20-30 cm; n=75) and root biomass (0-10 cm; n=28) were determined by consecutively sieving the soils to 2mm (stones), 1 mm and 0.5 mm (roots) under a gentle stream of water. The retained fine rock material and plant roots were thoroughly cleaned from soil particles, dried at 105°C and weighted to calculate fine rock volume and root biomass of the soils. Soil texture was determined with the pipette method according to Gee and Bauder (1986).

**Table 1:** Bulk density, clay content and soil pH in the different plots in 0-10 cm and 20-30 cm depth.

Land use Stand age	Altitude	BD	Clay (%)	pH	BD	Clay (%)	pH
		(kg dm <sup>-3</sup> )			(kg dm <sup>-3</sup> )		
		0 – 10 cm			20 – 30 cm		
Pasture	1450	0.87 (0.05)	53	5.5 (0.2)	1.05 (0.02)	ND	5.3 (0.2)
Pasture	1520	0.83 (0.05)	54	4.8 (0.1)	1.08 (0.03)	52	4.7 (0.1)
Pasture	1610	0.83 (0.03)	50	4.7 (0.1)	1.01 (0.02)	49	4.8 (0.2)
Pasture	1700	0.81 (0.03)	56	5.2 (0.1)	1.11 (0.02)	52	5.8 (0.3)
Forest 25 yr	1700	0.75 (0.02)	61	5.1 (0.2)	1.08 (0.03)	63	5.7 (0.2)
Forest 30 yr	1520	0.81 (0.03)	51	4.8 (0.2)	1.05 (0.04)	ND	5.0 (0.2)
Forest 30 yr	1610	0.84 (0.05)	58	5.7 (0.3)	1.14 (0.03)	ND	6.0 (0.3)
Forest 40 yr	1520	0.83 (0.05)	41	4.3 (0.1)	1.07 (0.04)	41	4.8 (0.1)
Forest 40 yr	1510	0.87 (0.02)	20	4.6 (0.2)	1.17 (0.04)	20	5.4 (0.3)
Forest 45 yr	1450	0.78 (0.02)	50	4.1 (0.1)	1.08 (0.02)	ND	4.9 (0.3)
Forest 120 yr	1450	0.72 (0.03)	49	4.5 (0.2)	1.01 (0.06)	ND	5.0 (0.2)

### 2.3 Tree biomass estimation

Diameter of all trees was measured in two areas of 25 to 100 m<sup>2</sup> in the younger stands (up to 30 years) and in a single larger area of 250 to 600 m<sup>2</sup> in the older afforestations to account for the bigger size and lower density of trees. In addition, the heights of 5-10 single trees per area were measured. Biomass of the single trees was calculated using allometric functions depending on stem diameter at breast height and tree height (Kaufmann, 2001) and total biomass was related to the basal area per ground area.

### 2.4 Chemical analyses of plant and soil samples

Prior to chemical analysis, soil samples were oven-dried (60 °C), sieved (2 mm) and ground. Soil pH was measured potentiometrically in 0.01 M CaCl<sub>2</sub> with a solid/extractant ratio of 1:2.

Since the parent material of the site is calcareous, mineral soil samples with pH >6 were fumigated with HCl in order to remove inorganic C (Walthert et al., 2010). C and N concentrations and the carbon isotopic ratio ( $\delta^{13}\text{C}$ ) of plant and organic layer material and a subset of the mineral soil samples (n = 250) were measured with an automated elemental analyser–continuous flow isotope ratio mass spectrometer (Euro-EA,

---

Hekatech GmbH, Germany, interfaced with a Delta-V Advanced IRMS, Thermo GmbH, Germany). C and N concentrations of the remaining mineral soil samples (n=350) were measured with an elemental analyser coupled with a thermal conductivity detector (NC-2500, CE-Instruments, Italy). Results of the C isotope analysis were expressed in  $\delta$  units (‰) relative to Vienna Pee Dee belemnite (V-PDB) standard.

In plant samples, Klason lignin was gravimetrically determined as the residue of 500mg of finely ground plant material after being consecutively extracted with 12.5ml of hot water (80°C), 12.5ml of ethanol, hydrolysed for 1h with 1.5ml of 72% sulphuric acid at 30°C and after the addition of 42ml of water, autoclaved for 1h at 120°C. Total lignin concentration was the sum of Klason lignin and the soluble lignin, which was estimated from the UV absorbance of the hydrolysate at 205 nm (Dence, 1992).

## **2.5. Microclimate and soil respiration**

Soil temperature and volumetric water content (VWC) was permanently recorded over one and a half year under pasture and in the 40 year old forest at 5, 25, and 50 cm depth using Decagon sensors (5TM) and data loggers (Em50R, Decagon Devices Inc., Pullman, WA, USA). Additional manual measurements were conducted concomitantly with the soil respiration measurements using a thermometer and time domain reflectometry (TDR) probes (TRIME-FM, IMKO, Germany).

Soil respiration was measured using a static chamber technique (Hiltbrunner et al., 2012). Per plot, four PVC-chambers (32cm diam. x 30cm; n=44) were inserted 20 cm into the soil, with the remaining headspace volume of each chamber was determined individually. The chambers were installed two months before the first measurement in order to ensure the recovery of soils from physical disturbance and grass was cut at several dates. Soil respiration rates were determined at six dates (July 17, August 10, 13 and 20, September 3, and October 1, 2010) by closing the chambers with an air-tight lid and sampling the headspace with a syringe through a septum after 5, 20, and 35 min. Gas samples were subsequently analyzed for CO<sub>2</sub> concentrations with a gas chromatograph (Agilent 7890, fitted to a flame ionisation detector (FID), Agilent Technologies Inc. CA, USA) and respiration rates were calculated by linear regression of the CO<sub>2</sub> concentrations against sampling time.

---

## 2.6 Substrate induced respiration (SIR) and C-mineralization

For the incubation experiments, sampled soils from the Oi horizon (n=8), Oe- horizon (n=4) and surface mineral soil (0-5 cm) (n=44) were immediately transported to the laboratory, where they were sieved (4 mm) and the roots were removed. Samples were stored at 4°C for one week prior to the incubation.

SIR, as an indicator for microbial biomass, was measured in a CO<sub>2</sub> measuring system with a continuous CO<sub>2</sub>-free air flow as described by Cheng and Virginia (1993). Briefly, 40 g fresh mineral soil and 10 g organic layer material was placed into airtight jars and amended with 8 ml glucose solution (50g L<sup>-1</sup> for mineral soils and 12.5 g L<sup>-1</sup> for organic layer material, resulting in 10 mg glucose g<sup>-1</sup> sample). CO<sub>2</sub> release of the samples was determined after one hour by measuring CO<sub>2</sub> concentration with an infrared gas analyzer (LI 6252, LI-COR, NE, USA) at an air flow rate of 160 ml min<sup>-1</sup>.

To estimate C mineralization of the soils under standardized conditions, mineral soil samples and organic layer material was incubated during 140 days at 20°C in the dark. 20 g mineral soil and 3 g organic layer material were placed into 120 ml jars and after measuring the respiration rates the first time under field moist conditions, water contents were adjusted to 50% of the water holding capacity. CO<sub>2</sub> production of the samples was measured periodically in a closed system; after tightening the jars with a rubber stopper, air from the headspace circled through an infrared gas analyzer (LI 6252, LI-COR, NE, USA) and respiration rates were calculated by linear regression of the CO<sub>2</sub> concentrations against time.

## 2.7 Calculations and statistical analysis

SOC stocks were calculated by multiplying SOC concentrations with soil bulk densities and thicknesses of each depth class. Bulk densities between 10 and 20 cm were interpolated from the measured values in 0-10 and 20-30 cm depth. In addition, bulk densities were corrected for stone contents according to measurements of the fine rock fraction and visual estimation of the coarse rocks in the soil pits (see results section) assuming a rock density of 2.65.

We analyzed our data by fitting mixed-effects models by maximum likelihood (lme function from the nlme package, R 2.10.1, R Development Core Team (2010)). The model included the sequential fixed effects land use (pasture vs. forest), forest stand age as a linear contrast and soil depth where appropriate. Reflecting the structure of the experiment, the nested random effects were block, plot, and where appropriate, soil depth. The homogeneity of the

slope was tested by applying a linear model (aov function) with the data from the pasture soils only. The variables included were either altitude (m a.s.l.) or position along the horizontal gradient and soil depth. Effects with  $P < 0.05$  were considered statistically significant, effects with  $0.05 < P < 0.1$  as marginally significant.

### 3. Results

#### 3.1 Bulk density, soil texture and soil pH

Soils were rather homogenous across the study site. They had a mean thickness of 80 cm and were carbonate-free to an average depth of 60 cm. Bulk density ranged between 0.7 - 0.9 kg dm<sup>-3</sup> in the surface soil (0-10 cm) and 1.0-1.2 kg dm<sup>-3</sup> between 20 and 30 cm depth with the afforestations being not significantly different from the pasture ( $P = 0.8$ ; Tab. 1). However, the lowest bulk density was recorded at both depths in the old forest resulting in a stand age effect close to significance ( $P = 0.06$ ). The stone content up to a depth of 50 cm was surprisingly low for alpine soils. Smaller rock fragments (>2 mm) determined by the sieving of soil material sampled with cores, accounted for about 1% of the soil volume in the surface layer and up to 8% in the 20-30 cm layer. In addition, we visually estimated the fraction of coarse stones not included in the sampled soil cores. The resulting total rock content, which was also used for C stock calculation, was on average 5, 10, 15, 20 and 40% for the 0-10, 10-20, 20-40, 40-60, and 60-80 cm layers, respectively.

All soils had clay contents around 50% except one of the 40-year old stands with a clay content of only 20% and a sand fraction of 70% (Tab.1). Soil pH ranged between 4 and 7 with significantly higher values in the deeper soils. Although the lowest pH values were found in the stands older than 30 years, neither the effect of vegetation type (pasture vs. forest) nor the one of stand age was statistically significant. In fact, pH was lowest in the 45 years old stand and not in the old forest. Soil pH under pasture was not affected by altitude but slightly increased along the horizontal gradient towards the Eastern part of the slope ( $P = 0.02$ ). Nevertheless, visual characterization of the soil profiles as well as analyses of the samples showed rather homogeneous soil properties within the whole study site.



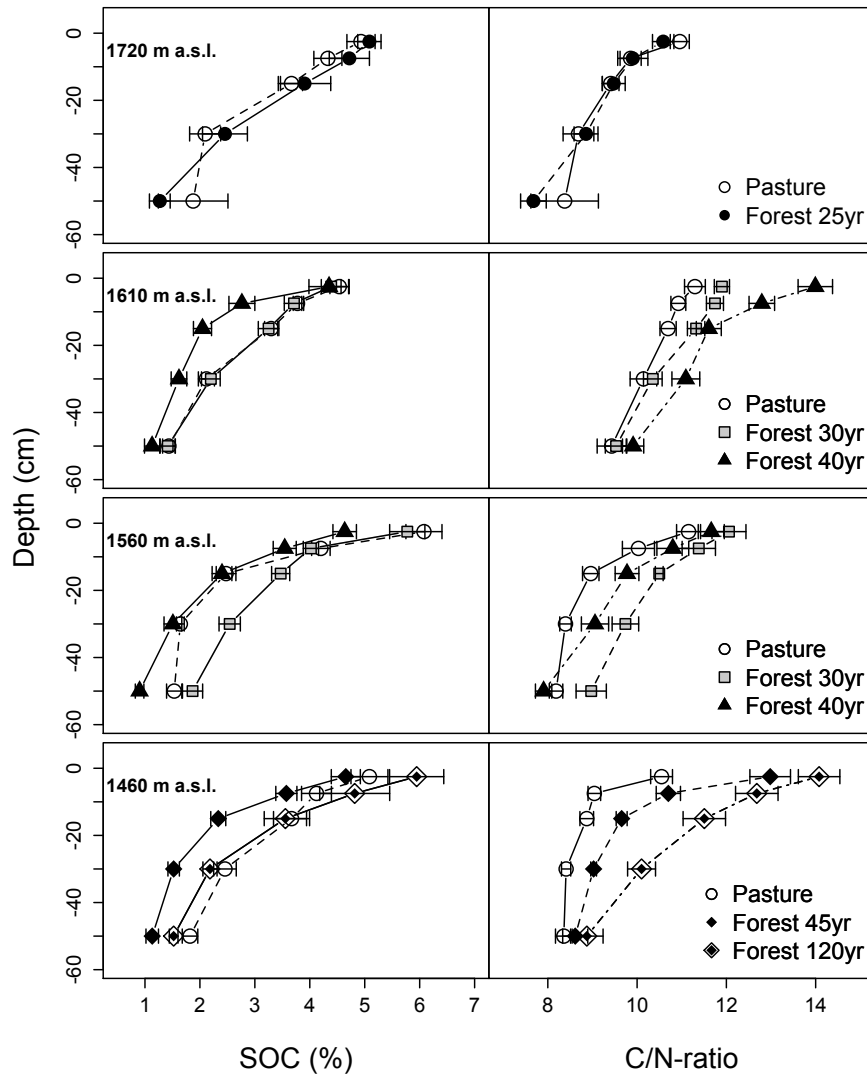
---

### 3.2 SOC concentrations and stocks

Soil carbon concentrations significantly decreased with soil depth in all the plots ( $P < 0.001$ ), indicating that the soils had not been disturbed or affected by erosion or landslides in the recent past. We tested the spatial homogeneity of SOC concentration of the study site by an altitudinal and horizontal gradient within the pasture. Neither altitude ( $P = 0.4$ ) nor horizontal position ( $P = 0.12$ ) significantly affected SOC concentrations. In fact, the soil C concentrations along the pasture transect at 1680 m a.s.l. slightly increased towards the Eastern part of the slope, which is in contrast to the lower C stocks of the forest stands on the Eastern part further down the slope at 1610 and 1560 m.a.s.l. (Fig. 1). By comparison to the small variability in the pasture, SOC concentration in the surface mineral soils (0-10 cm) showed a distinct pattern along the afforestation chronosequence, with the highest values under pasture and in the old forest ( $\sim 60 \text{ mg C g}^{-1}$ ) and the lowest values in the 40 and 45 years old stands ( $\sim 43 \text{ mg C g}^{-1}$ ) (Fig. 2). The same pattern was also apparent in the subsoils, with the lowest SOC concentrations in the 40 and 45 years old stands. Statistically, the transient decline of SOC was evidenced when the old forest was excluded from the analysis by linear mixed model ( $P_{\text{stand age}} < 0.01$ ).

Total SOC stocks ranged between 112 -154 t C ha<sup>-1</sup>, of which 83 – 100% was stored in the mineral soil and the remainder in the organic layer. Reflecting the SOC concentrations, SOC stocks of the mineral soils only moderately decreased in the first years following the tree plantation, but strongly declined after 30 years, with the lowest stocks in the 40 and 45 years old stands (Fig. 3). In the old forest, mineral soil SOC stocks were in the same range as the ones under pasture. Carbon stocks in the organic layer increased from 0 t C ha<sup>-1</sup> in the pasture to approximately 25 t C ha<sup>-1</sup> in the 45-year old forest and did hardly change to the old forest. The increasing C storage in the organic layer with stand age but slightly decreasing C stocks in the mineral soil resulted in unaltered total soil C stocks from the pasture to the 45 year old forest stands. The old forest had the highest total soil C stocks, with approximately 20 t ha<sup>-1</sup> more C stored than in the other soils of the pasture and younger stand ages. This increase corresponded to 60% higher C stocks when only the organic layer and the uppermost 10 cm were included and to 13% higher stocks when the total soil profiles were considered. Tree biomass strongly contributed to the increasing C storage with afforestation. In the 25 years old afforestation, an additional 86 t C ha<sup>-1</sup> was fixed in the plant biomass (above

and belowground) and this pool size increased up to 420t C ha<sup>-1</sup> in the old forest. Here, approximately three times the amount of SOC was stored in the tree biomass.



**Fig. 2:** Depth profiles of SOC concentrations and C:N-ratios ( $\pm$ SE) along the four transects under pasture and different aged spruce stands

### 3.3 Composition and quality of litter, fine roots and SOM

Chemical composition of the organic matter, as expressed by C:N ratios and <sup>13</sup>C natural abundances, strongly changed during decomposition (i.e. with soil depth in all soils; Fig 2; Fig.4). Fine root biomass (< 2 mm) in the uppermost 10 cm amounted to 0.6 kg m<sup>-2</sup> under forest and to 1 kg m<sup>-2</sup> under pasture. Grass roots were enriched in N (C:N ratio = 49) and had a lower lignin content (230 mg g<sup>-1</sup>) compared to the spruce fine roots (C:N ratio = 69, lignin content = 310 mg g<sup>-1</sup>, respectively) suggesting a higher decomposability of the grass roots. In addition, grass tissue contained a higher

proportion of N and less lignin than spruce needles (Table 2). Reflecting the altered above and belowground C input following the afforestation, soil C:N ratios were significantly narrower under pasture than under forest ( $p = 0.02$ ) and increased with stand age ( $P = 0.04$ ). However, the effect of vegetation on soil C:N ratios was only apparent up to a depth of 40 cm but not in the deeper soils. The  $\delta^{13}\text{C}$  values of soils under forest were significantly higher than the ones under pasture ( $P < 0.01$ ). Similar to the soil C:N ratios, the effect of vegetation on soil  $\delta^{13}\text{C}$  signature was most pronounced in the surface soil while it decreased with increasing soil depth and vanished below 20cm. In the uppermost mineral soil (0-5cm), the difference between the two land use types amounted to about 1.5‰. After tree plantation,  $\delta^{13}\text{C}$  values at 0-5cm became gradually more positive with increasing stand age (Fig. 4), shifting from -27‰ under pasture to -25.5‰ under the old forest.

**Table 2:** Chemical composition of the fresh plant material and the plant residues in the organic layer

	<b>C (%)</b>	<b>CN Ratio</b>	<b><math>\delta^{13}\text{C}</math> (‰)</b>	<b>Lignin (%)</b>
<b>Plant material</b>				
Gras foliage	42.1 (0)	26 (4.9)	-27.3 (0.4)	18.3 (0.1)
Gras roots	32.7 (2.0)	49 (2.3)	-27.2 (0.1)	23.2 (1.0)
<b>Spruce roots</b>	<b>42.6 (2.0)</b>	<b>69 (4.0)</b>	<b>-26.6 (0.2)</b>	<b>30.6 (1.0)</b>
Bark	48.9 (0.1)	67 -	-25.6 (0.6)	44 (1.0)
Wood	46.4 (0.3)	550 -	-26.0 (1.2)	-
Needle	47.4 (0.4)	40 (2.4)	-29.4 (0.5)	31 (0.5)
<b>Oranic layer</b>				
Oi	44.2 (0.8)	42 (2.3)	-28.0 (0.1)	-
Oe	41.6 (1.4)	25 (1.1)	-27.4 (2.0)	-
Oa	31.9 (4.1)	25 (0.3)	-27.3 (0.3)	-

### 3.4 Soil respiration, microbial activity, and microclimate

Soil respiration changed strongly along the chronosequence. Average soil respiration rates (July to October) ranged between  $2.3 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  in the youngest stand and  $6.4 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  in the pasture. As expected, soil respiration was strongly driven by soil temperature ( $P < 0.001$ ), but not by soil moisture ( $P = 0.4$ ). Microclimatic measurements showed about  $5^\circ\text{C}$  lower soil temperatures at 5 cm depth under forest than under pasture during the growing season (April – September). In winter, the

difference in temperature between the two vegetation types was smaller but the forest soils were frozen for a longer time period than the pasture soils. Moreover, forest soils had considerably lower volumetric water contents than pasture soils (Fig. 6). However, even after fitting soil temperature and soil moisture as covariates into the model, soil respiration was still significantly higher in the pasture plots ( $P = 0.02$ ).

Under standardized conditions in the laboratory, C mineralization per gram dry and root-free soil did not significantly differ between pasture and forest mineral soils, either for the cumulated amount of respired C at the end of the incubation ( $P = 0.11$ ) or the amount of C after glucose addition (as a measure for the microbial biomass  $P = 0.09$ ). Not unexpectedly, C mineralization was positively affected by the soil C content of the mineral soil samples ( $P < 0.001$ ). As a result, when C mineralization was related to g soil C, pasture soils released significantly more  $\text{CO}_2$  ( $P < 0.01$  for both), indicating that SOC contained a larger proportion of labile C in the pasture than in the forest. C mineralization in the organic horizons was 10 to 20 times higher than in the mineral soils and the mineralization rates of the organic layer material were positively correlated with the C:N ratios ( $P = 0.003$ ).

#### 4. Discussion

Land-use changes in alpine regions in Switzerland are dramatic: the forest area primarily covered by spruce has increased by as much as 8% during the last decade. Similar changes occur in other regions of European mountain ranges (FAO, 2010). Our chronosequence study shows that afforestation with spruce on an alpine pasture decreased soil respiration rates, altered soil organic matter quality and induced less favorable microclimatic conditions. However, the afforestation had only a moderate impact on total soil C stocks, with an accumulation of forest floor and a small transient decline of C stocks in the mineral soil.

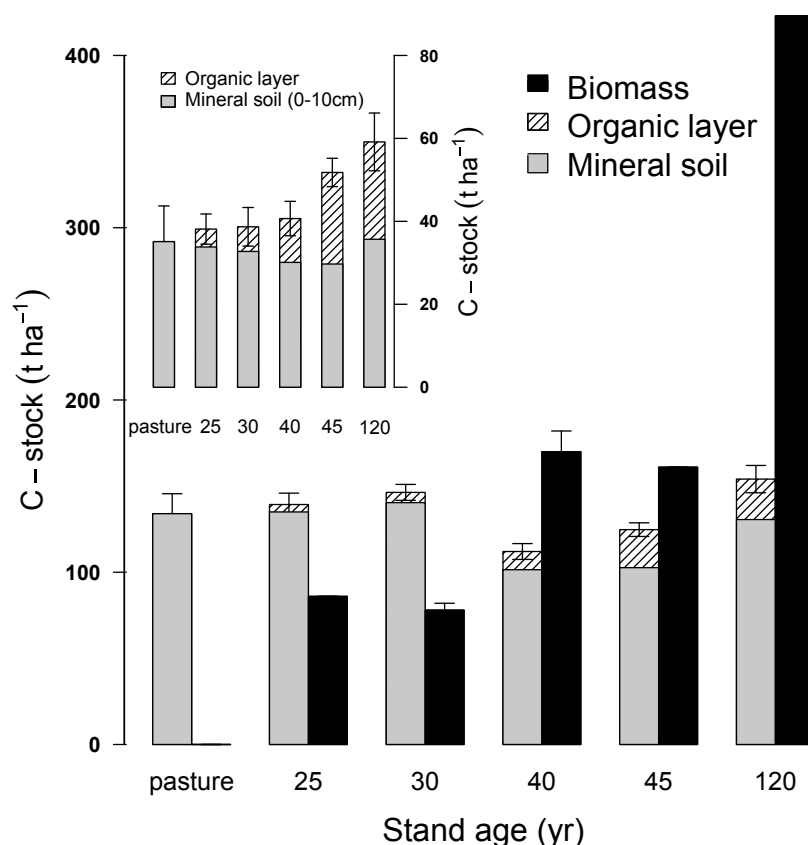
The detection of changes in soil C-stocks through afforestation by the “space for time” approach applied in this and other studies relies on homogeneous soil conditions. This is particularly important given that our study site spanned an altitudinal gradient of 250 m, along which temperatures and length of vegetation period changes, which, in turn, might affect organic matter input. For example, in a Swiss alpine pasture, root biomass increased by 25% and turnover times almost doubled between 1665 and 2520 m a.s.l. (Hitz et al., 2001). Accordingly, Leifeld et al. (2005) estimated for Swiss alpine

grasslands that SOC-concentrations at 0-20 cm depth increase with altitude at a rate of about 2mg C g<sup>-1</sup> per 100m. However, at our study site, we did not find statistical differences in SOC concentrations within the pasture soils, either along the altitudinal gradient or along an East-West transect encompassing 20 soil pits under pasture (Fig. 1). Moreover, soil properties such as soil texture, stone content, soil density, thickness of the soil horizons, and soil color were also very homogeneous across the whole slope (Table 1).

#### **4.1 Moderate effects on soil C pools**

Soil organic C stocks of the mineral soils ranged between 101 and 140 t C ha<sup>-1</sup> which is higher than the ones reported by Thuille and Schulze (2006) in a series of different aged spruce afforestations in the Alps and Thuringia (40-100 t C ha<sup>-1</sup> for the whole soil profile) but close to the mean value for Swiss alpine forest soils (~130 t C ha<sup>-1</sup>) (Hagedorn et al., 2010b). The gradual accumulation of forest floor at a rate of 0.19 t C ha<sup>-1</sup> yr<sup>-1</sup> resulted in an additional 23 t C ha<sup>-1</sup> stored in the old forest. Taking this C pool into calculation, the soil C pool in the old forest exceeded the one of the pasture soils by 20 t ha<sup>-1</sup> (Fig. 3), which is close to the average difference of 22 t ha<sup>-1</sup> between the two land use types in Switzerland (Bolliger et al., 2008). By comparison, SOC stocks of the mineral soil transiently decreased in the 40 and 45y old stands and increased again in the old forest (Fig. 3). This pattern is in line with the findings of other chronosequence studies along afforestation (Poeplau et al., 2011), although the main C loss in our study apparently occurred with a time lag of a few decades after the tree plantation. In general, the magnitude of soil C loss with afforestation principally depends on the degree of disturbance with site preparation (Jandl et al., 2007) and the persistence of the grass cover providing a sustained input of fine root C into mineral soils (Thuille and Schulze, 2006). In this study, soil disturbance at planting and hence C losses through mineralization were minimal as the spruce saplings were planted manually on the relatively steep subalpine slope. This might explain that the main C loss in the surface soil did not occur directly after the tree plantation but was more closely related to the cessation of the dense grass cover with tree canopy closure 20 to 30 years after afforestation. We also have hypothesized that the C-loss would primarily occur in the topsoil due to declining fine root densities and the development of deeper rooting systems following afforestation (Jackson et al., 1996). However, our data suggest the

opposite, C was primarily lost from the subsoil. Afforestation effects on SOC stored in the deeper soil layer are ambiguous to date as the majority of the LUC studies only comprised topsoils. While some studies found that afforestation increases C storage in the subsoil (Hooker and Compton, 2003; Ouimet et al., 2007; Wei et al., 2012), others reported declining C stocks (Thuille and Schulze, 2006; Tremblay et al., 2007). After afforesting cotton fields with loblolly pine, (Richter et al., 1999) observed increasing C stocks in the surface soil accompanied by a loss of C in the deeper soil layers. Their analysis of  $^{13}\text{C}$  and  $^{14}\text{C}$  isotopes indicated that these C losses from the deeper soil occurred despite a substantial input of new root C and DOC. These new C inputs, however, were not stabilized but probably instead might have stimulated the decomposition of inherent SOC (Mobley, 2011). Since subsoils are lacking energy-rich substrate needed for soil microbes to maintain their metabolism, they presumably are more prone to priming than surface soils with higher C availability (Fontaine et al., 2007).



**Fig. 3:** C-stocks in the different compartments ( $\pm$ SE) in the pasture and different aged spruce stands. Small figure shows soil C stocks in the uppermost 10cm of mineral soil and organic layers

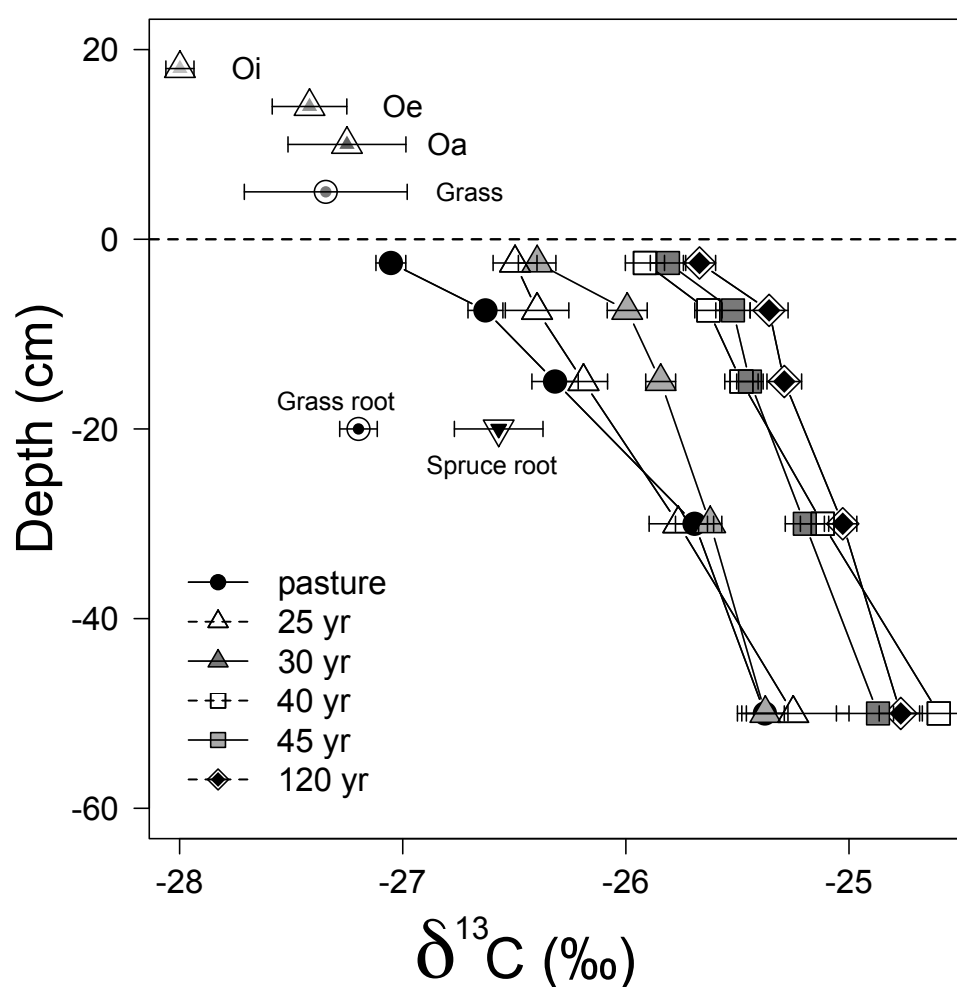
Our data clearly show the importance of the chosen sampling depth for the interpretation of the results (Harrison et al., 2011). If we had included only the organic layer and the uppermost mineral soil (0-10cm), the afforestation would have increased the soil C stocks by 60%. However, if the whole soil profile is taken into account, the afforestation effect is much smaller (~13%) and C stocks depend more strongly on subsoil C dynamics (Fig. 3). In any case, SOC pools on a mass basis were only marginally affected by the land use change as compared to C stored in tree biomass. Forty years after land use conversion, C stock of tree biomass exceeded the one of the soil. In the old forest, trees almost stored three times as much C as soils (Fig. 3), although the C pool of the trees in this study is much larger than the average pool of ~100 t C ha<sup>-1</sup> in the Swiss Alps (Hagedorn et al., 2010b). Thus, afforestation turned pasture into a C sink mainly by C accumulation in vegetation, whilst the effects on SOC were comparatively small; the increase of the SOC stocks of 20 t C ha<sup>-1</sup> following afforestation corresponds to only 20% and 5% of the average biomass C pool in the Alps or in this study, respectively.

#### **4.2 Effects on soil C quality**

In contrast to the moderate effects on total soil C stocks, spruce plantation strongly influenced the composition and quality of SOM and its vertical distribution in soils. After tree establishment,  $\delta^{13}\text{C}$  values in the surface mineral soil shifted from -27 ‰ in the pasture towards -25.5 ‰ in the mature forest. Despite the rather small difference of 1.5 ‰ between the two vegetation types, the increase of the isotopic ratio was very consistent (Fig 4). The impact of vegetation on  $\delta^{13}\text{C}$  values decreased with soil depth, which implies a stronger dominance of older soil C in the subsoils (e.g. Richter et al., 1999). The lower  $\delta^{13}\text{C}$  values of the upper mineral soils under pasture than under old forest might partly reflect the C input from grass roots that are more depleted in  $^{13}\text{C}$ , while the larger difference in  $\delta^{13}\text{C}$  values between spruce roots and SOM in the old forest soils might result from a stronger discrimination during decomposition, which is indicative for a slower root turnover (Chen et al., 2005). However, it is beyond the scope of this study to fully assess the processes leading to the different  $\delta^{13}\text{C}$  values under pasture and forest, but we interpret it as an evidence for a replacement of pasture by tree-derived C in SOM with time after afforestation.

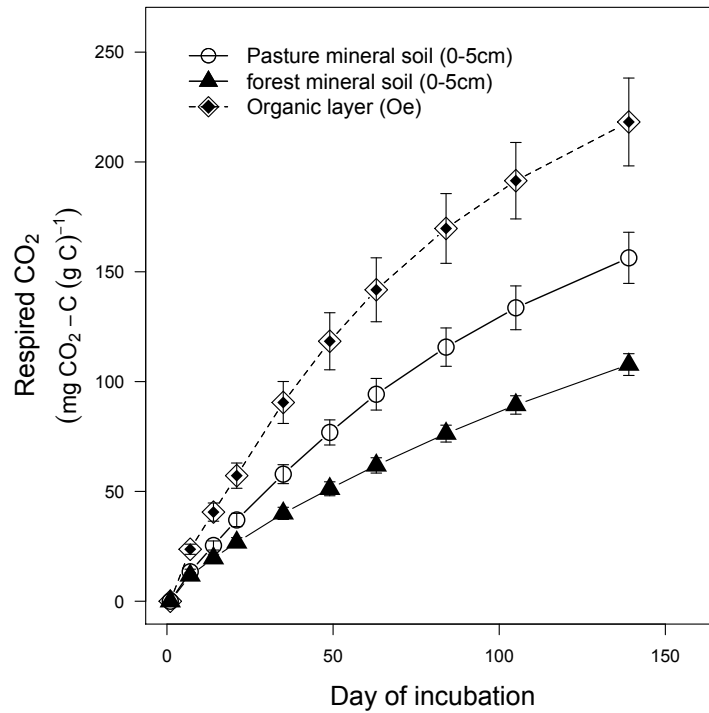
Afforestation directly affects SOM dynamics through changes of the input pathways and the quantity and quality of plant residues (De Deyn et al., 2008). In the pasture, C-inputs

into soils were mainly restricted to grass roots as most of the foliage was removed by grazing cattle. In the afforestations, by contrast, a substantial amount of C originates from needle fall. However, the accumulation of a forest floor suggests that only a small fraction of this needle litter was incorporated into the mineral soil. In a field experiment with  $^{13}\text{C}$  labeled pine needles, Bird and Torn (2006) reported that less than 1% of the pine C were allocated from the organic layer into the A horizon within 1.5 years and Tate et al. (2011) estimated that only 1 to 2.2% of fresh  $^{14}\text{C}$  labeled pine needles were transferred into the mineral soil within one year in a young pine stand. In addition to the aboveground litter input, afforestation also affects the belowground C dynamics through the characteristics of fine roots (Guo et al., 2007) representing a very important SOC source either through root exudates or as litter (Rasse et al., 2005). In our study, the amounts of fine roots (<2mm) in the spruce stands were 40% lower than under pasture.



**Fig. 4:** Natural abundance of the  $^{13}\text{C}$  Isotope ( $\pm\text{SE}$ ) of the plant material and the SOM at different depths under pasture and different aged spruce stands





**Fig. 5:** Cumulative fluxes of  $\text{CO}_2$  ( $\pm\text{SE}$ ) respired from the incubated pasture and forest mineral soils (0-5cm) and the organic layer (Oe) as an indicator for the fraction of labile carbon

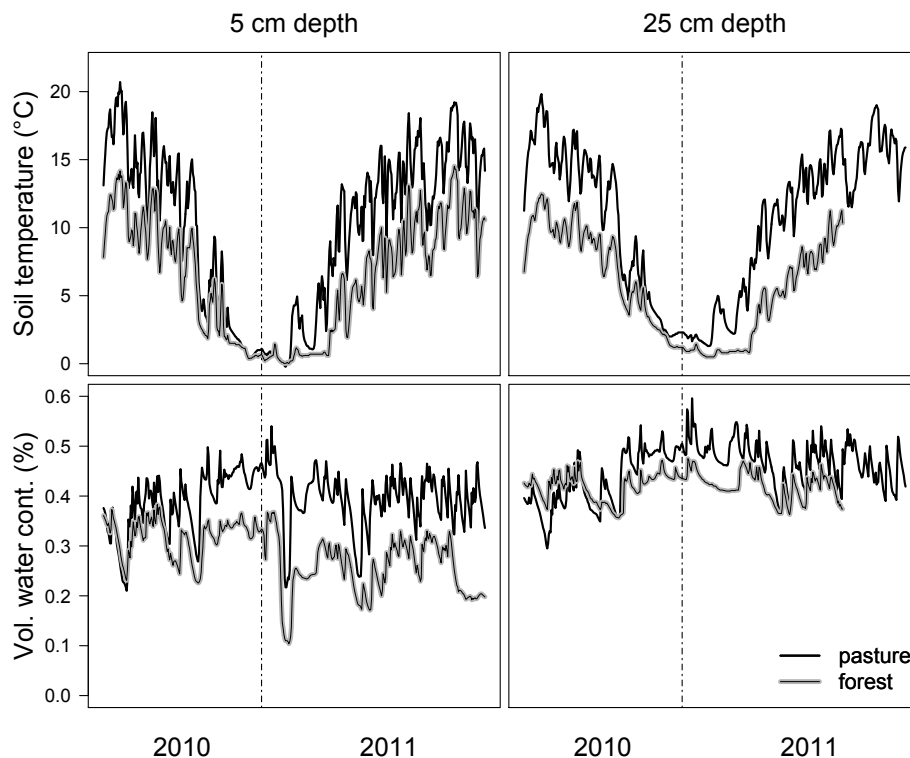
Moreover, fine roots from trees had a wider C:N ratio (spruce = 70; grass = 48) and lignin contents (spruce = 310  $\text{mg g}^{-1}$ ; grass = 240  $\text{mg g}^{-1}$ ), indicating lower decomposability. These changes in litter quality were also reflected in the higher C:N-ratio and smaller proportion of labile C in the forest than in the pasture soils which confirms our hypothesis that SOM quality declines with afforestation.

### 4.3 Afforestation decreases soil respiration

Soil respiration, as an indicator for the biological activity and carbon turnover, strongly declined after the tree plantation and only slowly increased thereafter. Though, in the old forest, respiration was 30% lower than prior to the land use change. What drives the reduced respiratory activity under forest?

The incubation experiment under standardized conditions revealed that the amount of mineralized C per gram mineral soil did not significantly differ between forest and pasture in, in spite of the higher fraction of labile C in the pasture soils (mineralized C per unit SOC) (Fig. 5, Fig. 7). Therefore, the higher C availability did not lead to significantly higher C mineralization in the pasture soils. This suggests that not soil C quality but other factors like microclimate or root respiration are primarily responsible

for the higher soil respiration in the grassland than in the forest. Averaged over the growing season, surface soils under forest were about 5°C cooler than under pasture. Given that soil temperature had a significant positive effect on the respiration ( $p < 0.001$ ) and assuming a  $Q_{10}$  of 3 (Hagedorn et al., 2010a), the lower soil temperature theoretically would lead to 50% smaller soil respiration rates in the forests, which approximately was the case for the middle aged stands (30 and 40 yr.). In agreement with our data, Smith and Johnson (2004) measured 38% smaller respiration rates after juniper encroachment on former grassland, which is also close to the 41% reduction reported by Kellman et al. (2007) in a comparison between pasture and two forests in Canada. In both studies, soil CO<sub>2</sub> efflux was mainly attributed to a decline of soil temperature after tree establishment.

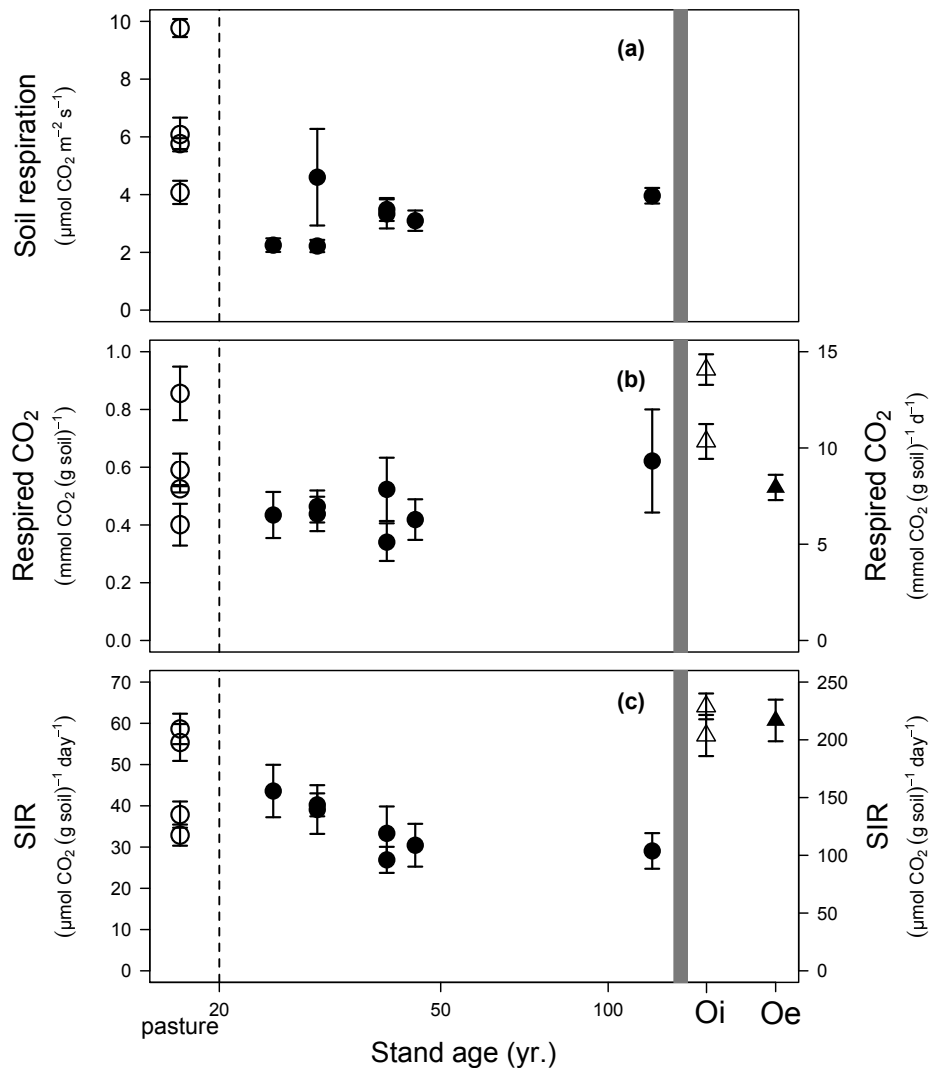


**Fig. 6:** Soil temperature and soil moisture at 5 and 25cm depth under pasture and in a 40 years old spruce stand

However, in addition to temperature, soil moisture can also be a critical factor for biological activity (Borken et al., 2003; Muhr and Borken, 2009), especially when it drops below a minimum threshold (Curiel Yuste et al., 2003). According to our model, volumetric water content did not significantly affect respiration rates when fitted after temperature ( $P = 0.4$ ). However, under dry conditions (volumetric SWC < 20%)

---

respiration was rather limited. Moreover, C mineralization of the incubated organic layers was up to 20 times lower when the soil samples were field moist and not adjusted to the optimal conditions during incubation (50% of water holding capacity). This strongly suggests that water availability contributes to the control of SOM decomposition on this South facing slope, especially during the dry summer months. This is even more reasonable as a considerable amount of the respired CO<sub>2</sub> in the forest originates from the organic layer (Buchmann, 2000; Kammer et al., 2009), which is most susceptible to dry out. However, in our linear-mixed model, land use type still remained significant even after correcting for the soil temperature and moisture effects ( $P = 0.02$ ). Therefore, additional factors must be at play, of which root respiration presumably is most important. Autotrophic respiration contributes about 50% to total soil respiration, but its proportion considerably varies throughout the year and among different vegetation types (Hanson et al., 2000). As the grassland soils had a 40% higher root biomass, it seems likely that autotrophic respiration was higher in the pasture than in the spruce stands. In summary, we interpret the 50% lower soil respiration in the forest than in the grassland as a combined effect of a colder and drier soil conditions, a reduced root respiration, and a lower SOM quality. The fact that the lower C turnover under forest did not increase SOC storage - at least not in the short term - suggests that it must have been offset by a decreased C input from plants. This is in line with the findings of Scott et al. (2006), who attributed the concomitant decreases in C stocks and soil respiration rates after pine afforestation to a declined C input into the mineral soil. On a larger scale compiling Eddy flux data across Europe, Schulze et al. (2010) reported that both heterotrophic respiration rates and net primary production in forests are considerably lower than in grasslands, which supports our conclusion of a reduced C input after afforestation. The highest SOC stocks in the more than 120-year old forest, however, show that afforestation has a positive effect on soil C storage in this subalpine ecosystem.



**Fig. 7:** Mean soil respiration in situ (a), cumulative CO<sub>2</sub> release per gram incubated soil after 140 days (b), and CO<sub>2</sub> release per gram incubated soil after glucose addition (Substrate Induced Respiration) (c) as affected by vegetation type and stand age. Mineral soils (0-5cm) are displayed in the left panel and organic layer in the right panel

## 5. Conclusions

Our results show that spruce afforestation on an extensively grazed alpine pasture had only small effects on total soil C storage. Soil C stocks in the mineral soil transiently decreased after tree establishment, reaching a minimum 40 to 45 years after afforestation and increased again thereafter. Soils of the mature spruce forest (>120 years) stored more C than pasture soils, mainly due to the accumulation of an organic layer. In contrast to soil C stocks, afforestation strongly altered SOM quality with forest soils containing lower fractions of labile C and having higher C:N ratios as pasture soils. Soil respiration significantly decreased after the land use change probably primarily due

to a less favorable microclimate and reduced root respiration in the forests. This implies, in conjunction with the moderate effects on total soil C stocks that C inputs into soils and hence also ecosystem C cycling rates must have decreased considerably with afforestation. However, the high soil C stocks in the old forest strongly suggest that on a centennial time scale, afforestation has positive effects on soil C storage, but this increase is small as compared to C accumulated in tree biomass.

## Acknowledgements

We gratefully thank P. A. Niklaus for the measurement of the gas samples, M. Walser and R. Köchli for the field assistance and G.D. Lieberherr, A. Zürcher and C. Herzog for assistance in the laboratory. This study was funded by the COST Action 639 (BurnOut) and the Swiss Federal Office for the Environment (FOEN)

## References

- Berg, G., Smalla, K., 2009. Plant species and soil type cooperatively shape the structure and function of microbial communities in the rhizosphere. *Fems Microbiology Ecology* 68(1), 1-13.
- Bird, J.A., Torn, M.S., 2006. Fine roots vs. Needles: A comparison of (13)C and (15)N dynamics in a ponderosa pine forest soil. *Biogeochemistry* 79(3), 361-382.
- Bolliger, J., Hagedorn, F., Leifeld, J., Boehl, J., Zimmermann, S., Soliva, R., Kienast, F., 2008. Effects of land-use change on carbon stocks in Switzerland. *Ecosystems* 11(6), 895-907.
- Borken, W., Davidson, E.A., Savage, K., Gaudinski, J., Trumbore, S.E., 2003. Drying and wetting effects on carbon dioxide release from organic horizons. *Soil Science Society of America Journal* 67(6), 1888-1896.
- Brändli, U.-B., 2010. Schweizerisches Landesforstinventar. Ergebnisse der dritten Erhebung 2004–2006. Birmensdorf, Eidgenössische Forschungsanstalt für Wald, Schnee und Landschaft WSL. Bern, Bundesamt für Umwelt, BAFU. 312 S.
- Buchmann, N., 2000. Biotic and abiotic factors controlling soil respiration rates in *Picea abies* stands. *Soil Biology & Biochemistry* 32(11-12), 1625-1635.
- Budge, K., Leifeld, J., Hiltbrunner, E., Fuhrer, J., 2011. Alpine grassland soils contain large proportion of labile carbon but indicate long turnover times. *Biogeosciences* 8(7), 1911-1923.
- Chen, Q.Q., Shen, C.D., Sun, Y.M., Peng, S.L., Yi, W.X., Li, Z.A., Jiang, M.T., 2005. Spatial and temporal distribution of carbon isotopes in soil organic matter at the Dinghushan Biosphere Reserve, South China. *Plant and Soil* 273(1-2), 115-128.
- Cheng, W.X., Virginia, R.A., 1993. Measurement of microbial biomass in arctic tundra soils using fumigation extraction and substrate-induced respiration procedures. *Soil Biology & Biochemistry* 25(1), 135-141.
- Curiel Yuste, J., Janssens, I.A., Carrara, A., Meiresonne, L., Ceulemans, R., 2003. Interactive effects of temperature and precipitation on soil respiration in a temperate maritime pine forest. *Tree Physiology* 23(18), 1263-1270.
- De Deyn, G.B., Cornelissen, J.H.C., Bardgett, R.D., 2008. Plant functional traits and soil carbon sequestration in contrasting biomes. *Ecology Letters* 11(5), 516-531.
- Dence, C.W., 1992. The determination of lignin. In: Lin, S.Y. & Dence, C.W. (Eds.) *Methods in lignin chemistry*. Springer-Verlag, Berlin Heidelberg. p. 33-61.
- FAO, 2010. *Global Forest Resources Assessment 2010, Main Report*. FAO Forestry Paper 163. Rome.

- 
- Fontaine, S., Barot, S., Barre, P., Bdioui, N., Mary, B., Rumpel, C., 2007. Stability of organic carbon in deep soil layers controlled by fresh carbon supply. *Nature* 450(7167), 277-U210.
- Gee, G.W., Bauder, J.W., 1986. Particle size analysis. In: Klute, A. (ed.), *Methods of Soil Analysis. Part 1. Physical and Mineralogical Methods* 386-411, Am. Soc. Agron., Madison, WI.
- Guo, L.B., Gifford, R.M., 2002. Soil carbon stocks and land use change: a meta analysis. *Global Change Biology* 8(4), 345-360.
- Guo, L.B., Wang, M., Gifford, R.M., 2007. The change of soil carbon stocks and fine root dynamics after land use change from a native pasture to a pine plantation. *Plant and Soil* 299(1-2), 251-262.
- Hagedorn, F., Martin, M., Rixen, C., Rusch, S., Bebi, P., Zuercher, A., Siegwolf, R.T.W., Wipf, S., Escape, C., Roy, J., Haettenschwiler, S., 2010a. Short-term responses of ecosystem carbon fluxes to experimental soil warming at the Swiss alpine treeline. *Biogeochemistry* 97(1), 7-19.
- Hagedorn, F., Moeri, A., Walthert, L., Zimmermann, S., 2010b. Kohlenstoff in Schweizer Waldböden – bei Klimaerwärmung eine potenzielle CO<sub>2</sub>-Quelle. *Schweizerische Zeitung für Forstwesen* 161(12), 530-535.
- Hagedorn, F., Mulder, J., Jandl, R., 2010c. Mountain soils under a changing climate and land-use. *Biogeochemistry* 97(1), 1-5.
- Hanson, P.J., Edwards, N.T., Garten, C.T., Andrews, J.A., 2000. Separating root and soil microbial contributions to soil respiration: A review of methods and observations. *Biogeochemistry* 48(1), 115-146.
- Harrison, R.B., Footen, P.W., Strahm, B.D., 2011. Deep Soil Horizons: Contribution and Importance to Soil Carbon Pools and in Assessing Whole-Ecosystem Response to Management and Global Change. *Forest Science* 57(1), 67-76.
- Hiltbrunner, D., Zimmermann, S., Karbin, S., Hagedorn, F., Niklaus, P.A., 2012. Increasing soil methane sink along a 120-year afforestation chronosequence is driven by soil moisture DOI: 10.1111/j.1365-2486.2012.02798.x. *Global Change Biology*.
- Hitz, C., Egli, M., Fitze, P., 2001. Below-ground and above-ground production of vegetational organic matter along a climosequence in alpine grasslands. *Journal of Plant Nutrition and Soil Science* 164(4), 389-397.
- Hooker, T.D., Compton, J.E., 2003. Forest ecosystem carbon and nitrogen accumulation during the first century after agricultural abandonment. *Ecological Applications* 13(2), 299-313.
- Houghton, R.A., 2003. Revised estimates of the annual net flux of carbon to the atmosphere from changes in land use and land management 1850-2000. *Tellus Series B-Chemical and Physical Meteorology* 55(2), 378-390.
- Jackson, R.B., Canadell, J., Ehleringer, J.R., Mooney, H.A., Sala, O.E., Schulze, E.D., 1996. A global analysis of root distributions for terrestrial biomes. *Oecologia* 108(3), 389-411.
- Jandl, R., Lindner, M., Vesterdal, L., Bauwens, B., Baritz, R., Hagedorn, F., Johnson, D.W., Minkinen, K., Byrne, K.A., 2007. How strongly can forest management influence soil carbon sequestration? *Geoderma* 137(3-4), 253-268.
- Kammer, A., Hagedorn, F., Shevchenko, I., Leifeld, J., Guggenberger, G., Goryacheva, T., Rigling, A., Moiseev, P., 2009. Treeline shifts in the Ural mountains affect soil organic matter dynamics. *Global Change Biology* 15(6), 1570-1583.
- Kaufmann, E., 2001. Estimation of standing timber, growth and cut. In: P. Brassel and H. Lischke (Ed.), *Swiss National Forest Inventory: Methods and Models of the Second Assessment*. Swiss Federal Research Institute WSL, Birmensdorf, S. 162-196.
- Kellman, L., Beltrami, H., Risk, D., 2007. Changes in seasonal soil respiration with pasture conversion to forest in Atlantic Canada. *Biogeochemistry* 82(1), 101-109.
- Leifeld, J., Bassin, S., Fuhrer, J., 2005. Carbon stocks in Swiss agricultural soils predicted by land-use, soil characteristics, and altitude. *Agriculture Ecosystems & Environment* 105(1-2), 255-266.
- Leifeld, J., Zimmermann, M., Fuhrer, J., Conen, F., 2009. Storage and turnover of carbon in grassland soils along an elevation gradient in the Swiss Alps. *Global Change Biology* 15(3), 668-679.
- Macdonald, C.A., Thomas, N., Robinson, L., Tate, K.R., Ross, D.J., Dando, J., Singh, B.K., 2009. Physiological, biochemical and molecular responses of the soil microbial community after afforestation of pastures with *Pinus radiata*. *Soil Biology & Biochemistry* 41(8), 1642-1651.

- MacDonald, D., Crabtree, J.R., Wiesinger, G., Dax, T., Stamou, N., Fleury, P., Lazpita, J.G., Gibon, A., 2000. Agricultural abandonment in mountain areas of Europe: Environmental consequences and policy response. *Journal of Environmental Management* 59(1), 47-69.
- Mobley, M.L., 2011. An Ecosystem Approach to Dead Plant Carbon over 50 years of Old-Field Forest. Dissertation University Program in Ecology in the Graduate School of Duke University.
- Muhr, J., Borken, W., 2009. Delayed recovery of soil respiration after wetting of dry soil further reduces C losses from a Norway spruce forest soil. *Journal of Geophysical Research-Biogeosciences* 114.
- Ouimet, R., Tremblay, S., Perie, C., Pregent, G., 2007. Ecosystem carbon accumulation following fallow farmland afforestation with red pine in southern Quebec. *Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere* 37(6), 1118-1133.
- Paul, K.I., Polglase, P.J., Nyakuengama, J.G., Khanna, P.K., 2002. Change in soil carbon following afforestation. *Forest Ecology and Management* 168(1-3), 241-257.
- Poeplau, C., Don, A., Vesterdal, L., Leifeld, J., Van Wesemael, B., Schumacher, J., Gensior, A., 2011. Temporal dynamics of soil organic carbon after land-use change in the temperate zone - carbon response functions as a model approach. *Global Change Biology* 17(7), 2415-2427.
- Post, W.M., Kwon, K.C., 2000. Soil carbon sequestration and land-use change: processes and potential. *Global Change Biology* 6(3), 317-327.
- R: Development Core Team, 2010. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rasse, D.P., Rumpel, C., Dignac, M.F., 2005. Is soil carbon mostly root carbon? Mechanisms for a specific stabilisation. *Plant and Soil* 269(1-2), 341-356.
- Richter, D.D., Markewitz, D., Trumbore, S.E., Wells, C.G., 1999. Rapid accumulation and turnover of soil carbon in a re-establishing forest. *Nature* 400(6739), 56-58.
- Schulze, E.D., Ciais, P., Luyssaert, S., Schrumpf, M., Janssens, I.A., Thiruchittampalam, B., Theloke, J., Saurat, M., Bringezu, S., Lelieveld, J., Lohila, A., Rebmann, C., Jung, M., Bastviken, D., Abril, G., Grassi, G., Leip, A., Freibauer, A., Kutsch, W., Don, A., Nieschulze, J., Boerner, A., Gash, J.H., Dolman, A.J., 2010. The European carbon balance. Part 4: integration of carbon and other trace-gas fluxes. *Global Change Biology* 16(5), 1451-1469.
- Scott, N.A., Tate, K.R., Ross, D.J., Parshotam, A., 2006. Processes influencing soil carbon storage following afforestation of pasture with *Pinus radiata* at different stocking densities in New Zealand. *Australian Journal of Soil Research* 44(2), 85-96.
- Smith, D.L., Johnson, L., 2004. Vegetation-mediated changes in microclimate reduce soil respiration as woodlands expand into grasslands. *Ecology* 85(12), 3348-3361.
- Tappeiner, U., Tappeiner, G., Hilbert, A., Mattanovich, E., 2003. *The EU Agricultural Policy and the Environment. Evaluation of the Alpine Region*. Blackwell, Berlin, Germany.
- Tasser, E., Walde, J., Tappeiner, U., Teutsch, A., Noggler, W., 2007. Land-use changes and natural reforestation in the Eastern Central Alps. *Agriculture Ecosystems & Environment* 118(1-4), 115-129.
- Tate, K.R., Lambie, S.M., Ross, D.J., Dando, J., 2011. Carbon transfer from  $^{14}\text{C}$ -labelled needles to mineral soil, and  $^{14}\text{C}$ - $\text{CO}_2$  production, in a young *Pinus radiata* Don stand. *European Journal of Soil Science* 62(1), 127-133.
- Thuille, A., Schulze, E.D., 2006. Carbon dynamics in successional and afforested spruce stands in Thuringia and the Alps. *Global Change Biology* 12(2), 325-342.
- Tremblay, S., Perie, C., Pregent, G., Ouimet, R., 2007. Ecosystem carbon accumulation following fallow farmland afforestation with red pine in southern Quebec. *Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere* 37(6), 1118-1133.
- Walthert, L., Graf, U., Kammer, A., Luster, J., Pezzotta, D., Zimmermann, S., Hagedorn, F., 2010. Determination of organic and inorganic carbon,  $\delta^{13}\text{C}$ , and nitrogen in soils containing carbonates after acid fumigation with HCl. *Journal of Plant Nutrition and Soil Science* 173(2), 207-216.
- Wei, X., Qiu, L., Shao, M., Zhang, X., Gale, W.J., 2012. The Accumulation of Organic Carbon in Mineral Soils by Afforestation of Abandoned Farmland. *Plos One* 7(3).

---

*Wiesmeier, M., Spoerlein, P., Geuss, U., Hangen, E., Haug, S., Reischl, A., Schilling, B., von Luetzow, M., Koegel-Knabner, I., 2012. Soil organic carbon stocks in southeast Germany (Bavaria) as affected by land use, soil type and sampling depth. Global Change Biology 18(7), 2233-2245.*



---

## Acknowledgements

Die Arbeit an meiner Dissertation war bei weitem meine interessanteste berufliche Beschäftigung bisher. Zum positiven Gelingen hat, neben der fachlichen Betreuung, vor allem das sehr angenehme Arbeitsklima an der WSL beigetragen; die Tage verminderter Motivation hielten sich vor allem auch dank des Umfeldes in engen Grenzen.

Zuerst möchte ich mich bei Stephan Zimmermann für seine umfassende Betreuung meiner Arbeit und die sehr angenehme Zusammenarbeit bedanken. Auch meinem zweiten Betreuer, Frank Hagedorn, gebührt ein spezieller Dank für die Unterstützung bei meinen Versuchen und die aktive Mitgestaltung meiner Paper. Pascal Niklaus vom Institut für Evolutionsbiologie und Umweltwissenschaften der Uni Zürich danke ich für die Betreuung der Treibhausgasmessungen und seine wertvollen Inputs zur Statistik. Euch ist es zu einem grossen Teil zuzuschreiben, dass diese Arbeit nun in dieser Form vorliegt!

Ausserdem danke ich folgenden Personen:

Michael Schmidt für die Oberaufsicht meiner Dissertation und, zusammen mit den anderen Mitgliedern der Bodengruppe des Geographischen Institutes der Universität Zürich, für neue Inputs zur Arbeit. Michael Hilf gebührt ein besonderer Dank für die Messungen meiner PLFA Proben.

Sandra Spielvogel von der Universität Koblenz für die Einführung in die Pflanzenbiomarker Analyse.

Allen Kollegen der Forschungseinheit Waldböden und Biogeochemie an der WSL. Vor allem Roger Köchli, Marco Walser, Dani Christen, Oli Schramm, Nouredine Hajjar und Yuk Ying Cheung-Tang für ihre Mithilfe in Feld und Labor. Meinen Büromitbewohnern Alois Zürcher, Adi Kammer, Beni Huber und Greg Tomlinson (Danke für all die Englischkorrekturen!) für die Fach- Fussball- Kletter- und anderen Diskussionen.

Den Mitarbeitern des Zentrallabors der WSL, Alessandro Schlumpf, Ursula Graf und Janka Bollenbach für Messungen und Methodenentwicklung.

Sebastian Schulze, der mit seiner Masterarbeit stark zum ersten Paper beigetragen hat und Gian Lieberherr für seine grosse Hilfe in Feld und Labor.

---

Allen Ex- und Mit-Doktorand(inn)en (Anita, Beni, Kathi, Kim, Juna, Tine) für Fachdiskussionen und (fast noch wichtiger) zerstreuende Stunden beim Bier.

Zu guter Letzt gehört ein besonderer Dank meiner Freundin Sandra für ihre Geduld und meinen Eltern, die es mir ermöglicht haben in schon leicht angegrautem Zustand überhaupt ein Studium absolvieren zu können.

---

## CURRICULUM VITAE

### David Hiltbrunner

Born 24<sup>th</sup> of June, 1975, Switzerland

### Scientific education

03/2009 – 10/2012	<b>Swiss Federal Research Institute WSL, University of Zurich</b> PhD: Turnover and stabilization of soil organic matter: effect of land-use change in alpine regions Supervisors: Dr. Stephan Zimmermann Dr. Frank Hagedorn Prof. Dr. Michael W. I. Schmidt
09/2002 – 09/2008	<b>University of Zurich</b> Master degree: Geography Subsidiary subj.: Geology, Chemistry Diploma thesis: Analysis of microbial phospholipids

### Publications

2012	<b>Hiltbrunner D</b> , Schulze S, Hagedorn F, Schmidt MWI, Zimmermann S (2012) Cattle trampling alters soil properties and changes soil microbial communities in a Swiss sub-alpine pasture. <i>Geoderma</i> , <b>170</b> , 369-377.  <b>Hiltbrunner D</b> , Zimmermann S, Hagedorn F, Niklaus PA (2012) Increasing soil methane sink along a 120-year afforestation chronosequence is driven by soil moisture. <i>Global Change Biology</i> <i>Accepted</i>  <b>Hiltbrunner D</b> , Hagedorn F, MWI Schmidt, Zimmermann S, (2012) Moderate effects of reforestation with Norway spruce ( <i>Picea abies</i> ) on carbon storage and turnover in a Swiss sub-alpine pasture (running title) <i>In preparation</i>
------	----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

## Conferences

- 01/2010      **COST Action FP0803 Conference, Birmensdorf**  
*Poster:*  
**David Hiltbrunner**, F. Hagedorn, A. Miltner  
Soil microbial communities in a CO<sub>2</sub>-enriched and <sup>13</sup>C-labelled treeline ecosystem with different tree species
- 04/2010      **European Geoscience Union (EGU), Vienna**  
*Talk:*  
**David Hiltbrunner**, F. Hagedorn, A. Miltner  
Soil microbial communities in a CO<sub>2</sub>-enriched and <sup>13</sup>C-labelled treeline ecosystem
- 05/2011      **European Geoscience Union (EGU), Vienna**  
*Poster 1:*  
**David Hiltbrunner**, P.A. Niklaus, S. Schulze, M.W.I. Schmidt, S. Zimmermann: Cattle trampling alters soil properties and greenhouse gas fluxes from soils in a Swiss sub-alpine pasture  
*Poster 2:*  
**David Hiltbrunner**, M.W.I. Schmidt, S. Zimmermann: Afforestation of former pastures with *Picea abies* in a subalpine ecosystem affects the microbial community composition
- 09/2011      **Deutsche Bodenkundliche Gesellschaft (DBG), Berlin**  
*Poster:*  
**David Hiltbrunner**, F. Hagedorn, P.A. Niklaus, M.W.I. Schmidt, S. Zimmermann: Beweidung verändert die Bodeneigenschaften und die Gasflüsse in einer voralpinen Weide der Schweiz
- 04/2012      **European Geoscience Union (EGU), Vienna**  
*Talk:*  
**David Hiltbrunner**, F. Hagedorn, P.A. Niklaus, M.W.I. Schmidt, S. Zimmermann: Moderate effects of reforestation with Norway spruce (*Picea abies*) on carbon storage and turnover in a Swiss sub-alpine pasture